

# LncRNA SNHG7 promotes the proliferation and inhibits apoptosis of renal cell cancer cells by downregulating CDKN1A

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**Abstract. – OBJECTIVE:** Recent studies have revealed that long non-coding RNAs (lncRNAs) have a crucial role in tumor progression. Renal cell cancer (RCC) is a common type of fatal gynecological cancer worldwide. This study aims to identify the role of lncRNA Small nucleolar RNA host gene 7 (SNHG7) in the progression of RCC.

**PATIENTS AND METHODS:** Expression of lncRNA SNHG7 in both RCC cells and 50 pairs of tissue samples was detected by Real-time quantitative polymerase chain reaction (RT-qPCR). Moreover, the function of SNHG7 was identified by performing cell apoptosis assay, colony formation assay and proliferation assay *in vitro*. The underlying mechanism assays including RT-qPCR and Western blot assay were conducted.

**RESULTS:** SNHG7 expression was remarkably upregulated in tumor tissues compared with adjacent tissues. Moreover, RCC cell proliferation was inhibited and cell apoptosis was promoted after knockdown of SNHG7. Moreover, after knockdown of SNHG7, CDKN1A was upregulated at mRNA and protein level *in vitro*. Furthermore, the expression of CDKN1A in tumor tissues was negatively correlated to the expression of SNHG7.

**CONCLUSIONS:** The results above suggest that SNHG7 could promote proliferation and inhibit apoptosis in RCC through downregulating CDKN1A, which may offer a new therapeutic intervention for RCC patients.

**Key Words:** long non-coding RNA, SNHG7, RCC, CDKN1A.

## Introduction

Renal Cell Cancer (RCC), accounting for nearly 10% of all adult malignancies, is the 3rd most frequent urologic carcinoma in the world. The morbidity of RCC has raised in the past two

decades<sup>1</sup>. Approximately 337,000 cases were newly diagnosed with RCC worldwide in 2012. Moreover, it was reported that 52,000 patients and 145,000 died of RCC in Europe and worldwide in 2012, respectively (<http://globocan.gco.net>). Though great advances have been made in RCC diagnosis and therapeutic treatment, the overall survival rate remains less than 10% in patients with distant metastases<sup>2</sup>. Thus, early detection and prevention are very crucial for patients with RCC. It's urgent to realize the underlying mechanism of RCC progression and find out potential biomarkers and therapeutic targets.

The majority of transcripts, which do not code protein, are called non-coding RNAs (ncRNAs). The ncRNAs greater than 200 nt are defined as long non-coding RNAs (lncRNAs). Numerous studies have indicated that lncRNAs play a crucial role in the development of cancers. For example, through targeting miR-221/SOCS3, lncRNA GAS5 suppresses cell proliferation, cell metastasis and gemcitabine resistance in pancreatic cancer<sup>3</sup>. LncRNA TP73AS1 dramatically promotes cell apoptosis, depresses cell proliferation in colorectal cancer and functions as a competing endogenous RNA for miR103 to modulate the expression of PTEN<sup>4</sup>. LncRNA HOTAIR facilitates cell invasion and metastasis in oral squamous cell carcinoma by indirectly recruiting EZH2 and depressing E-cadherin<sup>5</sup>. LncRNA ATB promotes cell migration and cell invasion in glioma by activating astrocytes through suppressing the expression of microRNA 2043p<sup>6</sup>. However, the clinical role of lncRNA AFAP1-AS1 and underlying biological mechanisms in RCC remains unknown.

In our study, lncRNA Small nucleolar RNA host gene 7 (SNHG7) expression was significantly upregulated in RCC samples. Moreover, func-

tion assays revealed that SNHG7 participated in the regulation of RCC cell apoptosis and proliferation. Furthermore, we discovered that lncRNA SNHG7 played its function in RCC by regulating CDKN1A.

## Patients and Methods

### Cell Culture and Clinical Samples

Caki-1, 769-P, 786-O, and ACHN RCC cell lines, a normal human kidney epithelial cell (HK-2) and 293T cell (Chinese Type Culture Collection, Chinese Academy of Sciences, Shanghai, China) were used in this study. Culture medium consisted of penicillin, Dulbecco's Modified Eagle's Medium (DMEM, HyClone, South Logan, UT, USA) and 10% fetal bovine serum (FBS; Invitrogen Life Technologies, Carlsbad, CA, USA). Besides, cells were cultured in a humidified incubator, which contained 5% CO<sub>2</sub> and was set at 37°C. RCC tissues were obtained from 50 RCC patients who underwent surgery at Affiliated Jiangyin Hospital of Southeast University Medical College. All tissues were stored at -80°C. Written informed consent was provided by RCC patients before surgery. This study was approved by the Ethics Committee of Shaoxing County People's Hospital.

