Novel insights on acute myocarditis in pediatric patients

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Abstract. – Acute myocarditis (AM) is an inflammatory affliction of the heart muscle characterized by recent onset with a broad spectrum of clinical manifestations that globally affect millions of individuals, notably children and young adults.

The absence of distinct patterns of onset or predictable progression poses a significant threat to survival, potentially leading to advanced heart failure and malignant arrhythmias.

Myocardial fibrosis, a hallmark of myocardial remodeling, is increasingly recognized as a contributor to adverse outcomes in acute myocarditis cases. Advances in molecular and immunological techniques have highlighted the intricate interplay between viral infections, dysregulated immune responses, and genetic susceptibility.

Currently, there is no clear consensus for diagnosis or ongoing follow-up in pediatric patients. The conventional diagnostic tool, endomyocardial biopsy (EMB), considered the gold standard, has been complemented by the effectiveness of cardiac magnetic resonance imaging (CMRI) techniques. Given the procedural complexities and associated complications, there is a pressing need to explore non-invasive alternatives. In this context, biomarkers emerge as promising contenders by evaluating both the inflammatory processes and cardiac remodeling, providing valuable observations into disease severity, progression, and treatment response. Therapeutic strategies in these cases, focusing on the specific pathways or immune components associated with the etiologies, have exhibited promise for better outcomes.

Acute myocarditis in children remains a multifaceted clinical challenge, necessitating a comprehensive understanding of its pathophysiology, diagnosis, and management. This review aims to delve into novel insights surrounding the pathophysiology, diagnosis, and management of acute myocarditis in pediatric patients.

Key Words:

Biomarkers, Cardiac fibroblasts, Circulating miRs, Children, Fibrillar collagens, Heart dysfunction, in-

flammatory signals, Myocarditis, Myocardial fibrosis, TGF- β signaling.

Introduction

Acute myocarditis (AM) stands as an inflammatory myocardial injury characterized by a recent onset (<1 month), presenting a diverse range of clinical trajectories that often pose challenges in diagnosis. Its lack of specific onset patterns and unpredictable evolution can negatively impact prognosis through advanced heart failure and malignant ventricular tachyarrhythmias¹. Globally, the prevalence of AM has been reported to range from 10.2 to 105.6 cases per 100,000 individuals, translating to an annual occurrence of approximately 1.8 million cases². However, the accurate incidence and prevalence in the pediatric population are likely underestimated due to numerous cases not necessitating medical attention. Clinically, acute myocarditis constitutes about 0.05% of hospital admissions, predominantly affecting male adolescents with a prevalence of 81%³. The age distribution displays a bimodal pattern with peaks at 1 year and 16 years, while the most severe forms are observed in the initial year of life, extending up to the age of 4 and in adolescents⁴. Histological studies indicate myocarditis prevalence varying between 0.12% and 12%⁵. Notably, AM contributes to up to 46% of dilated cardiomyopathies and approximately 4% of heart failure cases, with a proportion of these cases associated with a poor prognosis^{1,6,7}. In the age groups affected by myocarditis-related deaths, there has been a change with a shift from children and teenagers to adults as a result of the crucial role of healthcare systems in managing myocarditis⁸. In the pediatric population, is often challenging to establish whether newly diagnosed heart failure or arrhythmias originate from an infectious trigger or are secondary to pre-existing cardiac conditions^{9,10}. Currently, there is no consensus on the gold standard diagnostic approach for children. Histological verification, obtained through endomyocardial biopsy (EMB), is limited due to its high-risk nature and low specificity^{11,12}. Cardiac magnetic resonance imaging (CMRI), an advanced imaging method, can be complicated by the requirement for deep sedation in young children and, in some cases, limited utility due to unstable arrhythmias¹¹.

This review aims to delve into novel insights surrounding acute myocarditis in pediatric patients.

Pathogenesis

While the etiology of acute myocarditis frequently remains undisclosed, confirmed cases, as indicated by the American Academy of Pediatrics, predominantly point towards an infectious or immune-mediated process as the underlying trigger, as shown in Figure 1.

Immune Cells and Inflammasome Molecular Mechanisms

Inflammation serves as a crucial defense mechanism in higher organisms, constituting the immune system's initial response. This response functions to eliminate injurious stimuli, such as infectious pathogens, damaged cells, or irritants, thus paving the way for the commencement of the healing process^{13,14}. This fundamental process is intrinsic to all forms of myocardial injury¹⁵. The pathogenesis of myocarditis has been extensively elucidated through murine models, with these findings equally applicable to humans¹⁶. Among children, viral infection has emerged as the predominant cause of myocarditis¹⁷, with compelling evidence underscoring the direct role of viral myocardial triggers and concurrent pathological host immune responses¹⁵. This intricate interplay occurs through a stepwise progression, as shown in Figure 2.

Furthermore, the type of virus involved determines the specific cardiac cells that are affected (Table I). For instance, Parvovirus B19 (Parvo B19) targets vascular endothelial cells and cardiomyocytes through interactions with the erythrocyte P antigen (P-Ag) and integrin

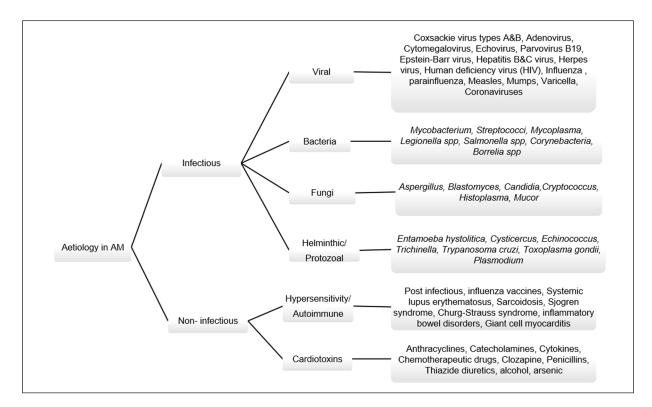


Figure 1. Diagram of major causes in AM.

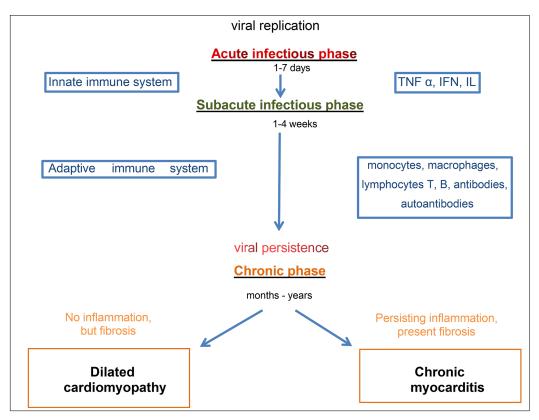


Figure 2. Three-phase model for the pathogenesis of viral myocarditis. TNF, tumour necrosis factor; IFN, interferon; IL, interleukin.

 $\alpha\nu\beta1$ as a co-receptor. Human Herpetic Virus-6 (HHV-6) employs a cluster of differentiation (CD) 46 as a cellular receptor for CD4+ lymphocyte (Ly) T and endothelial cells. Enteroviruses, on the other hand, exert a direct cytolytic effect on cardiomyocytes. Coxsackie B and adenoviruses trigger cytokine synthesis and cell-mediated immune responses *via* the coxsackievirus and adenovirus receptor (CAR), utilizing the decay-accelerating factor (DAF) and integrins ($\alpha\nu\beta3$ and $\alpha\nu\beta5$) as additional receptors¹⁷⁻²². A suggested mechanism of viral invasion of SARS-CoV-2 is through angiotensin-converting enzyme 2 (ACE2) receptor, helped by a host transmembrane serine protease 2, targeting the myocytes²³.

Table I. Cardiotropic viruses and their target-cells in the pathogenesis of acute myocarditis.

| Virus | Receptor/Co-receptor | Target cells | References |
|--------------------------|--|-----------------------------------|------------|
| Parvo-B19 | pAg, αvβ1 | Endothelial cells cardiomyocytes | |
| HHV-6 | CD46, CD4+ LyT | Endothelial cells | 17,19-22 |
| Coxsackie B Adenoviruses | CAR, DAF,ανβ3, ανβ5 | Cardiomyocytes | |
| SARS-CoV-2 | ACE2 receptor, transmembrane serine protease 2 | Cardiomyocytes interstitial cells | 23 |

(Parvo B-19) Parvovirus B19; (pAg) erythrocyte P antigen; (HHV-6) Human Herpetic Virus-6; (CD) cluster of differentiation; (CAR) Coxsackievirus and Adenovirus receptor; (DAF) Decay-accelerating factor; ($\alpha\nu\beta1,\alpha\nu\beta3,\alpha\nu\beta5$) integrins; (ACE2) angiotensin-converting enzyme 2.

