

USP22 down-regulation facilitates human retinoblastoma cell aging and apoptosis via inhibiting TERT/P53 pathway

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Abstract. – OBJECTIVE: Retinoblastoma is the most common malignant intraocular tumor in childhood, and still lacks effective treatment. The immortality of tumor cell can be attributed to elevated telomerase activity, which has been considered as tumor marker and treatment target. USP22 is one of the important targets for inhibiting tumor growth, but clear illustration regarding its effects of telomerase, tumor cell immortality and retinoblastoma cell aging or apoptosis via suppressing TERT/P53 pathway remains to be elusive.

MATERIALS AND METHODS: MTT was used for describing cell proliferation. Western blot was used to test protein expression level of USP22, TERT and P53. RT-qPCR was used to test USP22 mRNA level, followed by TRAP method to detect telomerase activity. Flow cytometry and comet assay were used to quantify cell apoptosis and DNA damage. Cell aging was measured by β -galactosidase.

RESULTS: The overexpression of USP22 significantly enhanced cell proliferation potency and telomerase activity. Elevated TERT expression level, inhibited P53 expression and cell aging, as well as reduced cell apoptosis or DNA damage. Down-regulation of USP22 contributed to opposite effects.

CONCLUSIONS: USP22 played an important role in retinoblastoma cell proliferation/aging and apoptosis. The reduction of USP22 expression facilitated human retinoblastoma cell aging or apoptosis via suppressing TERT/P53 signal pathway. USP22, thus, may work as a target for treating retinoblastoma.

Key Words:

USP22, Retinoblastoma, Telomerase, TERT/P53.

Introduction

Retinoblastoma is a kind of malignant tumor derived from embryonic retinoblastoma, and is the most common primary intraocular cancer associated with family inheritance. Currently, major treatment approaches for retinoblastoma include surgical resection of the eyeball or conservative treatment, although both efficacy and prognosis need to be further improved. Tumor cell immortalization is one of the most distinguished features apart from normal cells. The possible reason is elevated telomerase activity, which has been realized as a tumor marker and satisfactory target of tumor treatment¹. Telomere is a protein-DNA complex at chromosome ending, and can be shortened with repeated mitosis under normal circumstance to induce cell aging^{2,3}. The inhibition of telomerase activity can impede tumor cell immortalization. Moreover, differential expression of telomerase between cancer cell and normal cell could reduce toxic effects of drugs^{4,5}. Ubiquitin specific proteinase 22 (USP22) belongs to novel human de-ubiquitination gene with the function of predicting human cancer metastatic potency and treatment outcomes⁶. Previous studies^{7,8} showed that abnormal expression of USP22 was correlated with prognosis of infiltrative breast cancer or colorectal carcinoma. USP22 can modulate various cellular functions including cell growth/differentiation and cycle regulation, and is correlated with cellular gene transcriptional activation and signal transduction⁹⁻¹¹. Another study¹² showed that the

silence of USP22 gene inhibited prostate cancer cell proliferation and induced cell cycle arrest. USP22 depletion also facilitated human brain glioma cell apoptosis¹³. USP22, as an important target of tumor growth inhibition, however, requires to be further studied regarding its role in facilitating human retinoblastoma cell aging or apoptosis via suppressing TERT/P53 signal pathway. This study aimed to observe the effect of USP22 on TERT/P53 signal pathway, and its effects on retinoblastoma cell DNA damage, apoptosis and telomerase activity. The knowledge of USP22 in tumor cell aging and apoptosis may provide evidence for clinical exploration of novel drug target against intraocular tumors.

