## 2013; 17: 1211-1217

# Oxidative stress and antioxidant defense in Egyptian favism patients

H.G. OSMAN, F.M. ZAHRAN\*, A.M. A. EL-SOKKARY, A.M. SABRY

Biochemistry Division, Chemistry Department, Faculty of Science, Mansoura University, Mansoura, Egypt \*Biochemistry Division, Chemistry Department, Faculty of Science, Zagazig University, Zagazig, Egypt

**Abstract.** - BACKGROUND: Favism occurs as the result of intolerance to the ingesting of fava beans or to the inhalation of pollen from the *Vicia faba* plant. Patients with favism are always Glucose-6-phosphate dehydrogenase (G6PD)-deficient, but not all G6PD-deficient individuals develop hemolysis as a result of fava beans consumption.

**PATIENTS AND METHODS:** Blood samples were collected from children with favism (n = 55) between age (2-12 years) on EDTA tubes divided into 3 groups: group 1 control group (n = 15), group 2 before blood transfusion (during hemolytic action) (n = 20) and group 3 after blood transfusion (treated) (n = 20).

**RESULTS:** It was found that in group 2 GSH level was significantly low;  $(1.11 \pm 0.39, p < 0.001)$  compared to controls  $(26.31 \pm 5.26, p < 0.001)$ . In group 3 after blood transfusion Level of GSH rose but remained lower than normal level  $(5.88 \pm 2.33, p < 0.001)$  compared to controls. As for oxidative stress parameters, both levels of  $H_2O_2$  and MDA were highly significant in group 2;  $(213.49 \pm 57.56, p < 0.001)$ ,  $(98.05 \pm 22.34, p < 0.001)$  compared to controls  $(3.75 \pm 1.164, p < 0.001)$ ,  $(7.38 \pm 2.07, p < 0.001)$ , respectively.

Moreover, in group 3 after blood transfusion, levels of  $\rm H_2O_2$  and MDA were decreased but remained high compared to controls (66.55  $\pm$  22.49, p < 0.001), (47.18  $\pm$  9.62, p < 0.001) sequentially. Also, there was a negative correlation between GSH that acts as antioxidant defense enzyme and each one of oxidative stress parameters MDA &  $\rm H_2O_2$ . However, there was a positive correlation between  $\rm H_2O_2$  and MDA.

**CONCLUSIONS:** From this study, it could be concluded that the favic patients have high oxidative stress ( $H_2O_2$  and MDA) more than normal individuals and less antioxidant defense (GSH). With the passage of time these individuals, cells would be more vulnerable for  $H_2O_2$ -induced senescence.

Kev words:

Favisim, Oxidative stress, Antioxidant defense.

## Introduction

Exposure to fava beans (*Vicia faba*; broad bean) is known to be toxic and potentially fatal for some individuals since the era of the old Greeks. Favism is the acute hemolysis that follows the ingestion of fava beans. This syndrome appears to be limited to those who have the Mediterranean variant with more prevalence among males than females. There is an increased frequency between children who are between the ages of 2 and 6 years. Also, the breast-fed infant whose mother had ingested fava beans can develop favism<sup>1</sup>.

Favic patients are always G6PD deficient, but not all G6PD-deficient individuals show symptoms of hemolysis when they ingest fava beans. Therefore, G6PD deficiency is essential but not enough to cause favism. Favism is the most common in patients who develop G6PD class II variants, but it is uncommon in patients with the G6PD class V A-variant<sup>2</sup>. Fava beans are assumed to be the cause of oxidative damage produced by an unknown component, probably vicine, convicine, or isouramil<sup>3,4</sup>.

G6PD deficiency is the most common existing enzymatic disorder of red blood cells in human beings. About 400 million people are considered to be affected by this deficiency<sup>5</sup>. The G6PD enzyme is involved in catalyzing the first step in the pentose phosphate pathway (PPP), which leads to the production of antioxidants that protect cells against oxidative damage<sup>6</sup>. This pathway shows the production of NADPH, which maintains the reduced glutathione within the cell. Reduced glutathione acts as an antioxidant and protects cells against oxidative damage<sup>6</sup>. A G6PD-deficient patient, consequently, is not capable of protecting red blood cells against oxidative stresses from certain drugs, metabolic conditions, infections, and consumption of fava beans<sup>7</sup>.