### Cell Transfection

For transfection, lentivirus expressing short-hairpin RNA (shRNA) targeting SNHG7 was compounded and then cloned into pLenti-EF1a-EGFP-F2A-Puro vector (Bioset, Inc., San Diego, CA, USA). The lentivirus transfection was performed using Lipofectamine 2000 (Invitrogen, Carlsbad, CA, USA).

### RNA Extraction and Real-Time Quantitative Polymerase Chain Reaction (RT-qPCR)

TRIzol reagent (Invitrogen, Carlsbad, CA, USA) was utilized to isolate total RNA from tissue and cells. RBR green (Roche, Basel, Switzerland) was conducted to measure the relative expression levels between SNHG7 and CDKN1A. GAPDH. Messenger RNA expressions were normalized to the level of glyceraldehyde 3-phosphate dehydrogenase (GAPDH) mRNA. Following are the primers for RT-qPCR: SNHG7 primers forward 5'-GTGACTTCGCCTGTGATGGA-3', reverse 5'-GGCCTCTATCTGTACCTTTAT-3', GAPDH primers forward 5'-CCAAAT-CACTGGGGCAATGCTGG-3' and reverse 5'-TGATGGCATGGACTGTGGTCATTCA-3'.

The thermal cycle was as follows: 30 s at 95°C, 5 s at 95°C for 40 cycles, 35 s at 60°C.

### Colony Formation Assay

1.5×10<sup>3</sup> of 786-O cells were placed in a 96-well plate. 10 day later, colonies were then fixed with 10% formaldehyde for 30 min and stained for 5 min with 0.5% crystal violet. A camera (Tokyo, Japan) was used to take pictures of the colonies.

### Cell Proliferation Assay

Cell proliferation of the treated cells in 96-well plates was monitored every 24 h. Cell Counting Kit-8 (CCK-8) (Dojindo Molecular Technologies, Inc., Kumamoto, Japan). Spectrophotometer (Thermo Fisher Scientific, Waltham, MA, USA) was utilized to detect the absorbance at 450 nm.

### Flow Cytometric Analysis

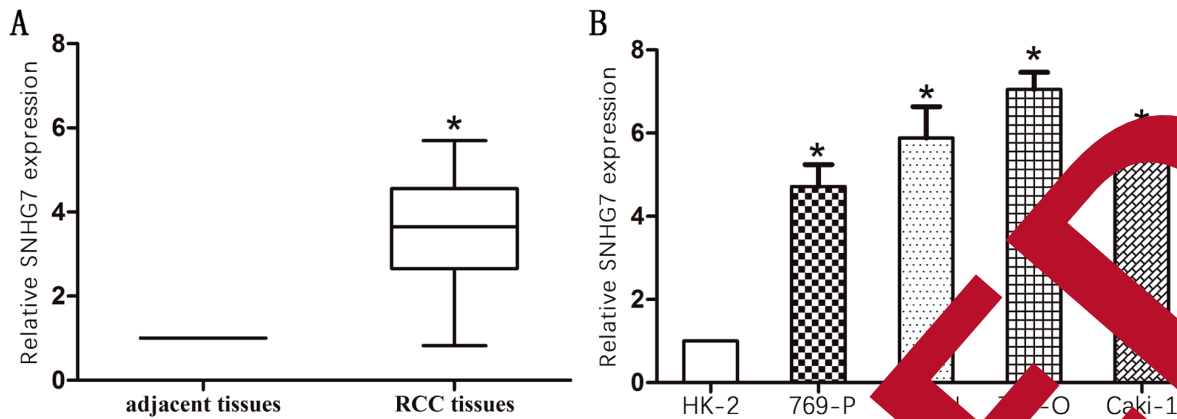
Annexin-V-FITC apoptosis detection kit (BD Biosciences, Franklin Lakes, NJ, USA) was used to detect RCC cell apoptosis. Briefly, harvested cells were washed twice using ice-cold PBS (Phosphate-buffered saline) and then, 100 μL flow cytometry binding buffer was added. After 5 μL Annexin V/FITC and PI were mixed, these cells were stained for 15 min in the dark at room temperature. Each tube was added with four hundred microliters binding buffer. FACSCalibur flow cytometer (BD Biosciences, Franklin Lakes, NJ, USA) was used for analyzing the apoptosis cells.