Materials and Methods

Reagents and Drugs

MTT reagent was purchased from Sigma-Aldrich (St. Louis, MO, USA). One-step RT-qPCR kit was purchased from Quanshijin Bio (Beijing, China). Dulbecco's Modified Eagle Medium (DMEM), fetal bovine serum (FBS) and other antibiotics were purchased from Gibco (Rockville, MD, USA). Human gastric cancer cell line MGC-803 was purchased from Cell Bank of Chinese Academy of Sciences (Beijing, China). Anti-rabbit CDKL1 polyclonal antibody was purchased from Abcam (Cambridge, MA, USA). Goat anti-rabbit IgG (H+L) and secondary antibodies were purchased from Bointec (Wuhan, Hubei, China). Lentiviral transfection vector was purchased from Jikai Gene (Shanghai, China). Annexin V-Propidium iodide (AV-PI) staining kit was purchased from Beyotime (Shanghai, China). Telomerase activity assay kit was purchased from Kaiji Biotech (Shanghai, China). TUNEL assay kit was purchased from Beyotime (Shanghai, China). AV double staining kit was purchased from Beyotime (Shanghai, China). 8-OHdG assay kit was purchased from Huamei (Luoyang, Henan, China). HEK293T cell was purchased from Cell Bank of Chinese Academy of Sciences (Beijing, China). Lipofectamine 2000 transfection reagent was purchased from Invitrogen (Carlsbad, CA, USA).

Cell Culture

Human retinoblastoma cell line HXO-Rb⁴⁴ was purchased from the Tumor Institute, Harbin Medical University. Cells were cultured in 1640 medium containing 10% fetal bovine se-

rum (FBS), 100 U/mL penicillin and 100 µg/mL streptomycin in a 37°C incubator with 5% CO₂ in suspension. Cell morphology and growth status were observed under an inverted phase contrasted microscope. Those cells at log-growth phase were used for experiments.

Lentiviral Transfection

Lentiviral vector for USP22 knockdown or over-expression was synthesized based on CDKL1 gene sequence (Jikai Gene, Shanghai, China), and was sub-cloned into pGCS-GFP plasmid with Age I/EcoR1 sites. When cell density reached 80-80%, Lipofectamine 2000 incorporated lentiviral particles was used to transfect HEK293T cells. 72 h later, USP22 knockdown and over-expression lentivirus were harvested from culture medium, enriched by Centricon Plus-20 ultrapure centrifuge, and were kept at -80°C. Retinoblastoma cells were inoculated in 6-well plate. When reaching 80% confluence, lentivirus was added based on viral titer. On the next day, normal culture medium was used for observing transfection efficiency under fluorescent microscope.

MTT Assay

Retinoblastoma cells at log-growth phase were adjusted and inoculated into 96-well plate (100 µL each well). After transfection by lentiviral vector for USP22 knockdown or over expression for 6 h, the plate was centrifuged, and normal medium was added for 18 h incubation. Each group included 8 paralleled replicated wells. After 24 h incubation, each well was added with 10 µL MTT solution for 4 h continuous incubation. Under microscopic observation, violet crystal was formed. The supernatant was discarded, with addition of 150 µL triple buffer into each well for mix well to solve the crystal. After 12-15 h, absorbent values were measured at 490 nm for calculating cell viability.

Western Blot

Retinoblastoma cells after transfection were centrifuged and were mixed with 100 µL lysis buffer (including proteinase inhibitor, phenylmethylsulfonyl fluoride (PMSF), phosphatase inhibitor). After homogenization, the mixture was centrifuged at 14000 g for 10 min. The supernatant was collected to obtain the whole protein solution. After adding equal volume of loading buffer, the mixture was boiled for 5 min for denaturing. The sample was loaded into sodium dodecyl sulphate-polyacrylamide gel electrophoresis (SDS-PAGE) and was transferred to polyvinylidene fluoride (PVDF)

membrane, which was blocked in defatted milk powder for 1 h. Antibody against USP22 (1:1000), p53 (1:1000), TERT (1:1000) and GAPDH (1:1000) was used for hybridization at 4°C overnight. After Tris-buffered saline (TBS) rinsing for three times, secondary antibody diluted at 1:1000 was added for 37°C incubation for 1 h. ECL development was then performed.