G6PD is reported to be more common in Africa, southern Europe, the Middle East, South-East Asia, and the central and Southern Pacific islands. However, deficient alleles are quite widespread in North and South America as well as in some areas of northern Europe due to migration<sup>8</sup>. G6PD deficiency is an X-linked hereditary genetic defect produced by mutations in the G6PD gene. The inheritance of G6PD deficiency, a typical X-linked, exhibits pattern with higher frequency in males than in females<sup>7</sup>.

In most cells, the generation of the necessary intracellular NADPH has been backed up by other metabolic pathways. In contrast, red blood cells do not have other options to produce NADPH. Thus, G6PD deficiency becomes fatal in red blood cells, where hemolytic anemia will be the outcome of any oxidative stress. Numerous factors may be the cause of oxidative stress including the ingesting of fava beans, certain drugs, infections, and certain metabolic conditions such as diabetic ketoacidosis<sup>6</sup>.

NADPH has a key function in the reduction of oxidized glutathione (GSSG) to a tripeptide known as reduced glutathione (GSH). This tripeptide acts as a reducing agent together with the enzyme glutathione peroxidase, which plays an important role in the detoxification of hydrogen peroxide. During this process, GSH is converted into GSSG which in turn decreases the GSH levels. Glutathione reductase catalyzes the reduction of GSSG to GSH in the presence of NADPH leading to the regeneration of GSH9. Red blood cells have no other sources of NADPH, i.e. G6PD is required for the protection of hemoglobin sulfhydryl groups and preventing red blood cell membrane oxidation<sup>10</sup>. Lipid peroxidation in red blood cells results in disorganization of the lipid moiety of cell membranes causing in lethal damage to the cell<sup>11</sup>.

The aim of our study is to find out the correlation between oxidative stress (H<sub>2</sub>O<sub>2</sub> and MDA) and antioxidant defense (GSH) in favism patients.

#### **Patients and Methods**

## **Patients**

This study was conducted on 55 children dividing to 3 groups:

Group 1, [control group, (n = 15)], group 2, (before blood transfusion (during hemolytic action) (n = 20) and group 3, after blood transfusion (treated) (n = 20). They were enrolled from those presented and followed up at Pediatric

Hospitals in both Mansoura and Zagazig University Children's Hospital and after taking a written formal consent from the patient's parents.

#### Methods

G6PD assay

All patients were screened for G6PD deficiency using qualitative visual method from the red cell hemolysate (Span Diagnostics Ltd, Surat, India)<sup>(12,13)</sup>.

## Estimation of glutathione reduced (GSH)

The concentration of GSH in RBC<sub>s</sub> was assessed by the method based on the reduction of 5,5'-dithiobis (2-nitrobenzoic acid) (DTNB) with glutathione (GSH) to produce a yellow compound. The reduced chromogen directly proportional to GSH concentration and its absorbance can be measured at 405 nm<sup>14</sup>.

## Estimation of hydrogen peroxide $(H_2O_2)$

The concentration of  $H_2O_2$  in plasma was assessed by the method based on the reaction of  $H_2O_2$  with 3, 5'dichloro-2-hydroxy benzene sulfonic acid (DHBS) and 4- aminophenazone (AAP) in the presence of peroxidase (HRP) to form achromophore<sup>15,16</sup>.

## Estimation of Malondialdehyde (MDA)

The lipid peroxidation products were estimated by the formation of thiobarbituric acid (TBA) and quantified in term of MDA, where TBA reacts with MDA in acidic medium at temperature of 95°C for 30 min. to form thiobarbituric acid reactive product, the absorbance of the resultant pink product can be measured at 534 nm<sup>17,18</sup>.

### Statistical analyses

Statistical differences between groups were evaluated using Student's t-test. Pearson-product correlation was used to test for linear relationship between different variables. The software SPSS 11.0 for Windows (SPSS Inc., Chicago, IL, USA) was used for statistical analyses. Results were reported as means  $\pm$  SD, and a p value < 0.05 was considered statistically significant.