Briefly, the immunological cascade in myocarditis appears to initiate with the involvement of innate immune system cells. This process involves encapsulating viral compounds and deceased cells as pathogen-associated molecular patterns (PAMPs) and danger-associated molecular patterns (DAMPs), respectively. These components interact with specialized membrane receptors known as pattern recognition receptors (PRRs)^{24,25}. Among these receptors, the NODlike receptor subtype 3 (NLRP3) is particularly well-described and implicated in the inflammatory disease process¹⁸. Cellular injury prompts nonspecific markers like reactive oxygen species to upregulate NLPR-3 and similar receptors, which identify antigens, whether exogenous or endogenous [e.g., Toll-like receptors (TLR)], ultimately functioning as inflammasomes. This multimeric protein complex assembles in the cytosol and activates the dormant pro-caspase-1, an enzyme responsible for cleaving pro-interleukin (IL)-1β and IL-18, as well as an inducer of pyroptosis^{18,26,27}. In turn, IL-1 β activates the innate immune system by recruiting specific cells to the site of inflammation and modulating the adaptive immune system. Furthermore, IL-18 triggers the synthesis of interferon and enhances the activation of natural killer (NK) cells, thereby augmenting major histocompatibility complex antigens on myocytes and eliminating infected cells. In the later stages of this process, T-lymphocytes are also activated by IL-18. Both CD4+ and CD8+ T cells respond to the infected cardiac cells through the complement system, antibody-dependent toxicity, and direct injury via T cytotoxic cells^{18,27}.

Human inflammatory caspase-4 and caspase-5, through a non-canonical, dependent caspase-1 activation pathway facilitated by their Caspase-Associated Recruitment Domain (CARD), are capable of autonomously activating pro-1 β and IL-18²⁸. Simultaneously, while inflammasomes are activated to eliminate infectious pathogens, another pathway that contributes to this process emerges pyroptosis, an inflammation-induced programmed cell death^{24,27}. Conversely, caspase-1 can facilitate the pyroptosis process in a canonical manner within infected cardiac cells, engendering the creation of pores and the suppression of mitochondrial action potential via gasdermin D (GSDMD), a pyroptotic factor that is significantly upregulated in both epithelial and inflammatory cells²⁷. These pores lead to the influx of ions and water into the cell, eventually causing a cellular rupture and releasing an abundance of cytoplasmic contents, including proinflammatory factors, alarmins, DAMPs, and nuclear and mitochondrial DNA. Additionally, GSDMD potentially plays a role in binding to cardiolipin (CL), contributing to the formation of pores on cell membranes, as well as on the inner endosome and phagosome membranes^{27,28}. Finally, to facilitate inflammation resolution, certain regulatory factors are indispensable. Modulating inflammasome activation necessitates phosphoinositide-3-kinase (PI3K) to function within a negative feedback loop. Similarly, IL-1 receptor-associated kinase-M (IRAK-M) and suppressor of cytokine signaling-1 (SOCS-1) contribute by dampening Toll-like receptor (TLR) signalling²⁹.

Autoimmune and Auto-Inflammatory Theories

From a physiological perspective, the inflammatory pathway facilitated by profibrotic signaling culminates in the healing process through replacement fibrosis. However, in some cases, this process can evolve into a chronic autoinflammatory or autoimmune state, resulting in myocardial scarring and progressive heart failure^{15,30}. While autoimmunity's involvement has been explored in murine models, research in humans remains limited¹⁶. Autoimmunity, understood as the outcome of self-tolerance loss, results from an imbalance between pro- and anti-inflammatory factors within both innate and adaptive immune systems. Recent studies^{22,30} have unveiled interconnected genes that may partially explain myocardial autoimmunity pathways. These genes encompass TLR-signalling adaptor Myd88, cytokines and IL-4, chemokines, and their receptors, complement C3, and complement receptor Cr2.

Conversely, when the immune system fails to effectively clear viral material, significant viral loads persist within the myocardium, leading to a chronic, low-grade, local inflammation. In this context, T-helper (Th)1 cells play a pivotal role, acting as a primary molecular factor contributing to the chronic inflammation response²⁹. The Th1 proinflammatory pathway, mediated by IL-12 and gamma interferon (IFN-y), drives myocardial inflammatory infiltrate. This process is modulated by INF- γ synthesis, a phenomenon altered in certain murine models with defective T-Box transcription factor TBX21 (T-bet) mice. T-bet prompts the development of Th1 lineage from naive Th1 precursor cells, both by activating Th1 and repressing opposing Th2 and Th17 genetic programs. Furthermore, the diminished response of Th2 cells results in deficient pro-healing signals²⁹, a susceptibility observed in those developing autoimmune myocarditis^{13,22}.

Furthermore, the Th2 pathway, mediated by IL-4, is involved in specific types of autoimmune myocarditis characterized by predominant eosinophil presence. Similarly, Th17 exerts its effects through IL-17 expression, playing a crucial role in heart failure progression rather than cardiac inflammation²⁵. A study³¹ demonstrated that CD4+ T cells, stimulated by an initial transient IL-23 signal, are essential for establishing cardiac autoimmunity. Without sustained IL-23 action, autoreactive T cells suppressed IL-17A production.

The persistence of inflammation may continue despite effective viral clearance, as circulating antibodies could serve as inflammatory triggers, as evidenced in biopsy-proven myocarditis³². Moreover, B-cells are activated by dendritic cells that present viral antigens, contributing to humoral immunity by producing specific antibodies. In certain high-risk patients, B-cells may generate autoantibodies targeting shared epitopes or self-cell proteins like β I-adrenergic receptor, cardiac myosin, troponin, mitochondrial compounds, and Na/K-ATP-ase²².

Finally, several studies^{3,15,22} underscore the role of B-cells in sustaining myocardial autoimmunity, identifying immunoglobulin (Ig) G and IgG3 subclasses (potent triggers for complement cascade activation following myocyte lysis) in patients with dilated cardiomyopathy and end-stage heart failure.

Etiological Diagnosis Tools

Endomyocardial Biopsy

EMB stands as the definitive diagnostic method for myocarditis¹. The European Society of Cardiology (ESC), guided by an expert consensus group on myocardial diseases, advocates for EMB application in all patients suspected of myocarditis. However, it is acknowledged that in the pediatric population, diagnosis remains largely clinical due to concerns regarding the potential risks of the procedure¹. In the late 1980s, the Dallas criteria emerged as a tool to furnish a histopathological depiction of myocarditis. These criteria encompassed the presence of inflammatory infiltrates, primarily lymphocytes and macrophages, along with concurrent myocyte necrosis or damage not indicative of an ischemic event. These criteria serve as a basis for diagnostic determination³³. Standard histological examination is a qualitative approach that introduces significant interobserver variability. Immunohistochemistry, when compared to histology alone, offers heightened sensitivity in myocarditis diagnosis³⁴. It enhances diagnostic accuracy by quantifying the number of leukocytes per square millimeter and characterizing inflammatory cells, particularly those bearing CD markers. Additionally, immunohistochemistry holds prognostic value^{35,36}. Moreover, the inclusion of nested real-time polymerase chain reaction (rt-PCR) to detect viral genomes in EMB samples contributes further to diagnostic capabilities¹⁵, as evidenced in Figure 3.

EMB diagnostic biases can arise due to the uneven distribution of inflammation across different myocardial regions. Notably, the left ventricular lateral free wall is more frequently affected than the more accessible right ventricle, potentially leading to sampling bias. To address this, the utilization of low-amplitude electrocardiograms has been proposed to better guide biopsy toward affected myocardial areas³⁷. Despite the perceived risks associated with EMB, a considerable cohort of German pediatric patients has shown a different perspective. The ESC Working Group for Myocardial and Pericardial Disease advocates for an increased application of EMB in the pediatric population³⁸. In such cases, the utilization of immunochemistry on EMB specimens and the accuracy of PCR in detecting viral genomes offer valuable insights crucial for accurate diagnosis and early therapeutic management³⁸. Furthermore, three cohorts studies³⁹⁻⁴¹ have reported that major adverse events stemming from the procedure occurred in 2.6%³⁹, 5%⁴⁰, and 13.2%⁴¹ of biopsied infants with suspected cardiomyopathy. A less invasive alternative to EMB entails conducting PCR analysis on tracheal aspirate samples in young patients with myocarditis. Interestingly, the EMB specimens showed similarities with those collected from the respiratory tract⁴². Nonetheless, the utilization of biopsy remains confined to carefully selected cases within the pediatric population¹.

Immunology Viral Straining

Viral genomes play a prominent role in the etiology of myocarditis^{17,36}. Among these, no-table culprits include enteroviruses, particularly coxsackievirus, adenovirus, parvovirus B19, human herpesvirus 6, Epstein Barr virus, and the more recent addition of SARS-CoV-2^{19,21,22}.

| FIRST BIOPSY Active myocarditis : > 5 cells infiltrate, myofibrillar degeneration, edema | IMMUNOHISTOLOGICAL CRITERIA (WHO/ISFC)* Active and ongoing myocarditis: >14 leucocytes/mm2 > 4 CD68 macrophages/mm2 ≥ 7 cells/mm2 CD3 positive T lymphocytes | | |
|--|---|--|--|
| Borderline myocarditis : > 5 cells infiltrate | Fulminant myocarditis: > 50leucocytes/mm2 | | |
| No myocarditis : no infiltrate | IMMUNOHISTOLOGICAL CRITERIA IN CHILDREN | | |
| SUBSEQUENT BIOPSIES Ongoing myocarditis : > 5 cells infiltrate, myofibrillar degeneration, edema Resolved myocarditis : no infiltrate, but focal | Active and ongoing myocarditis: 7 leucocytes/mm2 2.5 cells/mm2CD3 positive T lymphocytes 0.5 cells/mm2CD20 positive B lymphocytes 4cells/mm2 CD68 macrophages Fulminant myocarditis: > 50leucocytes/mm2 | | |
| fibrosis | RT-PCR ON VIRUSES | | |
| | NEGATIVE in autoreactive myocarditis | | |
| | POSITIVE in viral myocarditis and dilated cardiomyopathy | | |

Figure 3. Comparison of histological, immunohistochemical criteria and the predictability of rt-PCR on viral genome in EMB- patients^{11,12,25,36}. (WHO) World Health Organization, (ISFC) International Society and Federation of Cardiology.

