Long noncoding RNA TTN-AS1 promotes the proliferation and migration of prostate cancer by inhibiting miR-1271 level

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Abstract. – OBJECTIVE: Prostate cancer is one of the most common malignant tumors. Recently, the role of long noncoding RNAs (IncRNAs) in tumor progression has been well concerned by numerous researchers. In this research, IncRNA TTN-AS1 was studied to identify its biological function in the progression of prostate cancer.

PATIENTS AND METHODS: Firstly, Real Time-quantitative Polymerase Chain Reaction (RT-qPCR) was conducted to measure TTN-AS1 expression in prostate cancer tissues. Furthermore, *in vitro* role of TTN-AS1 in regulating prostate cancer cells was assessed. Tum mation assay was conducted in NOD/SC in to explore the *in vitro* function of TTN-1. In addition, the luciferase reporter gene assays performed to analyze the relationship beth TTN-AS1 and miR-1271.

RESULTS: TTN-AS1 expres remai ably higher in prostate ca es com pared with that of corres ding d . Moreover, proliferation and ration prostate cancer cells were inhibit silenced. MiR-1271 up silence of TTN-AS urther m sm assays 71 was a d showed that mil arget of TTN-AS1 in pr ncer. In add tumor abilities were inhibitleta: formation an ed after in vivo knocko TTN-AS1. CONC ONS: Our stu covers a poten-

tial once ene in prostate career and demonstrate nat TTN-AS1 enhances cell proliferation and ration sponging miR-1271.

Key W

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TN-AS1, Prostate cancer,

Introduction

of calcer-related death in 2018¹. The surgery or androgen deprivation therapy is available for most of the new er patients nosed prosta. wever, the currence and at an ear la_b recurrence rates o. tate cancer significantly increase in both devel and developing counth makes it the pard-leading cause of the in male malignancies all over the world²⁻⁵. tri erefore, it is y ent to find out the underlying hanism of pa genesis of prostate cancer and out a new atment strategy. ng RNAs (IncRNAs) are one

LncRNA TTN-ASI is a novel lncRNA, and its function in cancers has been focused on. Our study aimed to identify whether TTN-AS1 participates in the proliferation and migration of prostate cancer and its potential mechanism.

Patients and Methods

Tissue Specimens

58 prostate cancer patients received surgery at Baoji Central Hospital. Their tissue samples were surgically resected and immediately preserved at -80°C. All tissues were analyzed by an experienced pathologist. The Research Ethics Committee of Baoji Central Hospital ratified this study and the written informed consent was offered by the patients.

Cell Culture

The human prostate cancer cell lines (22Rv1, DU145, and LNCaP) and normal human prostate epithelial cell line (P69) were offered by the Institute of Biochemistry and Cell Biology, Chinese Academy of Science (Shanghai, China). The cell culture was conducted in Dulbecco's Modified Eagle's Medium (DMEM; Gibco, Rockville, MD, USA) containing 10% fetal bovine serum (FBS; Life Technologies, Gaithersburg, MD, USA) and 1% penicillin. Besides, the cells were cultured in an incubator containing 5% CO, at 37°C.

Cell Transfection

shRNAs directed against TTN-AS1 were synthesized by GenePharma (Shanghai, China). The complementary DNA encoding TTN-AS1 was amplified and inserted into pcDNA3.1 (Gene-Pharma, Shanghai, China), which were then used for the transfection of 22Rv1 cells with polybrene (Genepharma, Shanghai, China).

RNA Extraction and Real Time-Ouantitative Polymerase Chain Reaction (RT-qPCR)

The total RNA from tissues and ce extracted by using TRIzol reagent (I <u>-</u>0gen, Carlsbad, CA, USA). RNA was rev transcribed to cDNAs through reverse T scription Kit (TaKaRa Biotechne v Co., Li Dalian, China). Thermocycli ons wer as follows: pre-denaturati at 95° 🕻 5 min. denaturation at 95°C for s and a aling at 60°C for 30 s, for a total vc method was utiliz for C ng relative expression. The ler sequen e as folrd: 5'-TCC lows: TTN-AS GCAT-CACCTAGC erse: 5'-GAAGGAGa GAAGTAGAOTCATI glyceraldehyde 3-phosph dehydrogen GAPDH), forward: -GCACCGTCAA CTGAGAAC-3' '-TGGTGAAGACGCCAGTGand verse: GA

Coll Pro. tion

0³ transfer of cells were seeded in 96-well plat and cell proliferation was assessed by Cell Provide Eagent Kit I (MTT; Roche, Basel, atzerland, at 0 h, 24 h, 48 h, 72 h following the afacturer's protocol. Absorbance at 490 nm were essed using an enzyme-linked immunosorbeat assay (ELISA) reader system (Multiskan Ascent, LabSystems, Helsinki, Finland).