### Results

It was found that in group 2 GSH level was significantly low;  $(1.11 \pm 0.39, p < 0.001)$  com-

<b>Table I.</b> Minimum, Maximum, and Standard Deviation values of GSH, H <sub>2</sub> O <sub>2</sub> and MDA obtain	ed from groups 1, 2 and 3.
--	----------------------------

Group	Variable	No.	Min.	Max.	Mean	St. deviation
Control	GSH	15	19.89	35.00	26.3086*	5.25638
	$H_2O_2$	15	2.39	5.75	3.7497*	1.16428
	MDA	15	3.66	11.19	7.3831*	2.74692
Hemolytic action	GSH	20	0.42	1.98	1.1065*	0.38544
•	$H_2O_2$	20	104.50	300.00	213.4863*	57.55669
	MDA	20	65.60	153.80	98.0538*	22.33509
Treated	GSH	20	2.60	10.13	5.8806*	2.33060
	$H_2O_2$	20	38.36	100.20	66.5498*	22.49079
	MDA	20	32.35	60.50	47.1839*	9.62414

The mean difference is significant at 0.001 levels (p < 0.001).

pared to controls (26.31  $\pm$  5.26, p < 0.001). In group 3 after blood transfusion Level of GSH rose but remained lower than normal level (5.88  $\pm$  2.33, p < 0.001) compared to controls. As for oxidative stress parameters, both levels of H<sub>2</sub>O<sub>2</sub> and MDA were highly significant in group 2; (213.49  $\pm$  57.56, p < 0.001), (98.05  $\pm$  22.34, p < 0.001) compared to controls (3.75  $\pm$  1.164, p < 0.001), (7.38  $\pm$  2.07, p < 0.001), respectively.

Moreover, in group 3 after blood transfusion, levels of  $\rm H_2O_2$  and MDA were decreased but remained high compared to controls (66.55  $\pm$  22.49, p < 0.001), (47.18  $\pm$  9.62, p < 0.001) sequentially (Table I).

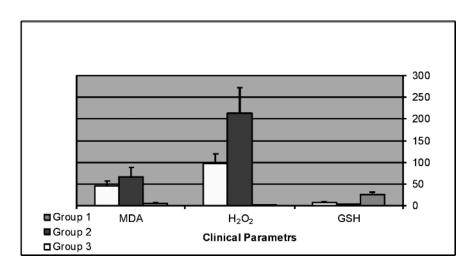
This high significant difference between studied groups was shown in (Figure 1).

Also, there was a negative correlation between GSH that acts as antioxidant defense enzyme and each one of the oxidative stress parameters MDA

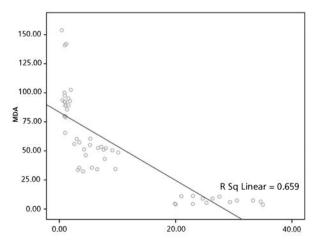
&  $H_2O_2$  (Figures 2 and 3). However, there was a positive correlation between  $H_2O_2$  and MDA (Figure 4). These correlations were significant at the (0.01 and 0.05 level) as shown in (Table II).

## Discussion

Oxidative stress results from a disturbance in the balance between the production of Reactive Oxygen Species (ROS) and the antioxidant defense efficiency. Furthermore, oxidative stress occurs if excessive production of ROS overcomes the antioxidant defense system or when there is a significant decrease of antioxidant defense<sup>19</sup>. Free radicals attack potential biological targets such as lipids, proteins and nucleic acids<sup>20</sup>. The epoxides are formed as the result of an increase in oxidative stress. These epoxides



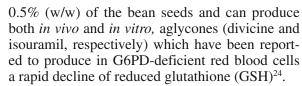
**Figure 1.** MDA, H<sub>2</sub>O<sub>2</sub> and GSH levels in studied groups.



**Figure 2.** The Pearson negative correlation between GSH and MDA.

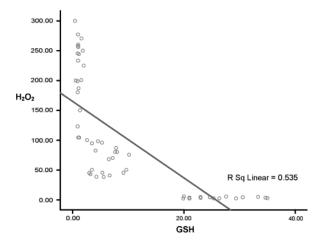
may spontaneously react with the nucleophilic centers present in the cell and, thereby, covalently bind to DNA, RNA and protein. These reactions leading to carcinogenicity and cytotoxicity are dependent on the properties of the epoxides<sup>21</sup>.

