The anticancer mechanism of caffeic acid phenethyl ester (CAPE): review of melanomas, lung and prostate cancers

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Abstract. – BACKGROUND: Caffeic acid phenethyl ester (CAPE), an active component of propolis extract, specifically inhibits NF-κB. It exhibits antioxidant, antiinflammatory, antiproliferative, cytostatic, and most improtantly, antineoplastic properties.

AIM: The aim of the present mini-review is to summarize and evaluate the anticancer mechanism of CAPE with examples of several cancer types.

RESULTS: In view of the mechanisms and findings in our laboratory and those of others in literature, we suggest that CAPE possess anticancer and apoptosis inducing activities.

CONCLUSIONS: Further researches are needed regarding the anticancer basis of CAPE in all disciplines of medicine. Also, clinical potential toxicities of CAPE should be revealed if it is going to be used as an anticancer agent.

Key Words:

Caffeic acid phenethyl ester (CAPE), Cancer, Effect, Treatment

Introduction

As a natural compound bearing polyphenolic ring, CAPE (Figure 1), is originated from a biologically active bee product, propolis. It is known to have antitumoral, antiinflammatory, antineoplastic and antioxidant properties to some extend. It is a white-color powder as a commercial product with a storage temperature -20° C and soluble in ethanol, DMSO, and ethyl acetate (50 mg/ml). Its empirical formula is $C_{17}H_{16}O_4$ and has 284.31 g/mol molecular weight (Figure 1).

CAPE is a specific inhibitor of NF-κB. It has been shown to significantly suppress the lipoxygenase pathway of arachidonic acid metabolism

during inflammation in μM concentrations. At a concentration of 10 μM , it completely blocks production of reactive oxygen species (ROS) in human neutrophils and the xanthine/xanthine oxidase system.

Possible Anticancer Mechanisms of CAPE

It has been suggested that CAPE potently stimulate glucose uptake in cultured skeletal muscle cells throughout the adenosine monophosphate-activated kinase-protein (AMKP) pathways showing an important anti-diabetic potential¹. CAPE has a higher hydrophobicity and stronger inhibition potency toward xanthine oxidase (XO) and it inhibits the enzymatic activity via binding to the molybdoptrein region of its active site². Because XO has an action to metabolyze both purine and pyrimidine bases, by this inhibition mechanism, CAPE can stop the nucleotide turnover salvage pathway showing anticancer activity in all cell types. It is suggested to be used in the treatment of gout and hyperuricemia because of that XO inhibitory effect³.

Oxidative stress is also suggested to be a major cause of cellular injuries in carcinogenesis. It was tested by using erythrocyte membrane ghost lipid peroxidation, plasmid pBR322 DNA, and protein damage initiated by the water-soluble initiator 2,2'-azabis(2-amidinopropane) hydrochloride and H₂O₂ monitored by formation of hydroperoxides and by DNA nicking assay, single-cell alkaline electrophoresis, and sodium dodecyl sulphate (SDS)-polyacrylamide gel (PAGE) electrophoresis. The results showed that CAPE and its related polyphenolic acid esters elicited remarkable inhibitory effects on erythrocyte membrane lipid peroxidation, cellular DNA strand breakage, and protein fragmentation⁴.

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Figure 1. The chemical structure of caffeic acid phenethyl ester.

Arachidonic acid metabolites are important mediators in cancer and inflammation processes, so when we look at the lipoxygenase inhibitor effect of CAPE, we can see that CAPE has a ability to inhibit 5-lipoxygenase by a complete uncompetitive mechanism⁵. Inhibition of the tumor promoter-mediated oxidative processes by CAPE has also been reported in the culture of HeLa cells⁶. The other studies has also shown that CAPE and the other caffeic acid esters inhibit azoxymethane-induced colonic preneoplastic lesions and enzyme activities, including ornithine decarboxylase, tyrosine kinase and lipoxygenase, associated with colon carcinogenesis⁷.