Colony Formation Assay

 1.5×10^3 22Rv1 cells per well were inoculated in a 6-well plate. 10 days later, the colored then fixed with 10% formaldehyde and 50 m and stained for 5 min with 0.5% crueal violet. A Nikon camera (Tokyo, Japan) ware ad for taking photographs of colonies.

Ethynyl Deoxyuriding dU) Incorporation Assay

is.

The EdU Kit (Roce Basel, ditzerland) was used for detecting central service. A representative photograph of taken of ss Axic of Photomicroscope (a Zeiss, Observermany).

Transwe

24 h arter tran on, 2 $\times 10^5$ cells in 100 µL serum-free DME e transformed to the are insert (Corning, to er of an 8-µm ning, NY, USA). 20% FBS-DMEM was addto the bottom hamber of the culture inserts. later, these erts were fixed by methanol min and st ed by hematoxylin for 20 min. f oscope (×40) was utilized for An atory cells in three random fields. counting

iferase Reporter Gene Assay

PGL3 vector (Promega, Madison, WI, USA) as wild-type (WT) 3'-UTR. The site-direction mutagenesis of the miR-1271 binding site in TTN-AS1 3'-UTR as mutant (MUT) 3'-UTR was conducted through quick-change site-directed mutagenesis kit (Stratagene, La Jolla, CA, USA). After co-transfection, the luciferase assay was conducted on the dual-luciferase reporter gene assay system (Promega, Madison, WI, USA).

Tumor Formation Assay

After transfection, the 22Rv1 cells (6×10^5 /mL) were injected into NOD/SCID mice (6 weeks old) subcutaneously. Tumor diameter was detected every 5 days. Tumor volume was calculated as the formula (volume = length × width² × 1/2). The tumors were extracted after 4 weeks.

Tumor Metastasis Assay

The transfected 22Rv1cells were injected into the tail vein of NOD/SCID mice (6 weeks old). The mice were sacrificed, and the lungs were extracted after 4 weeks. Then the number of metastatic nodules in the lung was counted. The research was approved by the Animal Ethics Committee of Baoji Central Hospital.

Statistical Analysis

All statistical analyses were carried out using GraphPad Prism 5.0 (La Jolla, CA, USA). The difference between the two groups was compared by the Student's *t*-test. The statistical significance was defined as p<0.05.

Results

TTN-AS1 Expression Level in Prostate Cancer Tissues and Cells

First, RT-qPCR was conducted for detecting TTN-AS1 expression in 58 prostate cancer tissues and 3 prostate cancer cell lines. As a result, TTN-AS1 was significantly upregulated in tumor tissue samples (Figure 1A). TTN-AS1 level in prostate cancer cells was higher than that of P69 (Figure 1B).

Knockdown of TTN-AS1 Inhibited Cell Proliferation and Migration in DU145 Cells

In our study, we chose the 22Rv1 cell line for establishing the knockdown model of TTL Then, the RT-qPCR was performed for the transfection efficacy (Figure 2A). M assay showed that the cell growth ability of cells was significantly repressed after TTNknockdown (Figure 2B). The format assay showed that the color was sig nificantly reduced after known 10wn [N-AS1] (Figure 2C). Furthermore EdU in poration assay also showed that the er tive cells was reduce fter k wn of Th

AS1 in 22Rv1 cells (Figure 2D). The transwell assay showed that the number of migratory cells was significantly reduced after TTN-through knocked down (Figure 2E).

The Interaction between No. 271 and TTN-AS1 in Prostate Ca.

tarbase Starbase v2.0 (http:/ du.cn/ o find the mirLncRNA.php) was us that contained a compl ntary base with AS1. Because miR-1 was a or suppres. or and was able to supply ell proliferation, g these we focused on -127 RNAs **C**1 gure 3A). which were int cted with Indeed, RT the expresassay showe higher in TN -AS1/shRNA sion of mi 27 cells than that in the ol cells (Figure 3B). Furthermore the dual-lue reporter gene assay at the co-trans. on of TTN-AS1-WT re miR-1271 mimics largely decreased the lucifse activity, w e the co-transfection of TTN-1271 mimics had no effect on MUT and n y (Figure 3C). Meanwhile, the ferase act th TTN-AS1 was negatively corexp related Sigure 3D). In summary, these data demonstrated 271 was a direct target of TTN-AS1.