Despite the diversity of these viruses, they converge on a common pathogenic autoimmune process, resulting in chronic inflammation and tissue remodeling that detrimentally impacts left ventricular systolic function²¹. Establishing effective treatment strategies and refining diagnostic tools are critical endeavors. Former techniques like *in situ* and slot blot hybridization have fallen out of favor due to their tendency to yield false positive results. Instead, emphasis has shifted towards enhancing the value of EMB through amplification methods⁴³. Presently, viral etiologies of acute or chronic cardiomyopathies

are substantiated through viral genome analysis using quantitative PCR methods, including real-time PCR and nested PCR with reverse transcription. These techniques facilitate the detection of low copies of viral RNA or DNA sequences from endomyocardial samples^{22,44}, although it is important to note that conventional PCR might not detect mutant species²⁵. While a positive PCR outcome is diagnostic, corroborating viral serology from peripheral blood obtained at the time of EMB is crucial to exclude the possibility of passive blood contamination. However, a negative PCR result does not definitively rule out viral disease, as it may be influenced by sample size bias⁴³. The presence of cardiac autoantibodies indicates an immune-mediated process and highlights the suitability of immunosuppressive therapy³⁶. Utilizing PCR analysis, including assessing viral replication status and viral load, is advised for follow-up biopsies in patients with viral myocarditis to evaluate the efficacy of specific antiviral therapy^{36,43}. A key advantage of PCR analysis lies in its capability to detect both latent and active infections, addressing etiologies that may be more elusive using conventional methods. Furthermore, it enables strain typing, detection of virulence factors, and determination of antimicrobial resistance⁴⁵.

Biomarkers

Currently, there exists a consensus on the utility of nonspecific circulating biomarkers such as C-reactive protein (CRP) and erythrocyte sedimentation rate (ESR) to indicate the systemic inflammatory status during the acute phase of myocarditis. However, these biomarkers lack specificity in terms of diagnosis, prognosis, or risk stratification¹. In contrast, troponins are more specific indicators of myocardial damage, particularly in cases of acute coronary syndrome, and hold significant prognostic value³². B-type natriuretic peptide (BNP), synthesized from its precursor N-terminal pro b-type natriuretic peptide (NT-proBNP) under physiological conditions, is released into the bloodstream when systolic or diastolic dysfunction occurs. Nonetheless, these markers exhibit low negative predictive values^{5,32}. Throughout the three-phase progression of myocarditis, following the acute phase, a disrupted healing process leads to necrosis evolving into extensive fibrosis, clinically manifested as heart failure or sudden cardiac death^{1,22,25}. Myocardial fibrosis plays a pivotal role in the cardiac remodeling process, characterized by excessive stimulation of fibroblasts and augmented production of the extracellular matrix (ECM), coupled with cardiomyocyte apoptosis²⁹. At the cellular level, persistent cardiac injury, regardless of its origin, triggers the release of various substances that propel the progression toward fibrosis^{29,46,47}. Among the various cell types constituting the myocardium, cardio fibroblasts play a critical role in maintaining cardiac ECM homeostasis and remodeling. They contribute to processes like angiogenesis, cell proliferation, cardiomyocyte hypertrophy, and apoptosis^{47,48}. Importantly,

these cells can differentiate into myofibroblasts (myoFBs), which perpetuate the inflammatory response to injury by producing cytokines such as IL-1 α , IL-1 β , IL-6, IL-10, and tumor necrosis factor-alpha (TNF- α). Additionally, myoFBs exhibit an enhanced synthetic capacity for ECM proteins⁴⁹.

During the pathological process of ECM remodeling, the degradation of collagen fibers results in the cleavage of telopeptides located at the amino-terminal or carboxy-terminal ends of collagen molecules. This cleavage leads to the release of pro-peptides, specifically the amino-terminal propeptide of type I procollagen (PINP) and carboxy-terminal propeptide of type I procollagen (PICP) for collagen type I, and the amino-terminal propeptide of type III procollagen (PIIINP) and carboxy-terminal propertide of type III procollagen (PIIICP) for collagen type III. These pro-peptides are liberated during the biosynthesis of these collagen types in a balanced manner and are thus regarded as biomarkers of collagen synthesis. Conversely, biomarkers of collagen degradation are generated when collagen type I (CITP, NITP) and type III (CIIITP, NIIITP) are broken down, a process that occurs during ECM degradation⁴⁸. In a physiological context, collagen is stable and resistant to most proteinase activity, apart from matrix metalloproteinases (MMPs), which are implicated in cardiac remodeling. The resistance or cross-linking of collagen type I fibers is indicative of the slower cleavage of the carboxy-terminal telopeptide of collagen type I (CITP) by the enzyme MMP-1. This interplay results in the serum CITP: MMP-1 ratio displaying an inverse correlation with myocardial collagen cross-linking. Therefore, maintaining a balance between the action of MMPs and the activity of their inhibitors (TIMPs) is crucial in modulating myocardial fibrosis46,47,50.

Research efforts have concentrated on assessing the predictive significance of the three established biomarkers - PINP, PIIINP, and CITP: MMP-1 ratio - and their correlation with myocardial collagen fraction in patients afflicted with advanced-stage heart failure and dilated cardiomyopathy, which often arise as late complications of myocarditis^{47,50}. Transforming growth factor β (TGF- β), a signaling cytokine secreted by diverse cell types, including macrophages, assumes a pivotal role in various disease contexts^{51,52}. In the realm of cardiac pathology, it functions as a mediator of remodeling, prompting fibroblast differentiation and driving myocyte apoptosis.

TGF- β engages the TGF- β type II receptor to initiate the Drosophila gene "mothers against decapentaplegic" Caenorhabditis elegans small protein (SMAD)-signaling pathway. Subsequently, the formed protein complex traverses into the cell nucleus, functioning as a transcription factor that orchestrates genes implicated in apoptosis via the mitogen-activated protein kinase 8 pathway. The SMAD pathway is governed by feedback regulation. Markedly elevated circulating levels of TGF- β have been detected in patients with dilated cardiomyopathy^{46,51,52}. Furthermore, emerging evidence indicates that endoglin and fibulin 2, ECM proteins, function as co-receptors for TGF- β , exerting a profibrotic influence on cardiac fibroblasts and modulating TGF-B1's impact on extracellular matrix synthesis. Caveolins, a group of membrane proteins, are involved in receptor-independent endocytosis. In myocardial cells, caveolae-3 (Cav-3) serves as a scaffolding protein. Studies⁵³ in murine models have shown that the expression of these proteins is linked to the prognosis and survival rates. These markers have garnered attention due to their potential as therapeutic targets, not solely for diagnosing cardiac fibrosis^{46,54,55}. Galectin-3 (Gal-3), a soluble beta-galactosidase-binding lectin belonging to the galectin family, plays a pivotal role in host defense against injury and in the modulation of the transition towards chronic inflammation, ultimately favoring fibrosis. Extensive animal studies⁵⁶⁻⁵⁸ have illuminated Gal-3's involvement in myocardial fibrosis, triggering the activation of cardiac fibroblasts by modulating TGF-B's function. Gal-3's overexpression is mediated by the profibrotic influences of aldosterone and angiotensin II, mechanisms contributing to the progression of heart failure. While Gal-3 carries informative value, it is important to note that it may serve as a marker for confirming the cardiac inflammatory process and fibrosis, dependent on the etiology of heart failure. However, circulating Gal-3 concentrations do not appear to accurately reflect endomyocardial Gal-3 levels or cardiac fibrosis^{45,56}. Recent studies⁵⁹⁻⁶¹ have proposed several potential diagnostic or prognostic indicators in cardiovascular disease, although they do not specifically target the diagnosis of myocarditis. Among these, notable mentions include growth differentiation factor-15 (GDF-15) and pentraxin 3. GDF-15, also known as macrophage inhibitory cytokine-1, belongs to the TGF-β cytokine superfamily. In physiological contexts, it exhibits high expression in placental cells, and a more subdued