### Western Blot Analysis

After separated with 12% sodium dodecyl sulfate-polyacrylamide gel electrophoresis (SDS-PAGE), the protein samples were transferred onto polyvinylidene difluoride (PVDF) membrane (Millipore, Billerica, MA, USA). Anti-CDKN1A (Abcam, Cambridge, MA, USA) and anti-GAPDH (Abcam, Cambridge, MA, USA) were used as the primary antibodies, which were utilized for incubating the membranes at 4°C overnight. Next, membranes were incubated with goat anti-rabbit secondary antibody (Abcam, Cambridge, MA, USA) for 2 h. Chemiluminescent film was applied for assessment of protein expression using ImageJ software (NIH, Bethesda, MD, USA).

### Statistical Analysis

Statistical Product and Service Solutions (SPSS) 17.0 (SPSS Inc., Chicago, IL, USA) was utilized to perform statistical analysis. Two-tailed Student's



**Figure 1.** Expression levels of SNHG7 were increased in RCC tissues and cell lines. **A**, SNHG7 expression was significantly increased in the RCC tissues compared with adjacent tissues. **B**, Expression levels of SNHG7 relative to GAPDH were determined in the human RCC cell lines and HK-2 (normal human kidney epithelial cells) by RT-qPCR. Data are presented as the mean  $\pm$  standard error of the mean. \* $p < 0.05$ .

*t*-test was performed to analyze the significance. When  $p < 0.05$ , the data were considered statistically significant.

## Results

### Expression Level of SNHG7 in Tissues and Cells of RCC

First, RT-qPCR was conducted for detecting SNHG7 expression in 50 patients' tissues and RCC cells. As a result, SNHG7 was significantly upregulated in tumor tissue compared with adjacent tissues (Figure 1A). SNHG7 expression level of RCC cells was higher than that of HK-2 (normal human kidney epithelial cells).

### Knockdown of SNHG7 Inhibited Cell Proliferation of RCC Cells

According to SNHG7 expression in RCC cells, we chose 786-O RCC cells for the knockdown of SNHG7. RT-qPCR was utilized for detecting the transfection efficiency (Figure 2A). Moreover, the outcome of CCK-8 assay also revealed that after SNHG7 was knocked down, RCC cell proliferation was remarkably suppressed (Figure 2B). The result of colony formation assay revealed that after SNHG7 was knocked down, the colonies of RCC cells were significantly decreased (Figure 2C).

### Knockdown of SNHG7 Promoted Cell Apoptosis in RCC Cells

To explore whether SNHG7 functioned in RCC cell apoptosis, flow cytometric analysis was performed to detect the apoptosis rate of these treat-