Total cell RNA Extraction and Real-time qPCR

All cells were cultured for 48 h after transfection. Culture medium was removed, and cells were washed with phosphate-buffered saline (PBS) for three times. 1 mL Trizol was added to each well for 5 min iced incubation. Pipette tip was used for homogenization. Lysate from each well was collected and mixed with 200 μ L chloroform for 15 s rough mixture. After room temperature incubation for 3 min, the mixture was centrifuged at 12000 g at 4°C for 15 min. The upper aqueous phase was carefully transferred and mixed with 500 μ L isopropanol for 10 min room temperature incubation. By 4°C centrifugation at 12000 g, the supernatant was discarded, and triple washing in 1 mL ethanol. The supernatant was carefully removed eventually, and mRNA was re-suspended in 20 μ L DEPC water. USP22 primers were synthesized by Signa Genes (St. Louis, MO, USA)^{14,15}. Sequences were forward primer, 5'-ATCTC CGTCC TACG TCG TCG-3'; reverse primer, 5'-ATCTC CCTC CTA TCA TCA TCA-3'. GAPDH was used as internal reference (forward primer, 5'-TGACT TAC AGC TAC CACCC A-3'; reverse primer, 5'-GCC TGT TTT GTGTA GCCAA A-3'). One-step RT-PCR was performed in a 50 μ L system following manual instruction. Reaction conditions were: 50°C for 30 min and 95°C for 5 min, followed by 40 cycles each containing 58°C 30 s, 55°C 30 s and 72°C 50 s, with ending 72°C elongation for 5 min. After amplification, amplifying curve and melting curve of RT-PCR were confirmed. Ct values of target gene were compared against Ct values of internal reference gene. Gene expression was then quantified by $2^{-\Delta\Delta C_t}$ method.

Cell Apoptosis Assay

Cell apoptosis was tested by AV-PI staining. Cells were inoculated into 6-well plate. After transfection using lentivirus, cells were centrifuged and changed for normal culture medium. AV and PI staining buffer were sequentially added for iced dark incubation in 10 min. Cells were

then loaded into FACS Calibur flow cytometry (BD, San Jose, CA, USA) for online testing. All experiments were repeated for three times.

Telomerase Activity Assay

This study utilized telomere repeated amplification method to test telomerase activity¹⁶. In brief, cells were precipitated in CHAPS buffer containing nuclease inhibitor for 1 h at 4°C for 30 min. Cell extracts (1 mg) were mixed with telomerase substrate (TS) primer, GAPDH primer and Maxima SYBR green qPCR Master Mix (TRIPeZe mixture). TS primer was elongated at 37°C for 30 min, followed by PCR amplification. PCR products from the same cell extract were analyzed for telomerase activity using $2^{-\Delta\Delta C_t}$ method.

Comet Assay

Comet assay kit was purchased from Kaiji (Shanghai, China). The assay was paved following manual instruction, with the addition of gel containing single cell suspension. When the gel was formed, the coverslip was carefully removed. The glass slide was immersed in pre-cold cell lysis buffer for 1 h incubation. After balancing in PBS, the slide was placed in horizontal electrophoresis tank filled with alkaline electrophoresis buffer. The mixture was denatured in the dark for 30 min, and was separated under 25 V, 300 mA current for 25 min. EB staining dye was added followed by coverslips, and observation under fluorescent microscope.

β -galactosidase Staining

Cellular β -galactosidase staining kit was purchased from Kaiji (Shanghai, China). Cells were collected by centrifugation. After rinsing in phosphate-buffered saline (PBS), 1 mL β -galactosidase fixation buffer was added for room temperature incubation for 10 min. Cell fixation buffer was removed by centrifugation, and cells were washed with phosphate-buffered saline (PBS) for three times (3 min each). The supernatant was discarded, and each tube was added with 0.5 mL staining working solution for 37°C overnight incubation. On the second day, staining cells were added onto glass slides for observation under light field microscope.

8-OHdG Assay

Intracellular 8-OHdG assay was performed using enzyme-linked immunosorbent assay. The kit was provided by Xitang Biotech (Shanghai, China) following manual instruction.

Statistical Analysis

Experimental data were obtained from at least three independent experiments and data were presented as mean±standard deviation (SD). Comparison between two groups was performed by student *t*-test. Comparison among multiple groups was performed by analysis of variance (ANOVA), followed by SNK approach between two groups. Statistical significance was defined when $p < 0.05$.

Results

Change of USP22 RNA Level

After infection by recombinant lentivirus of over-expression or knockdown of USP22, qPCR was used to test USP22 RNA expression level. As shown in Figure 1, over-expression lentivirus vector significantly up-regulated USP22 expression, whilst USP22 knockdown lentivirus remarkably suppressed USP22 RNA expression, with significant difference compared to blank control group ($p < 0.01$). These results proved that recombinant lentivirus vectors were successfully constructed and effectively enhanced or suppressed USP22 RNA level.