TTN-AS1 Knockdown Inhibited Tumor Formation and Metastasis in vivo

Next, *in vivo* effect of TTN-AS1 on tumor formation was detected. The tumor size in TTN-AS1/shRNA group was smaller compared with the empty vector (CONTROL) group (Figure 4A). The number of metastatic nodules in the lung



^{1.} The expression levels of TTN-AS1 were upregulated in prostate cancer tissues and cell lines. **A**, TTN-AS1 expression was a relative to GAPDH were determined in the human prostate cancer cell lines and P69 (normal human prostate epithelial cell lines) by RT-qPCR. The data are presented as the mean \pm standard error of the mean. *p<0.05.



ells (magnification: 10×). **D**, EdU incorporation assay showed that EdU-positive cells were reduced after the knockdown of N S1 in 22Rv1 cells. **E**, The transwell assay showed that the number of migratory cells was reduced after the knockdown of TTN-AS1 in 22Rv1 cells (magnification: 40×). The results represent the average of the three independent experiments (mean \pm standard error of the mean). **p*<0.05, as compared with the control cells.

rrela



Figure 3. Reciprocal repression between TTN-AS with was upregulated in TTN-AS1/shRNA group comp and TTN-AS1-WT strongly decreased the luciferase WT did not change the luciferase activity. **D**, The lin in prostate cancer tissues. The results represent the aver standard error of the mean. p<0.05.

roup from the TTN-AS1/shRN significantly reduced compare the er v vector (CONTROL) group (Figu expression levels of miR-1271-In extracted tumor ti es were de y RT-qP-CR. The result red that T 1 was downregulate 1/shRNA group compared with the empty CONTROL) group Conversely, (Figure 4 271 was highly roup compared express in TTN-AS1/shRN. empty ctor (CONTROL) group (Figure with 4D abo esults suggested that TTN-AS1 mor for could ion *in vivo*.

scussion

e has proved that lncRNAs play portant lies in prostate cancer. For instance, NA PCSEAT enhances cell proliferation of cancer by activating EZH2, which may offer a potential therapeutic target¹⁰. LncRNA SNHG7 promotes cell proliferation and cycle



A, The binding sites of miR-1271 on TTN-AS1. B, MiR-1271 CONTROL) group. C, Co-transfection of miR-1271 ty, white on of the negative control (NC) and TTN-AS1the expression levels of TTN-AS1 and miR-1271 independent experiments. Data are presented as the mean \pm

progression in prostate cancer through miR-503/ Cyclin D1 pathway¹¹. The downregulation of IncRNA PVT1 inhibits the metastasis of prostate cancer cells by regulating p3812. LncRNA MALAT1 and HOTAIR have been reported to play opposite roles in estrogen-induced transcriptional regulation in prostate cancer cells¹³.

LncRNA TTN antisense RNA 1 (TTN-AS1), located on 2q31.2, is a novel lncRNA that is reported to be upregulated in many cancers. For example, IncRNA TTN-AS1 is aberrantly expressed in hepatocellular carcinoma¹⁴. LncRNA TTN-AS1 functions as an oncogene in esophageal squamous cell carcinoma¹⁵. LncRNA TTN-AS1 enhances cell proliferation and aggressiveness in cervical cancer by targeting miR-573¹⁶. Our previous study showed that TTN-AS1 was upregulated in prostate cancer patients. Then, we conducted some functional experiments and found that after TTN-AS1 was knocked down, the proliferation and migration abilities of prostate cancer cells were inhibited, suggesting that TTN-AS1 could promote tumorigenesis of prostate cancer.



Figure 4. The knockdown of TTN-AS1 inhibited over form calculated respectively in empty vector (CONTROL) N-AS1 static nodules in the lung from the TTN-AS1/shRNA groups of group. **C**, The relative expression of TTN-AS1 in tumor in tumors were examined by RT-qPCR. The ba are presented as

erlying To further identify the chanism of how TTN-AS1 affects tate cancer, miR-12 out as me was oRNA of potential binding S1 by bioinformatic analy cation. experimenta MiR-1271 is y as a tumor suppressor ЛY gulates diverse bioin many carcinomas, wh logical pr sses. MiR-12 ownregulated in cancer and participues in regulating evelopment by targeting Wnt signaling colore tum M 271 could suppress cell metaspat Recently, miR-1271 is tic canc tasis o growth ability by targetorted ess o incer¹⁹. G in p ne presen dy, miR-1271 expression was lated after the knockdown of TTN-AS1. 1271 could directly bind to TTNand mR-1271 expression was negatively lated with TTN-AS1 in prostate cancer. results above suggested that TTN-AS1 might promote tumorigenesis of prostate cancer via sponging miR-1271. Besides, the tumorigen**A**. After tumor extraction, the tumor volume was p and made into a graph. **B**. The number of metaneantly educed compared to the empty vector (CONTROL) amined by RT-qPCR. **D**. The relative expression of miR-1271 s the mean \pm SD of three independent experiments. *p< 0.05.

esis assay revealed that knockdown of TTN-AS1 could inhibit tumor formation and metastasis *in vivo*. Through the detection of TTN-AS1 and miR-1271 expressions in those extracted tumors, we found that TTN-AS1 was downregulated and miR-1271 was upregulated in nude mice administrated with TTN-AS1 shRNA.

Conclusions

The results of the study indicated that TTN-AS1 could enhance proliferation and migration abilities of prostate cancer cells by sponging miR-1271. These findings implied that lncRNA TTN-AS1 could act as a prospective therapeutic target for prostate cancer.

Conflict of Interests

The Authors declared that they have no conflict of interests.

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