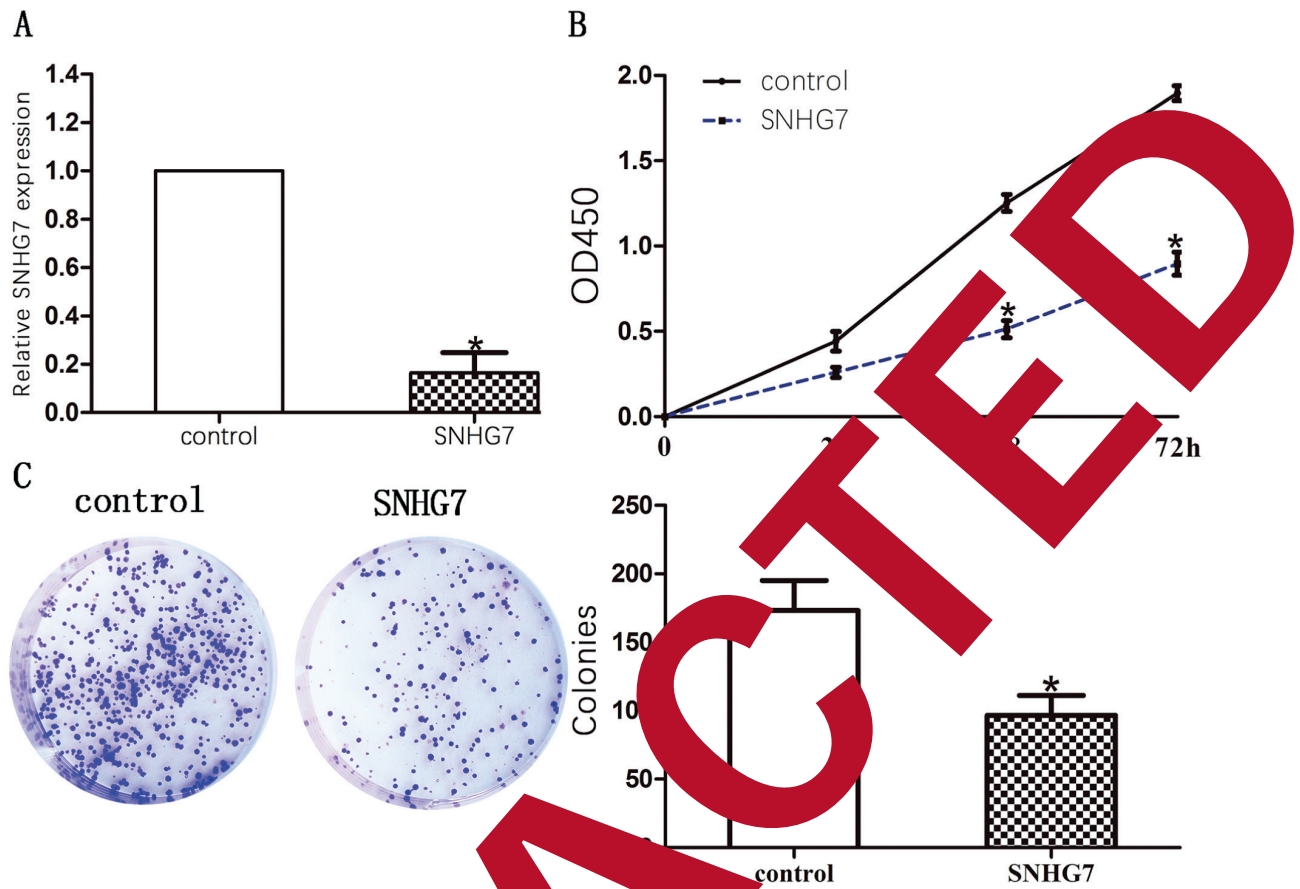
ed cells. The results revealed that after SNHG7 was knocked down in RCC cells, cell apoptosis rate of 786-O RCC cell was remarkably increased (Figure 3).

### SNHG7 Promoted RCC Tumorigenesis via CDKN1A

CDKN1A was a newly discovered gene in various cancers including RCC. We then conducted mechanism assays to identify the correlation between SNHG7 and CDKN1A. RT-qPCR results demonstrated that the CDKN1A mRNA expression was upregulated in RCC cells transfected with SNHG7 shRNA (Figure 4A). Western blot analysis results further verified that the expression level of CDKN1A protein was upregulated in RCC cells transfected with SNHG7 shRNA (Figure 4B). To explore the interaction between SNHG7 and CDKN1A, the expression level of CDKN1A was detected in RCC tissues. As a result, CDKN1A expression was significantly lower in RCC tissues compared with adjacent tissues (Figure 4C). The linear correlation analysis revealed that the CDKN1A expression negatively correlated to SNHG7 expression in RCC tissues (Figure 4D).

## Discussion

Emerging evidence suggested that lncRNAs play an important role in a variety of important biological processes, including the development of RCC. For instance, through negatively regulating the expression of miR-126, lncRNA DUXAP8



**Figure 2.** Silencing of SNHG7 inhibited RCC cell proliferation. **A**, SNHG7 expression in RCC cells transfected with control vector (control) or SNHG7 shRNA (SNHG7) was detected by qPCR. GAPDH was used as an internal control. **B**, CCK-8 assay showed that silencing of SNHG7 significantly inhibited cell proliferation in RCC cells. **C**, Colony formation assay showed that silencing of SNHG7 significantly reduced the number of colonies in RCC cells (magnification: 10×). The results represent the average of three independent experiments (mean ± standard error of mean). \* $p < 0.05$ .