USP22, TERT and P53 Protein Expression Level

We then used Western blotting to measure USP22, TERT and P53 protein expression. After infected by USP22 over-expression lentivirus, USP22 and TERT protein levels were significantly elevated, meanwhile, expression of

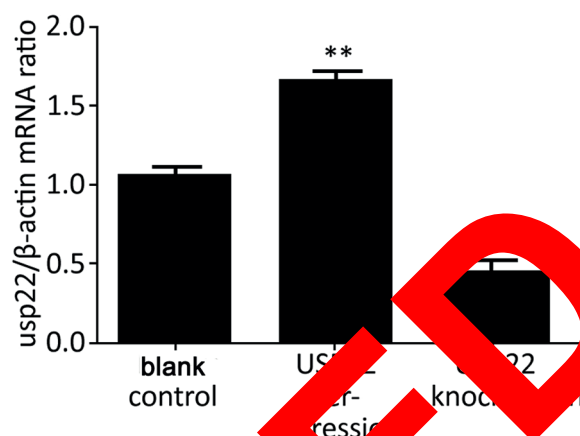


Figure 1. Effects of USP22 knockdown and over-expression lentivirus on USP22 expression. **, $p < 0.01$ compared to blank control group.

P53 protein was statistically down-regulated, as shown in Figure 2. In contrast, opposite effects were found with the infection by USP22-down-regulation lentivirus. These results suggested that USP22 activation could potentiate TERT/p53 signal pathway.

Effects of USP22 Up-Regulation and Down-Regulation on Cell Proliferation

Subsequently, MTT assay was performed to describe cell proliferation. Results showed (Figure 3) that the increase of USP22 level promoted cell proliferation whilst knockdown of USP22 remarkably suppressed proliferation level. These results indicated that the up-regulation of USP22 was correlated with enhancing cell proliferation potency.

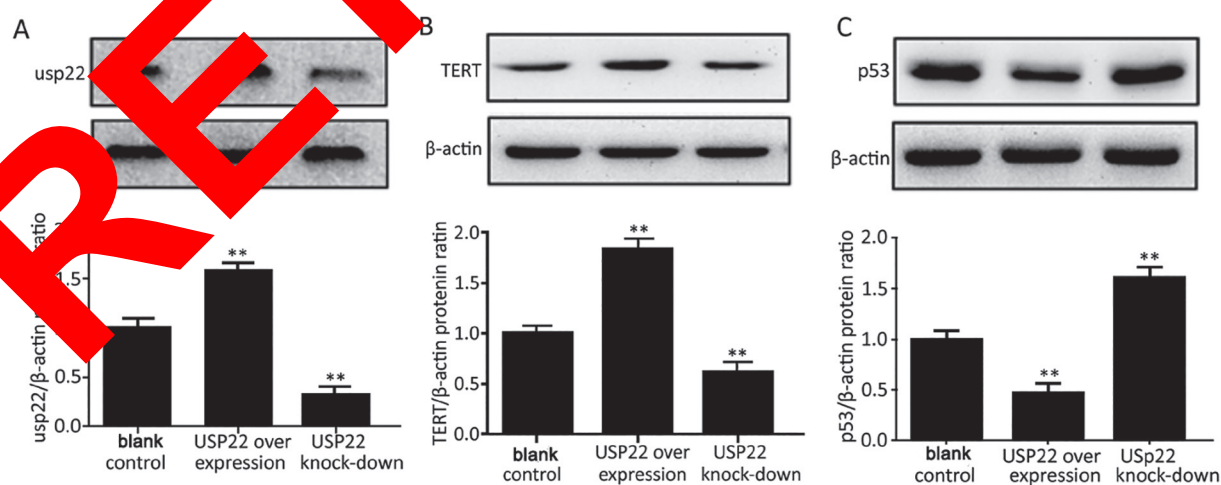


Figure 2. Effects of USP22 knockdown and over-expression on USP22, TERT and p53 protein expression. **, $p < 0.01$ compared to blank control group.