One of the anti-tumor promotion activities of CAPE is suggested to be the induction of apoptosis. CAPE suppressed 12-O-tetradecanylphorbol-13-acetate-induced cell transformation and induced apoptosis in mouse epidermal JB6 Cl 41 cell. No difference in induction of apoptosis was observed between nomal lymphoblasts and sphingomyelinase-deficient cell lines. CAPE treatment of two p53 mutant tumor cell lines, NCI-H358 and SK-OV-3, and p53 deficient (p53(-/-)) and Cl 41 cells caused the cleavage of caspase-3 as well as DNA fragmentation. However, caspase-3 cleavage was seen early only in cell expressing wild-type p53 (p53(+/+)) and Cl 41 cells. This shows p53 may be involved in the early stage of CAPE-induced apoptosis. In addition, CAPE may induce apoptosis through p53dependent and -independent pathways and its anti-tumor promotion activity may have occured through the induction of apoptosis⁸. To reveal the mechanism of CAPE-induced differential cytotoxicity, nontumorigenic cloned rat embryo fibroblasts (CREF) and adenovirus-transformed CREF cells (Wt3A) were used in a study. Nucleosomal-length DNA degradation, morphological alterations by electron microscopy, in situ labeling of 3'OH ends, and the appearance of a hy-

podiploid cell population by bivariant flow cytometry, cell death induced by CAPE in the transformed Wt3A cells revealed that the mechanism is apoptosis. CAPE has this effect by modulating the redox state (as GSH level) of the cells⁹. A series of oncogene transformed, oncogene-reverted and CAPE-resistant oncogene transformed CREF cells were used to observe the mechanism underlying the increased sensitivity of transformed cells to CAPE. A direct relationship exists between the cytotoxic effects of CAPE and the induction of DNA fragmentation and apoptosis. The expression of the transformed phenotype by rodent cells evokes sensitivity to CAPE-induced toxicity through apoptosis¹⁰. Evidence indicated that CAPE may represent a unique compound that can specifically target progressed transformed cells for growth suppression and toxicity. An understanding of the mechanism underlying this selective effect of CAPE could result in the identification of important biochemical pathways mediating cellular transformation and progression of the transformed state¹¹.

Angiogenesis is a fundamental pathogenic process in cancer and it is very valuable to find some compounds that has potential inhibitory effect on angiogenesis. One of these compounds is the biomimetic dimerization product of CAPE, benzo[k,l]xanthene lignan. The lignan showed a significant, dose-related inhibitory effect on new vessel growth in the angiogenesis bioassay and it inhibited vascular endothelial growth factor (VEGF) secretion in ovarian cell culture¹². In an in vitro tube formation assay, human umbilical vein endothelial cells and fibroblast cells were incubated for 14 days with VEGF for induction of proliferation and migration of cells and with potential inhibitor, CAPE. It significantly suppressed VEGF-induced in vitro tube formation and proliferation¹³.

The effect of CAPE on tumor invasion and metastasis by determining the regulation of matrix metalloproteinases (MMPs), which are zinc-dependent proteolytic enzymes playing pivotal role in tumor metastasis by cleavage of extracellular matrix as well as nonmatrix substrates, is an attractive area. Dose dependent decreases in MMP and tissue inhibitor of MMP-2 mRNA levels were observed in CAPE-treated HT1080 human fibrosarcoma cells as detected by reverse transcriptase-polymerase chain reaction (RT-PCR). Gelatin zymography analysis also exhibited a significant down-regulation of MMP-2 and MMP-9 expression in HT1080 cells trated with

CAPE. Moreover, CAPE inhibited the activated MMP-2 activity as well as invasion, motility, cell migration and colony formation of tumor cells¹⁴. One of cell culture cytotoxicity assays of CAPE in colon adenocarcinomas showed a dose-dependent decrease in cell viability, cell invasion by 47.8%, and expression of MMP-2 and -9. The other effects were inhibition of VEGF production, inhibition of pulmonary metastatic capacity accompanied with a decreased plasma VEGF (in BALB/c mice), and also prolonged survival of mice implanted by colon adenocarcinomas¹⁵.

In view of the above-mentioned mechanisms and findings in our laboratory and those of others in literature, we suggest that CAPE possess anticancer and apoptosis inducing activities. Further researches are needed regarding the clinical potential toxicities of CAPE if it is going to be used as an anticancer agent.