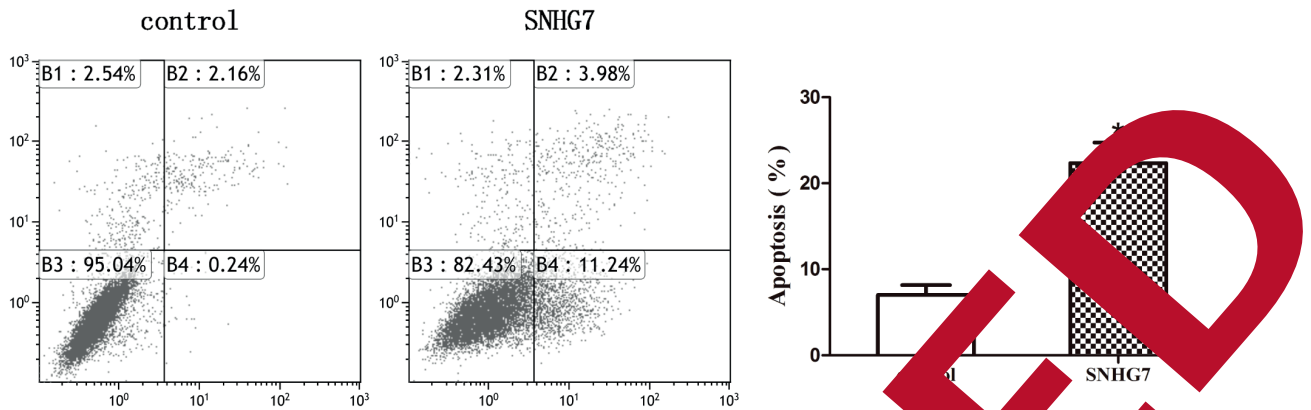
promotes the progression of RCC. Knockdown of lncRNA MAL-1 remarkably increases cell viability and migration in RCC by targeting miR-223p and activating the PI3K/Akt signaling pathway<sup>8</sup>. By binding to miR-193b, lncRNA TUG1 inhibits the proliferation and migration of RCC cell through regulating the YAP expression. Overexpression of lncRNA HOTTIP enhances cell proliferation, migration, and invasion in RCC by regulating the PI3K/Akt/Atg13 signaling pathway. In addition, lncRNA TUG1 functions as an oncogene in RCC<sup>11</sup> by suppressing the expression level of miR196a.

Small nucleolar RNA host gene 7 (SNHG7) is a long non-coding RNA located on chromosome 9q34.3. Numerous researches have revealed that SNHG7 promotes cell proliferation, cell invasion and migration in many cancers<sup>12</sup>. For example, SNHG7 facilitates the epithelial-to-mesenchymal transition

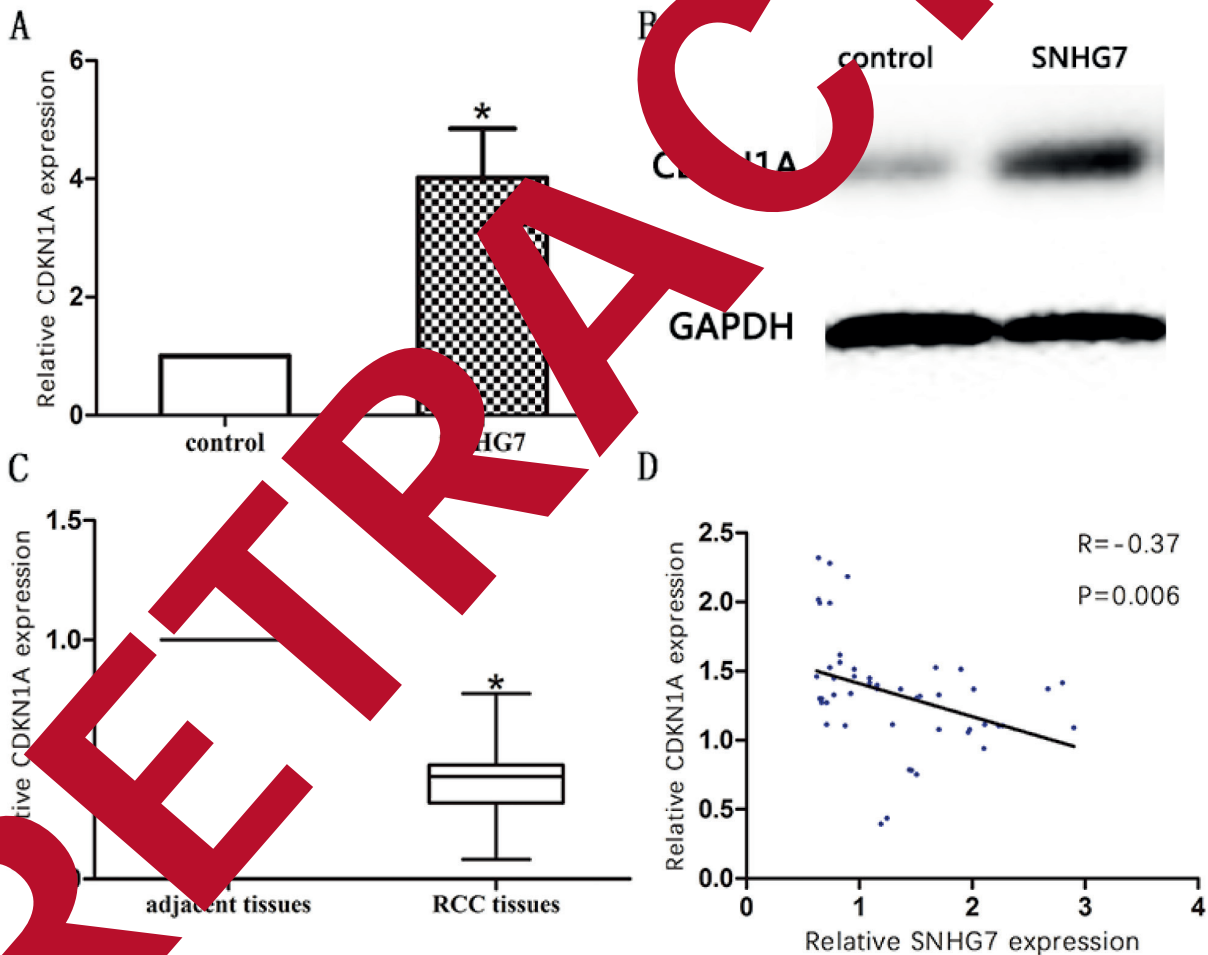
and tumor proliferation in osteosarcoma by regulation of miR-34a<sup>13</sup>. Knockdown of SNHG7 significantly inhibits cell proliferation and cell migration in glioblastoma through inhibition of miR-5095<sup>14</sup>. SNHG7 promotes the progression of non-small cell lung cancer by enhancing miR-193b level and reducing FAIM2 expression<sup>15</sup>. In addition, SNHG7 is upregulated in colorectal cancer and is found to be negatively related to the prognosis of colorectal cancer<sup>16</sup>. SNHG7 was found to be upregulated in both RCC tissues and cells in our study. Furthermore, after SNHG7 was knocked down, the ability of cell growth was inhibited, and cell apoptosis rate was enhanced. These data indicated that SNHG7 functioned as an oncogene and promotes the tumorigenesis of RCC.