This present study showed that the patients with favism had significantly low level of Reduced Glutathione (GSH),  $(26.31 \pm 5.26, p < 0.001)$  in comparison to those of healthy controls  $(26.31 \pm 5.26, p < 0.001)$ , (Figure 1). These findings are consistent with the results obtained by some investigators all over the world<sup>22,23</sup>, where it has been established that, the glycosides vicine and convicine play a role in generating the damage and ultimate lysis of the affected red cells. Actually, the latter substances represent about the

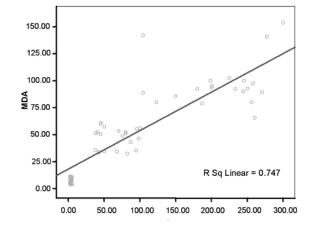


As G6PD is crucial in maintaining redox balance and detoxification of ROS, it is possible that G6PD deficiency retards the antioxidant defense, leading to oxidative damage and, therefore, cellular senescence<sup>24</sup>. Moreover, G6PD-deficient cells display increased tendency for H<sub>2</sub>O<sub>2</sub>-induced senescence<sup>25</sup>. Consequently, favism patients in the present study had high significant levels of  $H_2O_2$  (213.49 ± 57.56, p < 0.001), (Table I), even after blood transfusion (66.55  $\pm$  22.49, p < 0.001) when compared to controls (3.75  $\pm$  1.164, p <0.001), (Figure 1), which may lead to conclude that these favism patients are more vulnerable for H<sub>2</sub>O<sub>2</sub>-induced senescence. In other in vitro studies it was reported that high levels of H<sub>2</sub>O<sub>2</sub> cause damage to lipids, proteins, mitochondrial DNA and genomic DNA in a relatively undiscriminating manner. The buildup of this damage cripples the ability of cells to grow, provoking senescence<sup>26</sup>.

Furthermore, severe oxidative stress is not only known to be the reason for DNA damage and mutations of tumor suppressor genes, which are known to be the initial events in carcinogenesis <sup>19</sup>, but can also play a key role in the promotion of multistep carcinogenesis<sup>27</sup>. Lipids, especially polyunsaturated fatty acids (PUFA), are very vulnerable to free radical attack, which can result in lipid peroxidation<sup>28</sup>. Lipid peroxidation plays a vital role in the control of cell division<sup>29</sup>. Malon-



**Figure 3.** The Pearson negative correlation between GSH and  $H_2O_2$ .



**Figure 4.** The Pearson positive correlation between MDA and  $H_2O_2$ .

Table II. Correlations between studied groups

		group	GSH	H <sub>2</sub> O <sub>2</sub>	MDA
group	Pearson Correlation	1	- 693(**)	.185	.329(*)
	Sig. (2-tailed)		.000	.177	.014
	N	55	55	55	55
GSH	Pearson	-	1	-	-
	Correlation	.693(**)		.731(**)	.812(**)
	Sig. (2-tailed)	.000		.000	.000
	N	55	55	55	55
$H_2O_2$	Pearson		-		
	Correlation	.185	.731(**)	1	.865(**)
	Sig. (2-tailed)	.177	.000		.000
	N	55	55	55	55
MDA	Pearson		-		
	Correlation	.329(*)	.812(**)	.865(**)	1
	Sig. (2-tailed)	.014	.000	.000	
	N	55	55	55	55

<sup>\*\*</sup> Pearson Correlation is significant at the 0.01 level (2-tailed). \*Pearson Correlation is significant at the 0.05 level (2-tailed).