The effect of CAPE on Several Cancer Types in Experimental and Cell Culture Models

Lung Cancers

Lots of chemopreventive agents have been reported to interfere with the intracellular signal that is related to carcinogenesis, the proliferation of cancer cells, apoptosis, and cell migration. The effects of the representative chemopreventive agents on the transforming growth factor-beta (TGF-β)-induced invasive phenotype using A549 lung adenocarcinoma cells as a model system was investigated by using CAPE as therapeutic agent. CAPE effectively suppressed TGFβ-enhanced cell motility and TGF-β-induced Akt (protein kinase β) activation as well as a specific inhibitor of phosphatidyl inositol 3-kinase (PI3K)/Akt pathway, LY294002¹⁶. By this finding, the Authors suggest that CAPE can be applied not only as chemopreventive agent but also as a anti-metastatic therapeutic agent. The useful therapeutic dose of CAPE for A549 lung cancer cell line was found to be 6 µg/ml and the tolerance dose of normal lung fibroblast for CAPE appeared to be higher than the therapeutic dose¹⁷. It is claimed that CAPE inhibits the growth of tumor cells using oxidative stress pathways connected to p53-independent pathways and observed to inhibit oxidative processes by decreasing the generation of intracellular hydrogen peroxide (H_2O_2) in A549 cells. The decrease in H_2O_2 production and GSH level of A549 cells was seen

in a very rapid and profound manner¹⁷. Moreover, the depleting intracellular stores of GSH (reduced glutathione) by CAPE can render cells more susceptible to oxidative stress-induced apoptosis¹⁸.

Prostate Cancer

Prostate cancer is a highly common malignancy in Western countries and it is resistant to apoptosis. Because of this resistance, it is essential to develop new therapeutic strategies. Because CAPE is a nuclear factor-κB (NF-κB) inhibitor and 5α reductase inhibitor, it has a potential for the treatment of prostate cancer. It was found that CAPE can inhibit NF-κB activation in prostatic cancer-3 (PC-3) cells, by blocking the ability of paclitaxel and tumor necrosis factor-alpha (TNF- α) to activate NF- κ B. This effect is also associated with a reduction in the cellular levels of the inhibitors of apoptosis proteins (cIAP-1, cIAP-2 and XIAP). Note that these proteins are a family of structually homologous caspase inhibitors¹⁹. This is consistent with other reports showing that CAPE prevents NF-κB activation²⁰. CAPE can reduce constitutive activation levels. It is suggested that it could have a role in managing prostate cancer by blocking the NF-κB survival pathway. This is because CAPE may enable lower doses of chemotherapy or radiotherapy to be used to achieve a clinical response, with possibly less overall toxicity. In a study, CAPE was used as second hit agent in association with antisense oligonucleotide to target a region within the baculovirus IAP repeat (BIR) domain of cIAP-1 and examined its ability to facilitate apoptosis in prostate cancer cells²¹. Western blotting showed a down-regulation in cIAP-1 expression and higher levels of spontaneous apoptosis in PC3 and DU145 cells with no alteration in overall cell viability.

Melanomas

CAPE is suggested to suppress reactive-oxygen speies (ROS)-induced DNA strand breakage in human melanoma A2058 cells when compared other potential protective agents²². It was also studied in aspect of *in vitro* biochemical mechanism of its toxicity using tyrosinase enzyme as a molecular target in human SK-MEL-28 melanoma cells. The IC(50) of CAPE towards human skin melanoma (SK-MEL-28 melanoma) cells was found to be 15 μ M. Quinone formation is thought to play an important role in CAPE-induced cell toxicity. When