Elevated GDF-15 levels have been associated with inflammation and have been noted to be upregulated by factors like IL-1 β , TNF- α , and IL-2, particularly in peripheral tissues of patients with heart failure and dilated cardiomyopathy. Pentraxin 3 (PTX3), a soluble pattern recognition molecule, falls within the pentraxin superfamily alongside C-reactive protein. Its expression is induced by IL-1β, TNFa, and pathogenic compounds originating from cells such as macrophages, neutrophils, and fibroblasts. Conversely, high-density lipoproteins (HDL) and IL-10 act as down-regulators for PTX3. Endothelial cells contribute to PTX3 release, affording it a protective role. Operating through the regulation of complement activation, pathogen opsonization, innate immunity, and inflammation, PTX3 plays a crucial part in an extracellular matrix organization and cardiac remodeling facilitated by MMPs. Elevated circulating levels of PTX3 appear to correlate with severe heart failure and unfavorable outcomes, thereby positioning this molecule as a potential prognostic factor⁶²⁻⁶⁴. Suppression of tumorigenicity 2 protein (ST2), a member of the IL-1 receptors family, has emerged as a key player in inflammation and tissue fibrosis⁶⁵. ST2 acts through its ligand, IL-33, which activates a nuclear signal pathway, triggering an immunomodulatory response in tumor cells, as well as in heart and kidney cells. This interplay carries significant implications in these tissues, with potential consequences for inflammation and fibrosis processes⁶⁶. MicroRNAs The pursuit of identifying informative bio-

presence in cardiomyocytes and other cell types.

markers for the diagnosis and prognosis of various diseases, including the contentious area of acute myocarditis, has led to the discovery of novel RNA fractions associated with genetic, molecular processes. This journey was catalyzed by a seminal scientific breakthrough in the late 1980s involving the study of gene expressions of lin-4 and lin-14 in the temporal development of Caenorhabditis elegans larvae. This endeavor illuminated the existence of micro-RNAs (miRs), distinct genetic elements that do not encode proteins but instead interact with untranslated regions (5'UTR or 3'UTR) of messenger RNAs (mRNAs), thereby modulating the expression of specific genes in both physiological and pathological contexts⁶⁷⁻⁷². The prominence of miRs in the intracellular and extracellular milieu, coupled with their remarkable stability even under extreme conditions such as temperature or pH variations, has positioned them as desirable biomarkers for the diagnosis, progression, and prognosis of diverse diseases^{71,73}. Among cardiac tissue-specific miRs, certain molecules act as up-regulators (e.g., *miR208*) or down-regulators (e.g., miR126), and their roles have been more comprehensively elucidated in patients with coronary artery disease73. In the acute phase of myocarditis, as previously mentioned, Th17 cells contribute to the humoral immune response, and elevated levels of Th17-derived miRs have been observed, such as Mmu-miR-721 in murine models and has-miR-Chr8:96 in human myocarditis patients74,75. Several cardio-miRs are implicated in shaping the acute myocarditis process. MiR-21-5p and miR-208a, both connected with left ventricular systolic dysfunction, serve as prognostic indicators for recovery during the late stages of acute myocarditis^{76,77}. MiR-1-3p is associated with the extent of myocardial damage, mirroring troponin I levels⁷⁷. Further, the over-expression of miR-30a and miR-181d can alter the immune response to Coxsackievirus B3 by inhibiting SOCS-378. Conversely, a beneficial outcome was observed with the upregulation of both miR-155 and miR-148a. Increased miR-155 levels can effectively modulate the immune response to Coxsackievirus B3 by suppressing the nuclear signaling pathway, ultimately leading to improved survival rates among mice infected with the virus⁷⁹. Moreover, the diagnostic accuracy of acute myocarditis has been enhanced by the elevated expression of miR-146b, in correlation with cardiac troponin I, serving as an antigen (anti-cTNI)⁸⁰. Recent findings⁸¹ introduce a novel approach, suggesting a serum exosome miR panel for molecular diagnosis of myocarditis. Evaluating the circulating plasma levels of specific miRs, such as has-miR-30a, has-miR-192, has-miR-146a, has-miR-155, and has-miR-320a, collectively demonstrates higher diagnostic significance than individual miR assessment. This panel, especially characterized by has-miR-155 and has-miR-320a, offers superior differentiation between fulminant and non-fulminant myocarditis forms⁸¹. Beyond their diagnostic utility, cardiac miRs in acute myocarditis also hold prognostic significance and offer potential avenues for targeted treatment strategies. In summary, Table II provides an overview of the specificity and predictive value of biomarkers in the diagnosis of myocarditis.

Therapy

The diagnosis of myocarditis remains a complex challenge, and the treatment options are subject to ongoing debate. The diagnostic landscape continues to evolve as researchers explore various methods. Despite the gold standard status of EMB and PCR-related tests for diagnosing myocarditis, a significant number of cases within the pediatric population lack a clearly identified etiology. This underscores the need for improved diagnostic strategies that can provide more definitive answers. Moreover, the course of myocarditis can vary widely, with outcomes ranging from remission to unpredictable or unfavorable trajectories. This uncertainty underscores the importance of adopting a broad and supportive approach to management, especially given the lack of specific treatment options. As research advances and our understanding of myocarditis deepens, the hope is that more effective diagnostic methods and targeted therapeutic interventions will emerge, enhancing the care and outcomes of patients affected by this complex condition.

Symptoms-Based Therapy

In the absence of well-established treatment guidelines specifically tailored for pediatric patients with myocarditis, current strategies largely revolve around managing heart failure and arrhythmias, which are common manifestations of the condition. Beta-blockers and diuretics have shown promise in animal studies as beneficial interventions for managing heart failure in acute myocarditis patients⁸²⁻⁸⁴. Carvedilol, for instance, has demonstrated cardioprotective effects in autoimmune myocarditis by acting as an antioxidant and downregulating the synthesis of inflammatory cytokines⁸⁴. In terms of diuretics, torsemide has shown superiority over furosemide, primarily by improving left ventricular function and delaying the process of cardiac remodeling, which often leads to dilated cardiomyopathy⁸³. Arrhythmias are another significant concern in patients with suspected myocarditis, and they may manifest as sinus bradycardia, QRS complex prolongation, atrioventricular blocks, or tachyarrhythmias. Subacute phases of myocarditis can even present with ST-wave changes, further highlighting the electrical instability associated with the condition^{1,38}. The underlying mechanisms of arrhythmias can be multifactorial, encompassing structural changes due to viral invasion, ischemia secondary to endothelial tropism of viruses like

| Pathway | Biomarkers | Key Role | Specificity | Predictive value | References |
|------------------------------|--|---|-------------|------------------|-------------------------|
| Nonspecific | ESR CRP | Systemic inflammation | - | - | 1 |
| Specific | Troponin BNP, proBNP | Acute stress myocardial destruction Myocardial dysfunction | + + | + | 5,32 |
| Collagen | PINP, PIIICP CITP, NIIITP CITP: MMPs | Collagen synthesis Collagen degradation Collagen degradation | + + + | ++++++ | 47,82 |
| Fibrosis | TGF- β Fibulin -2 | Fibroblastic differentiation TGF β - co-receptor | - | ++++ | 46,51,52 |
| Inflammation and fibrosis | Galectin-3 Growth differentiation factor -15 PTX3 | IL-1β, TNF-α, IL-2- stimulation Complement opsonization | - - - | +++++++ | 45,56 59-61 62-64 |
| Gene expression | has-miR-Chr8:96 miR-21-5p, miR-208 a Panel has-miR-30a-has- miR-192-has- miR-146a-has- miR-155-has- miR-320a | Acute process Left ventricle dysfunction Fulminant vs. non-fulminant myocarditis | + + + | + + + | 75,77 76,80 81 |

| Table II. Specificity and predictive value of biomarkers in risk stratific | ation of myocarditis. |
|--|-----------------------|
|--|-----------------------|

(ESR) erythrocyte sedimentation rate; (CRP) C-reactive protein; (BNP) brain natriuretic peptide; (PINP) P-terminal propeptide of procollagen type II; (CITP) C-terminal propeptide of procollagen type III; (CITP) C-terminal propeptide of procollagen type III; (NIIITP) N-terminal propeptide of procollagen type III; (MMPs) Matrix metalloproteinases; (TGF- β) transforming growth factor beta; (TNF) tumor necrosis factor; (PTX3) Pentraxin 3.

Parvo B19, disruption of gap junctions and connexins, direct effects on calcium ion channels, and fibrotic scar formation⁸⁵. Given the persistent risk of arrhythmia development, even beyond the acute phase of the disease, antiarrhythmic medications are generally considered for these patients. Rigorous follow-up is crucial, particularly in the first year post-diagnosis. In cases where there is an increased risk of sudden death, consideration may be given to implanting a cardiac device or using temporary interventions like wearing a Life Vest to bridge transient cardiac rhythm disturbances^{1,22}. While there is a lack of standardized therapeutic protocols for pediatric myocarditis, these approaches aim to mitigate heart failure symptoms and manage arrhythmias, thereby improving patient outcomes.

Etiology Proven-Based Therapy

In pediatric populations, determining the precise etiology of myocarditis can indeed be

challenging, and reliance on peripheral blood serology is common due to the invasive nature of endomyocardial biopsy. However, when a viral cause is confirmed, targeted therapies can offer effective treatment options. For viral-induced myocarditis, interferon (IFN) β therapy has shown promising results. In cases of adenovirus or enterovirus-induced myocarditis, IF-N β therapy has been associated with enhanced viral clearance and improved left ventricular ejection fraction^{86,87}. It is important to note that the efficacy of IFN β therapy can vary depending on the specific viral agent causing the myocarditis. For instance, while high doses of IFN β have been found to improve endothelial function in Parvo B19-positive myocarditis cases, they may not lead to efficient viral clearance⁸⁸. In cases where myocarditis is caused by viruses like EBV, CMV, and HHV-6, antiviral drugs like acyclovir, ganciclovir, and valacyclovir can be considered as treatment options^{20,22,36,89}.