dialdehyde (MDA); the end product of lipid peroxidation, is suggested to act as a tumor promoter and a cocarcinogenic agent due to its high cytotoxicity and inhibitory action on protective enzymes<sup>30</sup>. In our study, it was found that the favic patients had significant high levels of MDA  $(98.05 \pm 22.34, p < 0.001)$ , even after blood transfusion (47.18  $\pm$  9.62, p < 0.001), when compared to controls (7.38  $\pm$  2.07, p < 0.001) (Figure 1), thus the cell growth of these patients will be affected with passage of time. These results are in accordance with the reports which have referred to fatty acid composition of red cells deficient in G6PD and their susceptibility to lipid peroxidation<sup>30</sup>. In addition, these reports have demonstrated that the red cell sensitivity to lipid per oxidation was found to be higher in patients with glucose-6- phosphate dehydrogenase deficiency than in normal subjects<sup>31</sup>. This may indicate the presence of increased oxidative stress. Increase in MDA levels could be caused by a rise in the generation of reactive oxygen species (ROS) due to the excessive oxidative damage generated in these patients produced from fava beans components divicine and isouramil.

In our study, it was also found that there was a negative correlation between GSH and MDA (Figure 2), in other reports it was stated that GSH is necessary for the activity of glutathione peroxidase (GPx) which efficiently protects the cell

membrane from lipid peroxidation and catalyzes the reaction of hydro peroxides with GSH to form GSSG<sup>32</sup>.

Moreover, it was found that there was another negative correlation between GSH and H<sub>2</sub>O<sub>2</sub> (Figure 3), this finding is consistent with other in vitro reports which described that the decrease of GSH in RBCs of aged rats could be the reason for the inhibited activities of GPx and glutathione S transferase (GST), since these enzymes act only in the expense of GSH<sup>32</sup>, as glutathione is a reducing agent because of its sulphydryl groups against oxidative stress, depletion of glutathione causes a proportional decrease of GPx activity. Reduced glutathione (GSH) is the main non-enzymatic antioxidant defense within the cell, reducing different peroxides, hydroperoxides and radicals (alkyl, alkoxyl, peroxyl, etc.)<sup>33</sup>. Thus, it is usually assumed that GSH depletion reflects intracellular oxidation.

It has been reported that divicine and isouramil react with oxygen to produce  $H_2O_2$ , and this reaction can be maintained as a redox cycle in the presence of reduced glutathione, which regenerates the reduced, autoxidizable pyrimidine form<sup>34</sup>. This mechanism can be responsible for the observed effect on sensitive red blood cells.

In other works it was found that, an acute hemolytic anemia can occur in some dehydrogenase-deficient subjects as a result of administration of exogenous substances other than fava beans<sup>35</sup>, particularly hemolytic drugs<sup>36</sup>. These compounds are likely to interact with oxygen, or, in the red cell, with oxyhemoglobin, and eventually produce  $H_2O_2$ ,  $via\ O_2^-$ , as intermediate<sup>37,38</sup>.

The present study showed a positive correlation between  $H_2O_2$  and MDA (Figure 4), where as  $H_2O_2$  level increased MDA would be increased consequently. Other investigators have reported, that when RBCs are incubated with high concentration of  $H_2O_2$  in the presence of a catalase inhibitor (Sodium Azide), membrane lipids are slowly and partly oxidized<sup>39</sup>. The result of that lipid peroxidation usually MDA, which is one of thiobarbituric acid (TBA) reactive species<sup>30</sup>.

#### Conclusions

Favic patients had increased oxidative stress parameters, (H<sub>2</sub>O<sub>2</sub> and MDA) more than normal individuals and less antioxidant defense enzyme (GSH). With the passage of time, these individuals cells would be more susceptible to H<sub>2</sub>O<sub>2</sub> induced senescence. The implication of oxidative stress in the etiology of several chronic and acute degenerative disorders suggests that antioxidant therapy represents a promising avenue for the treatment. Hence, it is recommended for these patients to take dietary antioxidant factors from food and medicinal plants.

Therefore, according to the significant correlations obtained between these parameters and favism, it is recommended that these parameters may be used as a biochemical marker for this disease.

#### Acknowledgements

The Authors would like to thank all the medical staff of Pediatric Hospitals in both Mansoura and Zagazig University Children's Hospital for providing help and assistance during the collection of samples.

#### **Conflict of Interest**

None declared.

## References

 NOORI-DALOII MR, DANESHPAJOOH M. Molecular Basis of G6PD Deficiency: Current Status and its Perspective. Acta Med Iran 2008; 46: 167-182.