the role of tyrosinase was investigated in CAPE toxicity, it was noticed that CAPE led to negligible antiproliferative effect, apoptotic cell death and ROS formation in shRNA plasmid-treated cells. Moreover, CAPE selectively caused escalation in the ROS formation and intracellular GSH depletion in melanocytic human SK-MEL-28 cells which express functional tyrosinase²³. The studies on five melanoma cell lines, B16-F0, B16F10, SK-MEL-28, SK-MEL-5, and MeWo and in vivo efficacy study in skin B16-F0 melanoma tumor model in C57BL/6 mice showed that CAPE (10 mg/kg/day) led to intracellular GSH depletion, 10-25 fold increase in ROS formation in B16-F0 cells, caused 5-7 fold increase in apoptosis in B16F0 cells, and led to tumor size growth inhibition²⁴. Tyrosinase bioactivates CAPE to an O-quinone, which reacts with GSH to form CAPE-SH conjugate. The researchers investigated CAPE as a selective glutathione S-transferase (GST) inhibitor in the presence of tyrosinase, which is abundant in melanoma cells. 90% of CAPE was metabolized by tyrosinase after a 60 minutes incubation. It showed 70-84% GST inhibition. CAPE-SG conjugate and CAPE quinone demostrated 85% GST inhibition via reversible and irreversible mechanisms. MK-571, a selective multidrug resistance protein (MRP) inhibitor, and probenecid, a nonselective MRP inhibitor, decreased the inhibitory concentration (IC)50 of CAPE by 13% and 21%, apoptotic cell death by 3% and 13%, and mitochondrial membrane potential in human SK-MEL-28 melanoma cells by 10% and 56%, respectively. Lastly, computational docking analyses suggest that CAPE binds to the GST catalytic active site indicating intracellularly formed quinones and GSH conjugates of CAPE may play major roles in the selective inhibition of GST in SK-MEL-28 melanoma cells²⁵.

References

- EID HM, VALLERAND D, MUHAMMAD A, DURST T, HADDAD PS, MARTINEAU LC. Structural constraints and the importance of lipophilicity for the mitochondrial uncoupling activity of naturally occurring caffeic acid esters with potential for the treatment of insulin resistance. Biochem Pharmacol 2010; 79: 444-454
- WANG SH, CHEN CS, HUANG SH, YU SH, LAI ZY, HUANG ST, LIN CM. Hydrophilic ester-bearing chlorogenic acid binds to a novel domain to inhibit xanthine oxidase. Planta Med 2009; 75: 1237-1240.

- Yoshizumi K, Nishioka N, Tsuji T. The xanthine oxidase inhibitory activity and hypouricemia effect of the propolis in rats. Yakugaku Zasshi 2005; 125: 315-231.
- 4) Wang T, Chen LX, Wu WM, Long Y, Wang R. Potential cytoprotection: antioxidant defence by caffeic acid phenethyl ester against free radical-induced damage of lipids, DNA, and proteins. Can J Physiol Pharm 2008; 86: 279-287.
- Sud'ina GF, Mirzoeva OK, Pushkareva MA, Korshunova GA, Sumbatyan NV, Varrolomeev SD. Caffeic acid phenethyl ester as a lipoxygenase inhibitor with antioxidant properties. FEBS Lett 1993; 329: 21-24.
- BHIMANI RS, TROLL W, GRUNBERGER D, FRENKEL K. Inhibition of oxidative stress in HeLa cells by chemopreventive agents. Cancer Res 1993; 53: 4528-4533.
- FRENKEL K, WEI H, BHIMANI R, YE J, ZADUNAISKY JA, HUANG MT, FERRARO T, CONNEY AH, GRUNBERGER D. Inhibition of tumor promoter-mediated processes in mouse skin and bovine lens by caffeic acid phenethyl ester. Cancer Res 1993; 53: 1255-1261.
- NOMURA M, KAJI A, MA W, MIYAMOTO K, DONG Z. Suppression of cell transformation and induction of apoptosis by caffeic acid phenethyl ester. Mol Carcinogen 2001; 31: 83-89.
- CHIAO C, CAROTHERS AM, GRUNBERGER D, SOLOMON G, PRESTON GA, BARRETT JC. Apoptosis and altered redox state induced by caffeic acid phenethyl ester (CAPE) in transformed rat fibroblast cells. Cancer Res 1995; 55: 3576-3583.
- SU ZZ, LIN J, PREWETT M, GOLDSTEIN NI, FISHER PB. Apoptosis mediates the selective toxicity of caffeic acid phenethyl ester (CAPE) toward oncogenetransformed rat embryo fibroblast cells. Anticancer Res 1995; 15: 1841-1848.
- 11) SU ZZ, LIN J, GRUNBERGER D, FISHER PB. Growth suppression and toxicity induced by caffeic acid phenethyl ester (CAPE) in Type-5 adenovirus-transformed rat embryo cells correlate directly with transformation progression. Cancer Res 1994; 54: 1865-1870.
- 12) BASINI G, BAIONI L, BUSSOLATI S, GRASSELLI F, DAQUINO C, SPATAFORA C, TRINGALI C. Antiangiogenic properties of an unusual benzo[k,l]xanthene lignan derived from CAPE (Caffeic Acid Phenethyl Ester). Invest New Drugs 2012; 30: 186-190.
- Izuta H, Shimazawa M, Tsuruma K, Araki Y, Mishima S, Hara H. Bee products prevent VEGF-induced angiogenesis in human umbilical vein endothelial cells. BMC Comp Alternative Med 2009; 9: 45. doi: 10.1186/1472-6882-45.
- 14) HWANG HJ, PARK HJ, CHUNG HJ, MIN HY, PARK EJ, HONG JY, LEE SK. Inhibitory effects of caffeic acid phenethyl ester on cancer cell metastasis mediated by the down-regulation of matrix metalloproteinase expression in human HT1080 fibrosarcoma cells. J Nutr Biochem 2006; 17: 356-362.