These medications target the specific viruses causing myocarditis and aim to reduce viral replication and associated inflammation. In addition to the mentioned therapies, other antiviral drugs like pleconaril and pocapavir have shown efficacy in decreasing cellular alterations in cases of enteroviral-induced myocarditis^{38,90-92}. These treatments are designed to directly impact the viral replication and associated effects on the heart tissue. It is important to emphasize that treatment decisions should be based on a comprehensive evaluation of the patient's clinical condition, the specific viral etiology, and potential benefits and risks associated with each therapy. As research continues to advance, more targeted and effective therapies may become available for the management of myocarditis in pediatric patients.

Pathogenesis-Based Therapy

Immunosuppression therapy, often involving prednisone or prednisolone, either alone or in combination with medications like azathioprine or cyclosporine, has shown promise in improving left ventricular ejection fraction in cases of virus-negative myocarditis^{1,25,93,94}. These therapies work by suppressing the immune response that contributes to the inflammatory process and cardiac damage. Intravenous immunoglobulin (IVIG) is indeed an important immunomodulatory treatment option. It contains a collection of antibodies obtained from healthy donors, which exert various effects on the immune system. The mechanisms through which IVIG exerts its therapeutic effects are complex and involve both fragment antigen-binding (Fab)-dependent and fragment crystallizable (Fc)-dependent pathways. Fab-dependent mechanisms involve interactions with B-cells and complement factors, leading to the suppression of autoreactive clones and inflammation^{3,95}. Fc-dependent mechanisms involve interactions with macrophages, regulatory T-cells, and other immune cells, resulting in the modulation of immune responses and suppression of inflammation^{3,96}. The multifactorial immunomodulatory effects of IVIG extend beyond its antiviral properties. It has been shown to neutralize pathogens and inhibit inflammatory cytokines, contributing to its anti-inflammatory effects. Recent data⁹⁷ from a Cochrane metanalysis further support the use of IVIG in patients with acute myocarditis, both in children and adults. This analy-

sis highlighted the beneficial effects of IVIG treatment, showing improved left ventricular ejection fraction in patients who received IVIG therapy compared to those who did not. For cases of fulminant myocarditis with a rapidly deteriorating clinical course, non-therapeutic options such as circulatory mechanical support and heart transplantation become crucial to providing life-saving interventions and addressing severe cardiac impairment. Overall, the approach of immunomodulation, including IVIG, represents an important therapeutic avenue in cases of non-viral proven or auto-reactive myocarditis, aiming to mitigate the immune response and its damaging effects on the heart tissue.

Futures Perspectives

These potential therapies offer innovative approaches to targeting specific pathways and mechanisms involved in the inflammatory and fibrotic processes associated with myocarditis. Immunoadsorption followed by intravenous immunoglobulin administration is an interesting approach to address autoantibodies, such as beta (1)-adrenergic receptor ($\beta(1)AR$) autoantibodies, that contribute to cardiac inflammation. By removing these autoantibodies and modulating the immune response with IVIG, this treatment strategy aims to reduce inflammation and improve left ventricular function^{36,98,99}. Aldosterone antagonists like eplerenone, which have been used in other cardiac conditions, are being explored for their potential benefits in cardiac remodeling by targeting mast cell gene expression and reducing inflammation¹⁰⁰. Targeting specific cytokines has also gained attention. Inhibiting IL-1ß with anti-mouse IL-1β antibodies has shown promise in reducing inflammation and fibrotic scar formation, potentially preventing aberrant cardiac remodelling¹⁰¹. Similarly, Secukinumab, an anti-IL-17 monoclonal antibody, is being considered to interrupt the profibrotic pathways induced by IL-17 in autoimmune cardiac impairment¹⁰². The use of antisense miRNA complements (antagomirs) as targeted therapy is a novel approach in autoimmune myocarditis. By administering antagomiR-21a-5p, researchers were able to attenuate myocardial inflammation and fibrosis in experimental models of induced myocarditis¹⁰³. This highlights the potential of miRNA-based therapies in modulating gene expression associated with pathological pro-

| Therapy | Target | Drug class/medication | Role | References |
|------------------------|---|---|--|----------------------------|
| Symptoms-based | Heart failure arrhythmias sudden death risk | Beta-blokers (Carvedilol) Diuretics (Torsemide) Anti-arrhythmics | Antioxidant cardiac remodeling | 84 83 1,85 |
| Aetiology proven-based | Parvo B-19, EBV, CMV, HHV-6 Enteroviruses | IFNβ Antivirals (Acyclovir, Ganciclovir,Valacyclovir) Pleconaril, Pocapavir | Improve viral clearance Improve left ventricular function | 20-22,36,86,89 38,90-92 |
| | SARS-CoV 2 | Remdesivir | | 23 |
| Pathogenesis-based | Immuno- modulation | Prednisone, Prednisolone IVIG | Improve left ventricular function | 1,25,93,94 97 |
| Perspectives | Inflammatory/ Fibrotic process | Immunoadsorbtion + IVIG fibrotic process Aldosterone antagonist (Eplerenone) | Cardiac remodelling IVIG | 36,98,99 100 |
| | | Anti-mouse IL-1β antibodies Anti-IL-17 monoclonal | | 101 102 |
| | | antibodies (Secukinumab) Ulinastatin + creatine phosphate sodium | | 104 |
| | | antisense miRNA complements (<i>antagomir-21a-5p</i>) | | 103 |

| Table III. Proposed therapy in AM | including accepted and p | erspective therapies. | aims and accomplishments. |
|-----------------------------------|--------------------------|-----------------------|---------------------------|
| | | | |

cesses. Ulinastatin, an acid-resistant protease inhibitor naturally present in human urine, has gained clinical recognition across various diseases due to its robust anti-inflammatory properties, making it a viable alternative to steroids. Recent research¹⁰⁴ has revealed its remarkable protective impact on cardiomyocytes, significantly improving cardiac function and reducing myocardial injury, especially when combined with creatine phosphate sodium. These findings hold promising implications for the treatment of viral myocarditis in pediatric patients. Ongoing human trials are crucial to validate the efficacy and safety of these emerging treatments and to further refine their therapeutic potential. These perspectives hold promise for providing more targeted and effective therapeutic options for patients with myocarditis, addressing the underlying immune and inflammatory mechanisms while aiming to improve cardiac function and patient outcomes. Table III summarizes the accepted treatment and prospects for future therapies in acute myocarditis.

Conclusions

The diagnosis of myocarditis involves various methods, with endomyocardial biopsy serving as the gold standard for confirmation. However, due to the risks involved, especially in pediatric patients, other diagnostic tools like immunohistochemistry and rt-PCR play a significant role in investigating viral etiology in myocardial samples. Biomarkers have emerged as promising candidates for both diagnostic and prognostic purposes, helping clinicians assess the inflammatory and cardiac remodeling processes. These markers can offer insights into disease severity, progression, and treatment response. Understanding the underlying pathogenesis of myocarditis is crucial for tailoring effective treatments. Different etiologies, such as viral infections or autoimmune reactions, drive distinct inflammatory and immune responses. Targeted therapies that focus on specific pathways or immune components associated with these etiologies have shown potential for better outcomes. Research efforts are ongoing to uncover more precise diagnostic methods, effective therapeutic interventions, and a deeper understanding of myocarditis complexities. This comprehensive approach will ultimately contribute to improved patient care, better treatment outcomes, and a more informed clinical management of this challenging cardiac condition.

Conflict of Interest

The authors declare that they have no conflict of interests.

Ethics Approval

Not applicable.

Informed Consent

Not applicable.

Funding

The review was carried out without funding.

Authors' Contribution

L. Agoston-Coldea conceptualized the manuscript's design. A. Popa and C. Lazea significantly contributed to the manuscript's conception; A. Popa performed the literature search and drafted the manuscript, L. Agoston-Coldea and C. Lazea supervised, verified the contents, and revised the manuscript. The authors reviewed and approved the final version of the article to be published.

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Data Availability

The authors confirm that the data supporting the findings of this study are available within the article.

References

 Caforio ALP, Pankuweit S, Arbustini E, Basso C, Gimeno-Blanes J, Felix SB, Fu M, Heliö T, Heymans S, Jahns R, Klingel K, Linhart A, Maisch B, McKenna W, Mogensen J, Pinto YM, Ristic A, Schultheiss HP, Seggewiss H, Tavazzi L, Thiene G, Yilmaz A, Charron P, Elliott PM. Current state of knowledge on aetiology, diagnosis, management, and therapy of myocarditis: A position statement of the European Society of Cardiology Working Group on Myocardial and Pericardial Diseases. Eur Heart J 2013; 34: 2636-2648d.