- 2) JALLOH A, TANTULAR IS, PUSARAWATI S, KAWILARANG AP, KERONG H, LIN K, FERREIRA MU, MATSUOKA H, ARAI M, KITA K, KAWAMOTO F. Rapid epidemiologic assessment of glucose-6-phosphate dehydrogenase deficiency in malaria-endemic areas in Southeast Asia using a novel diagnostic kit. Trop Med Int Health 2004; 9: 615-623.
- 3) GREGG XT, PRCHAL JT. RED CELL ENZYMOPATHIES. IN: HOFFMAN R, ED. Hematology: basic principles and practice. 4th ed. Philadelphia: Churchill Livingstone 2000: pp. 57-60.
- OBLENDER M. Index of suspicion. Case 3. Kernicterus in a baby girl homozygous for glucose-6-phosphate dehydrogenase deficiency. Pediatr Rev 1993; 14:191- 193.
- 5) GLADER BE. Glucose-6-phosphate dehydrogenase deficiency and related disorders of hexose monophosphate shunt and glutathione metabolism. In:Wintrobe's Clinical Hematology. 10th ed. Baltimore: Williams & Wilkins 2008; pp. 1176-1190.
- 6) LUZZATTO L, METHA A, VULLIANYT. Glucose-6phosphate dehydrogenase deficiency. In: Scriver CR, Beaudet AL, Sly WS, et al, eds:The Metabolic and Molecular Basis of Inherited Disease.8th ed.Columbus: McGraw-Hill 2001: pp.4517-4553.
- CAPPELLINI MD, FIORELLI G. Glucose-6-phosphate dehydrogenase deficiency. Lancet 2008; 371:64-74.
- WORLD HEALTH ORGANIZATION. Working group glucose-6- phosphate dehydrogenase deficiency. Bull WHO 1989; 67: 601-611.
- NJÁLSSON R, NORGREN S. Physiological and pathological aspects of GSH metabolism. Acta Paediatr 2005; 94: 132-137.
- 10) PANDOLFI PP, SONATI F, RIMI R, MASON P, GROSVELD F, LUZZATTO L. Targeted disruption of the housekeeping gene encoding glucose 6-phosphate dehydrogenase (G6PD): G6PD is dispensable for pentose synthesis but essential for defense against oxidative stress. EMBO J 1995; 14: 5209-5215.
- 11) MIHARA M, UCHIYAMA M. Determination of malonaldehyde precursor in tissues by thiobarbituric acid test. Anal Biochem 1978; 86: 271-278.
- ELLIS HA, KIRKMAN HN. A colorimetric method for assay of erythrocytic glucose-6-phosphate dehydrogenase. Proc Soc Exp Biol Med 1961; 106: 607-609.
- 13) UBIN BH, OSKI FA, BRIGANDI PE. An evaluatin of screening procedures for red cell glucose-6-phosphate dehydrogenase deficiency in the newborn infant. J Pediatr 1967; 70: 788-792.
- 14) BEUTLER E, DURON O, KELLY MB. Improved method for the determination of blood glutathione. J Lab Clin Med 1963; 61: 882-888.
- AEBI H. Catalase in vitro. Methods Enzymol 1984; 105: 121-126.
- 16) FOSSATI P, PRENCIPE L, BERTI G. Use of 3, 5-dichloro-2-hydroxybenzenesulforic acid/4-aminophenazone chromogenic system in direct enzymic assay of uric acid in serum and urine. Clin Chem 1980; 26: 227-231.