- 15) LIAO HF, CHEN YY, LIU JJ, HSU ML, SHIEH HJ, LIAO HJ, ET AL. Inhibitory effect of caffeic acid phenethyl ester on angiogenesis, tumor invasion, and metastasis. J Agric Food Chem 2003; 51: 7907-7912.
- 16) SHIGEOKA Y, IGISHI T, MATSUMOTO S, NAKANISHI H, KO-DANI M, YASUDA K, HITSUDA Y, SHIMIZU E. Sulindac sulfide and caffeic acid phenethyl ester suppress the motility of lung adenocarcinoma cells promoted by transforming growth factor-beta through Akt inhibition. J Cancer Res Clin Oncol 2004; 130: 146-152.
- 17) CHEN MF, WU CT, CHEN YJ, KENG PC, CHEN WC. Cell killing and radiosensitization by caffeic acid phenethyl ester (CAPE) in lung cancer cells. J Radiation Res 2004; 45: 253-260.
- COTGREAVE IA, GERDES RG. Recent trends in glutathione biochemistry—glutathione-protein interactions: a molecular link between oxidative stress and cell proliferation? Biochem Biophys Res Commun 1998; 242: 1-9.
- 19) McEleny K, Coffey R, Morrissey C, Fitzpatrick JM, Watson RW. Caffeic acid phenethyl ester-induced PC-3 cell apoptosis is caspase-dependent and mediated through the loss of inhibitors of apoptosis proteins. BJU Int 2004; 94: 402-406.
- NATARAJAN K, SINGH S, BURKE TR, GRUNBERGER D, AG-GARWAL BB. Caffeic acid phenethyl ester is a po-

- tent and specific inhibitor of activation of nuclear transcription factor NF-kappa B. P Natl Acad Sci USA 1996; 93: 9090-9095.
- 21) McEleny K, Coffey R, Morrissey C, Williamson K, Zangemeister-Wittke U, Fitzpatrick JM, Watson RW. An antisense oligonucleotide to cIAP-1 sensitizes prostate cancer cells to fas and TNFalpha mediated apoptosis. Prostate 2004; 59: 419-425.
- 22) CHEN CN, Wu CL, LIN JK. Apoptosis of human melanoma cells induced by the novel compounds propolin A and propolin B from Taiwenese propolis. Cancer Lett 2007; 245: 218-231.
- 23) KUDUGUNTI SK, VAD NM, WHITESIDE AJ, NAIK BU, YUSUF MA, SRIVENUGOPAL KS, MORIDANI MY. Biochemical mechanism of caffeic acid phenylethyl ester (CAPE) selective toxicity towards melanoma cell lines. Chem Biol Interact 2010; 188: 1-14.
- 24) KUDUGUNTI SK, VAD NM, EKOGBO E, MORIDANI MY. Efficacy of caffeic acid phenethyl ester (CAPE) in skin B16-F0 melanoma tumor bearing C57BL/6 mice. Invest New Drugs 2011; 29: 52-62.
- 25) KUDUGUNTI SK, THORSHEIM H, YOUSEF MS, GUAN L, MORIDANI MY. The metabolic bioactivation of caffeic acid phenethyl ester (CAPE) mediated by tyrosinase selectively inhibits glutathione S-transferase. Chem Biol Interact 2011; 192: 243-256.