- Golpour A, Patriki D, Hanson PJ, McManus B, Heidecker B. Epidemiological Impact of Myocarditis. J Clin Med 2021; 10: 1-15.
- Di Filippo S. Improving outcomes of acute myocarditis in children. Expert Rev Cardiovasc Ther 2016; 14: 117-125.
- Butts RJ, Boyle GJ, Deshpande SR, Gambetta K, Knecht KR, Prada-Ruiz CA, Richmond ME, West SC, Lal AK. Characteristics of Clinically Diagnosed Pediatric Myocarditis in a Contemporary Multi-Center Cohort. Pediatr Cardiol 2017; 38: 1175-1182.
- Dasgupta S, Iannucci G, Mao C, Clabby M, Oster ME. Myocarditis in the pediatric population: A review. Congenit Heart Dis 2019; 14: 868-877.
- Cooper LT, Keren A, Sliwa K, Matsumori A, Mensah GA. The global burden of myocarditis: part 1: a systematic literature review for the Global Burden of Diseases, Injuries, and Risk Factors 2010 study. Glob Heart 2014; 9: 121-129.
- 7) Wang X, Bu X, Wei L, Liu J, Yang D, Mann DL, Ma A, Hayashi T. Global, Regional, and National Burden of Myocarditis From 1990 to 2017: A Systematic Analysis Based on the Global Burden of Disease Study 2017. Front Cardiovasc Med 2021; 8: 692990.
- Liu ZY, Su ZH, Li WC, Zhang FW, Ouyang WB, Wang SZ, Xia RB, Li YK, Pan XB. Global, regional, and national time trends in myocarditis-related mortality, 1990-2019: An age-period-cohort analysis. Eur Rev Med Pharmacol Sci 2023; 27: 9183-9191.
- 9) Bozkurt B, Colvin M, Cook J, Cooper LT, Deswal A, Fonarow GC, Francis GS, Lenihan D, Lewis EF, McNamara DM, Pahl E, Vasan RS, Ramasubbu K, Rasmusson K, Towbin JA, Yancy C. Current Diagnostic and Treatment Strategies for Specific Dilated Cardiomyopathies: A Scientific Statement From the American Heart Association. Circulation 2016; 134: e579-e646.
- Peretto G, Sala S, Rizzo S, Luca G De, Campochiaro C, Sartorelli S, Benedetti G, Palmisano A, Esposito A, Tresoldi M, Thiene G, Basso C, Bella P Della. Arrhythmias in myocarditis: State of the art. Heart Rhythm 2019; 16: 793-801.
- Ammirati E, Veronese G, Bottiroli M, Wang DW, Cipriani M, Garascia A, Pedrotti P, Adler ED, Frigerio M. Update on acute myocarditis. Trends Cardiovasc Med 2021; 31: 370-379.
- 12) Ammirati E, Kaski JP. Resident inflammatory cells in the myocardium of children: On the way to set histologic reference standards to differentiate normal myocardium from myocarditis. Int J Cardiol 2020; 303: 64-65.
- 13) Chen L, Deng H, Cui H, Fang J, Zuo Z, Deng J, Li Y, Wang X, Zhao L. Inflammatory responses and

inflammation-associated diseases in organs. Oncotarget 2017; 9: 7204-7218.

- 14) Guo H, Callaway JB, Ting JPY. Inflammasomes: mechanism of action, role in disease, and therapeutics. Nat Med 2015; 21: 677-687.
- Trachtenberg BH, Hare JM. Inflammatory Cardiomyopathic Syndromes. Circ Res 2017; 121: 803-818.
- Błyszczuk P. Myocarditis in Humans and in Experimental Animal Models. Front Cardiovasc Med 2019; 6: 64.
- Tunuguntla H, Jeewa A, Denfield SW. Acute Myocarditis and Pericarditis in Children. Pediatr Rev 2019; 40: 14-23.
- He Y, Hara H, Núñez G. Mechanism and Regulation of NLRP3 Inflammasome Activation. Trends Biochem Sci 2016; 41: 1012-1021.
- Heymans S, Eriksson U, Lehtonen J, Cooper LT. The Quest for New Approaches in Myocarditis and Inflammatory Cardiomyopathy. J Am Coll Cardiol 2016; 68: 2348-2364.
- Pollack A, Kontorovich AR, Fuster V, Dec GW. Viral myocarditis--diagnosis, treatment options, and current controversies. Nat Rev Cardiol 2015; 12: 670-680.
- Schultheiss HP, Baumeier C, Aleshcheva G, Bock CT, Escher F. Viral Myocarditis-From Pathophysiology to Treatment. J Clin Med 2021; 10: 5240.
- 22) Tschöpe C, Ammirati E, Bozkurt B, Caforio ALP, Cooper LT, Felix SB, Hare JM, Heidecker B, Heymans S, Hübner N, Kelle S, Klingel K, Maatz H, Parwani AS, Spillmann F, Starling RC, Tsutsui H, Seferovic P, Linthout S Van. Myocarditis and inflammatory cardiomyopathy: current evidence and future directions. Nat Rev Cardiol 2021; 18: 169-193.
- Lovell JP, Čiháková D, Gilotra NA. COVID-19 and Myocarditis: Review of Clinical Presentations, Pathogenesis and Management. Heart Int 2022; 16: 20-27.
- 24) Abbate A, Toldo S, Marchetti C, Kron J, Tassell BW Van, Dinarello CA. Interleukin-1 and the Inflammasome as Therapeutic Targets in Cardiovascular Disease. Circ Res 2020; 126: 1260-1280.
- Maisch B. Cardio-Immunology of Myocarditis: Focus on Immune Mechanisms and Treatment Options. Front Cardiovasc Med 2019; 6: 48.
- Bracamonte-Baran W, Čiháková D. Cardiac Autoimmunity: Myocarditis. Adv Exp Med Biol 2017; 1003: 187-221.
- 27) Liu X, Zhang Z, Ruan J, Pan Y, Magupalli VG, Wu H, Lieberman J. Inflammasome-activated gasdermin D causes pyroptosis by forming membrane pores. Nature 2016; 535: 153-158.
- Man SM, Karki R, Kanneganti TD. Molecular mechanisms and functions of pyroptosis, inflammatory caspases and inflammasomes in infectious diseases. Immunol Rev 2017; 277: 61-75.
- 29) Suthahar N, Meijers WC, Silljé HHW, Boer RA de. From Inflammation to Fibrosis-Molecular and Cellular Mechanisms of Myocardial Tissue Remodelling

and Perspectives on Differential Treatment Opportunities. Curr Heart Fail Rep 2017; 14: 235-250.

- 30) Singh K, Fang H, Davies G, Wright B, Lockstone H, Williams RO, Ciháková D, Knight JC, Bhattacharya S. Transcriptomic Analysis of Inflammatory Cardiomyopathy Identifies Molecular Signatures of Disease and Informs in silico Prediction of a Network-Based Rationale for Therapy. Front Immunol 2021; 12: 640837.
- 31) Wu L, Diny NL, Ong S, Barin JG, Hou X, Rose NR, Talor MV, Čiháková D. Pathogenic IL-23 signaling is required to initiate GM-CSF-driven autoimmune myocarditis in mice. Eur J Immunol 2016; 46: 582-592.
- 32) Dominguez F, Kühl U, Pieske B, Garcia-Pavia P, Tschöpe C. Update on Myocarditis and Inflammatory Cardiomyopathy: Reemergence of Endomyocardial Biopsy. Rev Esp Cardiol (Engl Ed) 2016; 69: 178-187.
- 33) Aretz HT, Billingham ME, Edwards WD, Factor SM, Fallon JT, Fenoglio JJ, Olsen EG, Schoen FJ. Myocarditis. A histopathologic definition and classification. Am J Cardiovasc Pathol 1987; 1: 3-14.
- 34) Katzmann JL, Schlattmann P, Rigopoulos AG, Noutsias E, Bigalke B, Pauschinger M, Tschope C, Sedding D, Schulze PC, Noutsias M. Meta-analysis on the immunohistological detection of inflammatory cardiomyopathy in endomyocardial biopsies. Heart Fail Rev 2020; 25: 277-294.
- 35) Kindermann I, Kindermann M, Kandolf R, Klingel K, Bültmann B, Müller T, Lindinger A, Böhm M. Predictors of outcome in patients with suspected myocarditis. Circulation 2008; 118: 639-648.
- 36) Caforio ALP, Marcolongo R, Jahns R, Fu M, Felix SB, Iliceto S. Immune-mediated and autoimmune myocarditis: clinical presentation, diagnosis and management. Heart Fail Rev 2013; 18: 715-732.
- 37) Liang JJ, Hebl VB, DeSimone CV, Madhavan M, Nanda S, Kapa S, Maleszewski JJ, Edwards WD, Reeder G, Cooper LT, Asirvatham SJ. Electrogram guidance: a method to increase the precision and diagnostic yield of endomyocardial biopsy for suspected cardiac sarcoidosis and myocarditis. JACC Heart Fail 2014; 2: 466-473.
- 38) Howard A, Hasan A, Brownlee J, Mehmood N, Ali M, Mehta S, Fergie J. Pediatric Myocarditis Protocol: An Algorithm for Early Identification and Management with Retrospective Analysis for Validation. Pediatr Cardiol 2020; 41: 316-326.
- 39) Seidel F, Opgen-Rhein B, Rentzsch A, Boehne M, Wannenmacher B, Boecker D, Reineker K, Grafmann M, Wiegand G, Hecht T, Kiski D, Fischer M, Papakostas K, Ruf B, Kramp J, Khalil M, Kaestner M, Steinmetz M, Fischer G, Özcan S, Freudenthal N, Schweigmann U, Hellwig R, Pickardt T, Klingel K, Messroghli D, Schubert S. Clinical characteristics and outcome of biopsy-proven myocarditis in children - Results of the German prospective multicentre registry 'MYKKE'. Int J Cardiol 2022; 357: 95-104.