CDKN1A (p21<sup>Cip1</sup>) belongs to cyclin-dependent kinase inhibitors, which acts as a tumor suppressor in many cancers<sup>17</sup>. For example, by regulating CD-





**Figure 3.** Silence of SNHG7 promoted RCC cell apoptosis. Flow cytometric analysis may show that the cell apoptosis rate of RCC cells was significantly increased *via* silence of SNHG7 in RCC cells. The results represent the average of three independent experiments (mean  $\pm$  standard error of the mean). \* $p < 0.05$ .



**Figure 4.** Interaction between CDKN1A and SNHG7 in RCC. **A**, The RNA expression level of CDKN1A in SNHG7 shRNA (SNHG7) cells was significantly increased compared with empty control cells in RCC cells. **B**, Protein expression of CDKN1A was increased after silence of SNHG7 in RCC cells. **C**, CDKN1A was significantly downregulated in RCC tissues compared with adjacent tissues. **D**, The linear correlation between the expression level of CDKN1A and SNHG7 in RCC tissues. The results represent the average of three independent experiments. Data are presented as the mean  $\pm$  standard error of the mean. \* $p < 0.05$ .

KN1A transcription expression, LRH-1 inhibits cell proliferation in breast tumor<sup>18</sup>. CDKN1A enhances the response of cutaneous tumors to radiotherapy by manipulating Langerhans cell survival and promoting Treg cell generation<sup>19</sup>. In addition, CDKN1A is closely correlated with prognosis of patients who receive gastric adenocarcinoma resection<sup>20</sup>. Further, by suppressing CDKN1A, EZH2 controls the proliferation of germinal centers B cell and enables cell cycle progression<sup>21</sup>. The result of Western blot analysis indicated that CDKN1A was upregulated after SNHG7 was knocked down *in vitro*. Moreover, negative correlation between CDKN1A and SNHG7 expression was discovered in tumor tissues. The results above revealed that CDKN1A was a target regulated by SNHG7 in RCC development.

### Conclusions

We found that lncRNA SNHG7 might be a new biomarker in the carcinogenesis of RCC and could be served as a promising mark for RCC.

### Conflict of Interests

The Authors declare that they have no conflict of interests.

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