- SATOH K. Serum lipid peroxide in cerebrovascular disorders determined by a new colorimetric method. Clin Chem Acta 1978; 90: 37-43.
- OHKAWA H, OHISHI W, YAGI K. Assay of lipid peroxidation in animal tissue by thiobarbutyric acid reaction. Anal Biochem 1979; 95: 351-358.
- 19) Kang DH. Oxidative stress, DNA damage and breast cancer. AACN Clin Issues 2002; 13: 540-549.
- PRYOR WA. Cancer and free radicals. Basic Life Sci 1986; 39: 45-59.
- 21) TAMPO Y, TSUKAMOTO M. The antioxidant action of 2-methyl-6-(pmethoxyphenyl-3,7-dihydroimidazo[1,2-alpha]pyrazin-3-one (MCLA), a chemiluminescence probe to detect superoxide anions. FEBS Lett 1998; 430: 348-352.
- SALEN G, GOLDSTEIN F, HAURAN FC, WIRTS W. Acute hemolytic anemia complicating viral hepatitis in patients with glucose-6-phosphate dehydrogenase deficiency. Ann intern Med 1966; 65: 1210-1220.
- KATTAMIS AC, TJARTJATOU F. The hemolytic process of Viral hepatitis in children with normal or G6PD activity. J Pediatr 1970; 77: 422-430.
- 24) Mager J, Glaser G, Razin A, Izak G, Bien S, Noam M. Metabolic effects of pyrimidines derived from fava bean glycosides on human erythrocytes deficient in glucose-6-phosphate dehydrogenase. Biochem Biophys Res Commun 1965; 20: 235-240.
- CHENG ML, HO HY, WU YH, CHIU DT. Glucose-6phosphate dehydrogenase-deficient cells show an increased propensity for oxidant-induced senescence. Free Radic Biol Med 2004; 36: 580-591.
- 26) ORR WC, MOCKETT RJ, BENES JJ, SOHAL RS. Effects of over expression of copper-zinc and manganese superoxide dismutases, catalase, and thioredoxin reductase genes on longevity in Drosophila melanogaster. J Biol Chem 2003; 278: 26418-26422.
- AHMED MI, FAYED ST, HOSSEIN H, TASH FM. Lipid peroxidation and antioxidant status in human cervical carcinoma. Dis Markers 1999; 15: 283-291.
- HALLIWELL B, GUTTERIDGE MC. Free Radicals in Biology and Medicine. 3rd ed. London: Oxford, 1999.

- DIPLOCK AT, RICE-EVANS AC, BURTON RH. Is there a significant role for lipid peroxidation in the causation of malignancy and for antioxidants in cancer prevention? Cancer Res 1994; 54: 1952-1956.
- OTAMIRI T, SJODAHL R. Increased lipid peroxidation in malignant tissues of patients with colorectal cancer. Cancer 1989; 61: 122-125.
- CLEMENS MR, EINSELE H, WALLER HD. The fatty acid composition of red cells deficient in glucose-6phosphate dehydrogenase and their susceptibility to lipid peroxidation. Klin Wochenschr 1985; 63: 578-582.
- 32) SPECTOR A, WANG GM, WANG RR, GARNER WH, MOLL H. The prevention of cataract caused by oxidative stress in cultured rat lenses by H<sub>2</sub>O<sub>2</sub> and photochemically induced cataract. Curr Eye Res 1993; 12: 163-179.
- VIÑA J. Glutathione: metabolism and physiological functions. CRC Press, Boston 1990; pp. 222-228.
- 34) CHEVION M, NEVOK T, GLASER G, MAGER J. The chemistry of favism-inducing compounds: the properties of isouramil and divicine and their reaction with glutathione. Eur J Biorhem 1982; 127: 405-409.
- 35) BEUTLER E. IN: STANBURY JB, WYNGAARDEN JB, FREDRICKSON DS, GOLDSTEIN JI, BROWN MS, eds, The metabolic basis of inherited disease. McGraw Hill, New York 1983: pp. 1629-1653.
- CARREL RW, WINTERBOURN CC, RACHMIKWITZ EA. Activated oxygen and hemolysis. Br J Haematol 1975; 30: 259-264.
- 37) GOLDBERG B, STERN A, PEISACH J. The mechanism of superoxide anion generation by the interaction of phenylhydrazine with hemoglobin. J Biol Chem 1976; 251: 3045-3051.
- 38) GOLDBERG B, STERN A, PEISACH J, BLUMBERG WE. The detection of superoxide anion from the reaction of oxyhemoglobin and phenylhydrazine using EPR spectroscopy. Experientia 1979; 35: 488-489.
- STOCKS J, DORMANY TL. A direct thiobarbituric-acid reaction chromogen in human red blood cells. Clin Chim Acta 1970; 27: 117-120.