- 40) Mills KI, Vincent JA, Zuckerman WA, Hoffman TM, Canter CE, Marshall AC, Blume ED, Bergersen L, Daly KP. Is Endomyocardial Biopsy a Safe and Useful Procedure in Children with Suspected Cardiomyopathy? Pediatr Cardiol 2016; 37: 1200-1210.
- 41) Brighenti M, Donti A, Giulia Gagliardi M, Maschietto N, Marini D, Lombardi M, Vairo U, Agnoletti G, Milanesi O, Pongiglione G, Bonvicini M. Endomyocardial biopsy safety and clinical yield in pediatric myocarditis: An Italian perspective. Catheter Cardiovasc Interv 2016; 87: 762-767.
- 42) Carturan E, Milanesi O, Kato Y, Giacometti C, Biffanti R, Thiene G, Calabrese F. Viral detection and tumor necrosis factor alpha profile in tracheal aspirates from children with suspicion of myocarditis. Diagn Mol Pathol 2008; 17: 21-27.
- 43) Basso C, Calabrese F, Angelini A, Carturan E, Thiene G. Classification and histological, immunohistochemical, and molecular diagnosis of inflammatory myocardial disease. Heart Fail Rev 2013; 18: 673-681.
- 44) Nguyen Y, Renois F, Leveque N, Giusti D, Picard-Maureau M, Bruneval P, Fornes P, Andreoletti L. Virus detection and semiquantitation in explanted heart tissues of idiopathic dilated cardiomyopathy adult patients by use of PCR coupled with mass spectrometry analysis. J Clin Microbiol 2013; 51: 2288-2294.
- Calabrese F, Thiene G. Myocarditis and inflammatory cardiomyopathy: microbiological and molecular biological aspects. Cardiovasc Res 2003; 60: 11-25.
- 46) An Z, Yang G, Zheng H, Nie W, Liu G. Biomarkers in patients with myocardial fibrosis. Open Life Sci 2017; 12: 337-344.
- 47) González A, Schelbert EB, Díez J, Butler J. Myocardial Interstitial Fibrosis in Heart Failure: Biological and Translational Perspectives. J Am Coll Cardiol 2018; 71: 1696-1706.
- 48) Fan D, Takawale A, Lee J, Kassiri Z. Cardiac fibroblasts, fibrosis and extracellular matrix remodeling in heart disease. Fibrogenesis Tissue Repair 2012; 5: 15.
- 49) Baum J, Duffy HS. Fibroblasts and myofibroblasts: what are we talking about? J Cardiovasc Pharmacol 2011; 57: 376-379.
- 50) López B, Ravassa S, González A, Zubillaga E, Bonavila C, Bergés M, Echegaray K, Beaumont J, Moreno MU, San José G, Larman M, Querejeta R, Díez J. Myocardial Collagen Cross-Linking Is Associated With Heart Failure Hospitalization in Patients With Hypertensive Heart Failure. J Am Coll Cardiol 2016; 67: 251-260.
- 51) Khan S, Joyce J, Margulies KB, Tsuda T. Enhanced bioactive myocardial transforming growth factor-β in advanced human heart failure. Circ J 2014; 78: 2711-2718.
- 52) Komai T, Inoue M, Okamura T, Morita K, Iwasaki Y, Sumitomo S, Shoda H, Yamamoto K, Fujio K. Transforming Growth Factor-β and Interleukin-10 Synergistically Regulate Humoral Immuni-

ty via Modulating Metabolic Signals. Front Immunol 2018; 9: 1364.

- 53) Jia HL, Liu J. Investigation into the correlations of expressions of Cav-3 and Smad3 with pathogenesis and prognosis of viral myocarditis. Eur Rev Med Pharmacol Sci 2017; 21: 3262-3269.
- 54) Khan SA, Dong H, Joyce J, Sasaki T, Chu ML, Tsuda T. Fibulin-2 is essential for angiotensin II-induced myocardial fibrosis mediated by transforming growth factor (TGF)-β. Lab Invest 2016; 96: 773-783.
- 55) Shyu KG. The Role of Endoglin in Myocardial Fibrosis. Acta Cardiol Sin 2017; 33: 461-467.
- 56) Besler C, Lang D, Urban D, Rommel KP, Roeder M Von, Fengler K, Blazek S, Kandolf R, Klingel K, Thiele H, Linke A, Schuler G, Adams V, Lurz P. Plasma and Cardiac Galectin-3 in Patients With Heart Failure Reflects Both Inflammation and Fibrosis: Implications for Its Use as a Biomarker. Circ Heart Fail 2017; 10: e003804.
- 57) Noguchi K, Tomita H, Kanayama T, Niwa A, Hatano Y, Hoshi M, Sugie S, Okada H, Niwa M, Hara A. Time-course analysis of cardiac and serum galectin-3 in viral myocarditis after an encephalomyocarditis virus inoculation. PLoS One 2019; 14: e0210971.
- 58) Suthahar N, Meijers WC, Silljé HHW, Ho JE, Liu FT, Boer RA de. Galectin-3 Activation and Inhibition in Heart Failure and Cardiovascular Disease: An Update. Theranostics 2018; 8: 593-609.
- 59) Lok SI, Winkens B, Goldschmeding R, Geffen AJP Van, Nous FMA, Kuik J Van, Weide P Van Der, Klöpping C, Kirkels JH, Lahpor JR, Doevendans PA, Jonge N De, Weger RA De. Circulating growth differentiation factor-15 correlates with myocardial fibrosis in patients with non-ischaemic dilated cardiomyopathy and decreases rapidly after left ventricular assist device support. Eur J Heart Fail 2012; 14: 1249-1256.
- Wischhusen J, Melero I, Fridman WH. Growth/ Differentiation Factor-15 (GDF-15): From Biomarker to Novel Targetable Immune Checkpoint. Front Immunol 2020; 11: 951.
- Wollert KC, Kempf T, Wallentin L. Growth Differentiation Factor 15 as a Biomarker in Cardiovascular Disease. Clin Chem 2017; 63: 140-151.
- 62) Doni A, Stravalaci M, Inforzato A, Magrini E, Mantovani A, Garlanda C, Bottazzi B. The Long Pentraxin PTX3 as a Link Between Innate Immunity, Tissue Remodeling, and Cancer. Front Immunol 2019; 10: 712.
- Ristagno G, Fumagalli F, Bottazzi B, Mantovani A, Olivari D, Novelli D, Latini R. Pentraxin 3 in Cardiovascular Disease. Front Immunol 2019; 10: 823.
- Zlibut A, Bocsan IC, Agoston-Coldea L. Pentraxin-3 and endothelial dysfunction. Adv Clin Chem 2019; 91: 163-179.
- 65) AbouEzzeddine OF, McKie PM, Dunlay SM, Stevens SR, Felker GM, Borlaug BA, Chen HH, Tra-

cy RP, Braunwald E, Redfield MM. Suppression of Tumorigenicity 2 in Heart Failure With Preserved Ejection Fraction. J Am Heart Assoc 2017; 6: e004382.

- 66) Homsak E, Gruson D. Soluble ST2: A complex and diverse role in several diseases. Clin Chim Acta 2020; 507: 75-87.
- 67) Ambros V, Horvitz HR. The lin-14 locus of Caenorhabditis elegans controls the time of expression of specific postembryonic developmental events. Genes Dev 1987; 1: 398-414.
- Chalfie M, Horvitz HR, Sulston JE. Mutations that lead to reiterations in the cell lineages of C. elegans. Cell 1981; 24: 59-69.
- 69) Ferguson EL, Sternberg PW, Horvitz HR. A genetic pathway for the specification of the vulval cell lineages of Caenorhabditis elegans. Nature 1987; 326: 259-267.
- 70) Horvitz HR, Sulston JE. Isolation and genetic characterization of cell-lineage mutants of the nematode Caenorhabditis elegans. Genetics 1980; 96: 435-454.
- 71) O'Brien J, Hayder H, Zayed Y, Peng C. Overview of MicroRNA Biogenesis, Mechanisms of Actions, and Circulation. Front Endocrinol (Lausanne) 2018; 9: 402.
- 72) Ying SY, Chang DC, Lin SL. The MicroRNA. Methods Mol Biol 2018; 1733: 1-25.
- Small EM, Frost RJA, Olson EN. MicroRNAs add a new dimension to cardiovascular disease. Circulation 2010; 121: 1022-1032.
- 74) Blanco-Domínguez R, Sánchez-Díaz R, la Fuente H de, Jiménez-Borreguero LJ, Matesanz-Marín A, Relaño M, Jiménez-Alejandre R, Linillos-Pradillo B, Tsilingiri K, Martín-Mariscal ML, Alonso-Herranz L, Moreno G, Martín-Asenjo R, García-Guimaraes MM, Bruno KA, Dauden E, González-Álvaro I, Villar-Guimerans LM, Martínez-León A, Salvador-Garicano AM, Michelhaugh SA, Ibrahim NE, Januzzi JL, Kott-witz J, Iliceto S, Plebani M, Basso C, Baritussio A, Seguso M, Marcolongo R, Ricote M, Fairweather D, Bueno H, Fernández-Friera L, Alfonso F, Caforio ALP, Pascual-Figal DA, Heidecker B, Lüscher TF, Das S, Fuster V, Ibáñez B, Sánchez-Madrid F, Martín P. A Novel Circulating MicroRNA for the Detection of Acute Myocarditis. N Engl J Med 2021; 384: 2014-2027.
- 75) Düsing P, Zietzer A, Jansen F. MicroRNA-Based Diagnostics in Heart Diseases: Current Limitations and Future Perspectives. JACC Basic Transl Sci 2021; 6: 897-899.
- 76) Goldberg L, Tirosh-Wagner T, Vardi A, Abbas H, Pillar N, Shomron N, Nevo-Caspi Y, Paret G. Circulating MicroRNAs: a Potential Biomarker for Cardiac Damage, Inflammatory Response, and Left Ventricular Function Recovery in Pediatric Viral Myocarditis. J Cardiovasc Transl Res 2018; 11: 319-328.
- 77) Marketou M, Kontaraki J, Patrianakos A, Kochiadakis G, Anastasiou I, Fragkiadakis K, Plevritaki A, Papadaki ST, Chlouverakis G, Parthenakis

F. Peripheral Blood MicroRNAs as Potential Biomarkers of Myocardial Damage in Acute Viral Myocarditis. Genes (Basel) 2021; 12: 420.

- 78) Fan KL, Li MF, Cui F, Feng F, Kong L, Zhang FH, Hao H, Yin MX, Liu Y. Altered exosomal miR-181d and miR-30a related to the pathogenesis of CVB3 induced myocarditis by targeting SOCS3. Eur Rev Med Pharmacol Sci 2019; 23: 2208-2215.
- 79) Bao JL, Lin L. MiR-155 and miR-148a reduce cardiac injury by inhibiting NF-κB pathway during acute viral myocarditis. Eur Rev Med Pharmacol Sci 2014; 18: 2349-2356.
- 81) Zhang Y, Li X, Wang D, Jiang X, Zhang M, Lv K. Serum exosome microRNA panel as a noninvasive biomarker for molecular diagnosis of fulminant myocarditis. Mol Ther Methods Clin Dev 2020; 20: 142-151.
- 82) López B, González A, Ravassa S, Beaumont J, Moreno MU, San José G, Querejeta R, Díez J. Circulating Biomarkers of Myocardial Fibrosis: The Need for a Reappraisal. J Am Coll Cardiol 2015; 65: 2449-2456.
- 83) Veeraveedu PT, Watanabe K, Ma M, Thandavarayan RA, Palaniyandi SS, Yamaguchi K, Suzuki K, Kodama M, Aizawa Y. Comparative effects of torasemide and furosemide in rats with heart failure. Biochem Pharmacol 2008; 75: 649-659.
- 84) Yuan Z, Shioji K, Kihara Y, Takenaka H, Onozawa Y, Kishimoto C. Cardioprotective effects of carvedilol on acute autoimmune myocarditis: anti-inflammatory effects associated with antioxidant property. Am J Physiol Heart Circ Physiol 2004; 286: H83-H90.
- 85) Steinke K, Sachse F, Ettischer N, Strutz-Seebohm N, Henrion U, Rohrbeck M, Klosowski R, Wolters D, Brunner S, Franz WM, Pott L, Munoz C, Kandolf R, Schulze-Bahr E, Lang F, Klingel K, Seebohm G. Coxsackievirus B3 modulates cardiac ion channels. FASEB J 2013; 27: 4108-4121.
- 86) Kühl U, Pauschinger M, Schwimmbeck PL, Seeberg B, Lober C, Noutsias M, Poller W, Schultheiss HP. Interferon-beta treatment eliminates cardiotropic viruses and improves left ventricular function in patients with myocardial persistence of viral genomes and left ventricular dysfunction. Circulation 2003; 107: 2793-2798.
- 87) Schultheiss HP, Piper C, Sowade O, Waagstein F, Kapp JF, Wegscheider K, Groetzbach G, Pauschinger M, Escher F, Arbustini E, Siedentop H, Kuehl U. Betaferon in chronic viral cardiomyopathy (BICC) trial: Effects of interferon-β treatment in patients with chronic viral cardiomyopathy. Clin Res Cardiol 2016; 105: 763-773.
- 88) Schmidt-Lucke C, Spillmann F, Bock T, Kühl U, Linthout S Van, Schultheiss HP, Tschöpe C. Interferon beta modulates endothelial damage in patients with cardiac persistence of human parvovirus b19 infection. J Infect Dis 2010; 201: 936-945.
- 89) Jensen LD, Marchant DJ. Emerging pharmacologic targets and treatments for myocarditis. Pharmacol Ther 2016; 161: 40-51.

- 90) Abzug MJ, Michaels MG, Wald E, Jacobs RF, Romero JR, Sánchez PJ, Wilson G, Krogstad P, Storch GA, Lawrence R, Shelton M, Palmer A, Robinson J, Dennehy P, Sood SK, Cloud G, Jester P, Acosta EP, Whitley R, Kimberlin D. A Randomized, Double-Blind, Placebo-Controlled Trial of Pleconaril for the Treatment of Neonates With Enterovirus Sepsis. J Pediatric Infect Dis Soc 2016; 5: 53-62.
- 91) Amdani SM, Kim HS, Orvedahl A, John AO, Said A, Simpson K. Successful treatment of fulminant neonatal enteroviral myocarditis in monochorionic diamniotic twins with cardiopulmonary support, intravenous immunoglobulin and pocapavir. BMJ Case Rep 2018; 2018: bcr2017224133.
- 92) Yen MH, Huang YC, Chen MC, Liu CC, Chiu NC, Lien R, Chang LY, Chiu CH, Tsao KC, Lin TY. Effect of intravenous immunoglobulin for neonates with severe enteroviral infections with emphasis on the timing of administration. J Clin Virol 2015; 64: 92-96.
- 93) Blagova O, Nedostup A, Kogan E, Zaitsev A, Fomin V. Immunosuppressive therapy of biopsy proved immune-mediated lymphocytic myocarditis in the virus-negative and virus-positive patients. Cardiovasc Pathol 2020; 49: 107260.
- 94) Frustaci A, Russo MA, Chimenti C. Randomized study on the efficacy of immunosuppressive therapy in patients with virus-negative inflammatory cardiomyopathy: the TIMIC study. Eur Heart J 2009; 30: 1995-2002.
- 95) Gelfand EW. Intravenous immune globulin in autoimmune and inflammatory diseases. N Engl J Med 2012; 367: 2015-2025.
- 96) Maddur MS, Othy S, Hegde P, Vani J, Lacroix-Desmazes S, Bayry J, Kaveri S V. Immunomodulation by intravenous immunoglobulin: role of regulatory T cells. J Clin Immunol 2010; 30: S4-S8.
- 97) Huang X, Sun Y, Su G, Li Y, Shuai X. Intravenous Immunoglobulin Therapy for Acute Myocarditis in

Children and Adults. Int Heart J 2019; 60: 359-365.

- 98) Mobini R, Staudt A, Felix SB, Baumann G, Wallukat G, Deinum J, Svensson H, Hjalmarson Å, Fu M. Hemodynamic improvement and removal of autoantibodies against β1-adrenergic receptor by immunoadsorption therapy in dilated cardiomyopathy. J Autoimmun 2003; 20: 345-350.
- 99) Staudt A, Schäper F, Stangl V, Plagemann A, Böhm M, Merkel K, Wallukat G, Wernecke KD, Stangl K, Baumann G, Felix SB. Immunohistological changes in dilated cardiomyopathy induced by immunoadsorption therapy and subsequent immunoglobulin substitution. Circulation 2001; 103: 2681-2686.
- 100) Xiao J, Shimada M, Liu W, Hu D, Matsumori A. Anti-inflammatory effects of eplerenone on viral myocarditis. Eur J Heart Fail 2009; 11: 349-353.
- 101) Kraft L, Erdenesukh T, Sauter M, Tschöpe C, Klingel K. Blocking the IL-1β signalling pathway prevents chronic viral myocarditis and cardiac remodeling. Basic Res Cardiol 2019; 114: 11.
- 102) Stebut E von, Boehncke WH, Ghoreschi K, Gori T, Kaya Z, Thaci D, Schäffler A. IL-17A in Psoriasis and Beyond: Cardiovascular and Metabolic Implications. Front Immunol 2020; 10: 3096.
- 103) Mirna M, Paar V, Topf A, Kraus T, Sotlar K, Aigner A, Ewe A, Watzinger S, Podesser BK, Hackl M, Pistulli R, Hoppe UC, Kiss A, Lichtenauer M. A new player in the game: treatment with antagomiR-21a-5p significantly attenuates histological and echocardiographic effects of experimental autoimmune myocarditis. Cardiovasc Res 2022; 118: 556-572.
- 104) Li CL, Jia LB, Gao J, Wang ZZ, An XJ. The efficacy observation of ulinastatin combined with creatine phosphate sodium in pediatric viral myocarditis. Eur Rev Med Pharmacol Sci 2019; 23: 7144-7151.