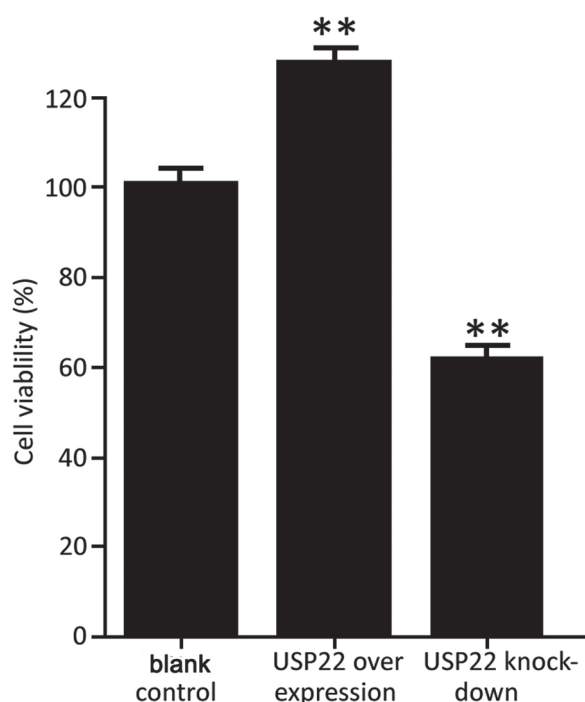


Figure 3. Effects of USP22 on cell proliferation. **, $p < 0.01$ compared to blank control group.

Telomerase Activity Change

In an assay for telomerase activity, we found that the rise of USP22 level enhanced telomerase activity, whilst the decrease of USP22 weakened telomerase activity, which showed significant difference compared to the control group (Figure 4).

Effects of USP22 Knock-Down and Over-Expression Lentivirus Transfection on Cell Apoptosis and DNA Damage

Flow cytometry was used to quantify the effect of USP22 on cell apoptosis. As shown in Figure 5A, the over-expression of USP22 significantly inhibited cell apoptosis, while a low level of USP22 significantly increased apoptotic cell number compared to control group. In the test of cellular DNA oxidative injury produce by H₂O₂ and FPG levels, we also found significantly decreased level of such DNA damage products in USP22 over-expression group, whilst DNA damage was elevated in USP22 knockdown group (Figure 5B). Similar results were obtained in comet assay, insignificant “tail-dragging” pattern was found in USP22 over-expression group, which was enhanced in USP22 knockdown group (Figure 5C). These results manifested that the suppression of USP22 could

weaken cell proliferation and enhance cellular DNA injury, thus may exert inhibitor roles on cancer cells.

Effects of USP22 knockdown or Overexpression Lentivirus on Cell Aging

Using cellular β -galactosidase staining, the increase of USP22 weakened staining intensity, whilst the reduction of USP22 significantly amplified number of β -galactosidase positive staining cells. These results illustrated that USP22 expression was correlated with cell aging, and the decrease of USP22 remarkably facilitated cell aging (Figure 6).

Discussion

Retinoblastoma is a type of malignant tumor derived from embryonic retinal cells. Current approaches mainly include conservative treatment or removal of eyeball. The prognosis is, however, unsatisfactory, leading to the requirement of identification of effective target and signal pathway necessary. An early study showed that USP22 was a critical target for the suppression of tumor growth in cells, and telomerase activity was related with malignant proliferation of cancer cells. However, in retinoblastoma cell line, whether USP22 can facilitate retinoblastoma cell aging or apoptosis via suppression TERT/P53 signal pathway has not been reported. This study thus used lentiviral infection to up-regulate and down-regulate USP22 RNA and protein expression level, for the study of the role of USP22 on TERT/P53 signal pathway and on

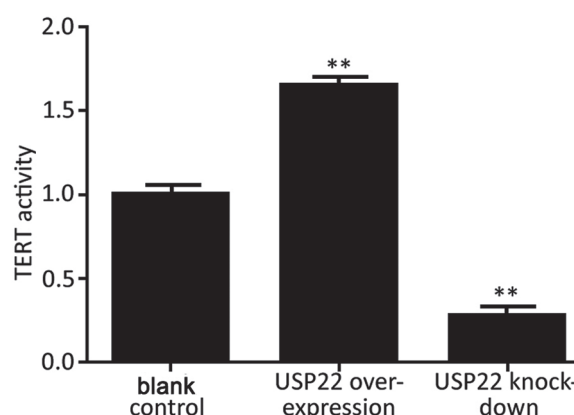


Figure 4. Effects of USP22 knockdown and over-expression lentivirus transfection on telomerase activity. **, $p < 0.01$ compared to blank control group.

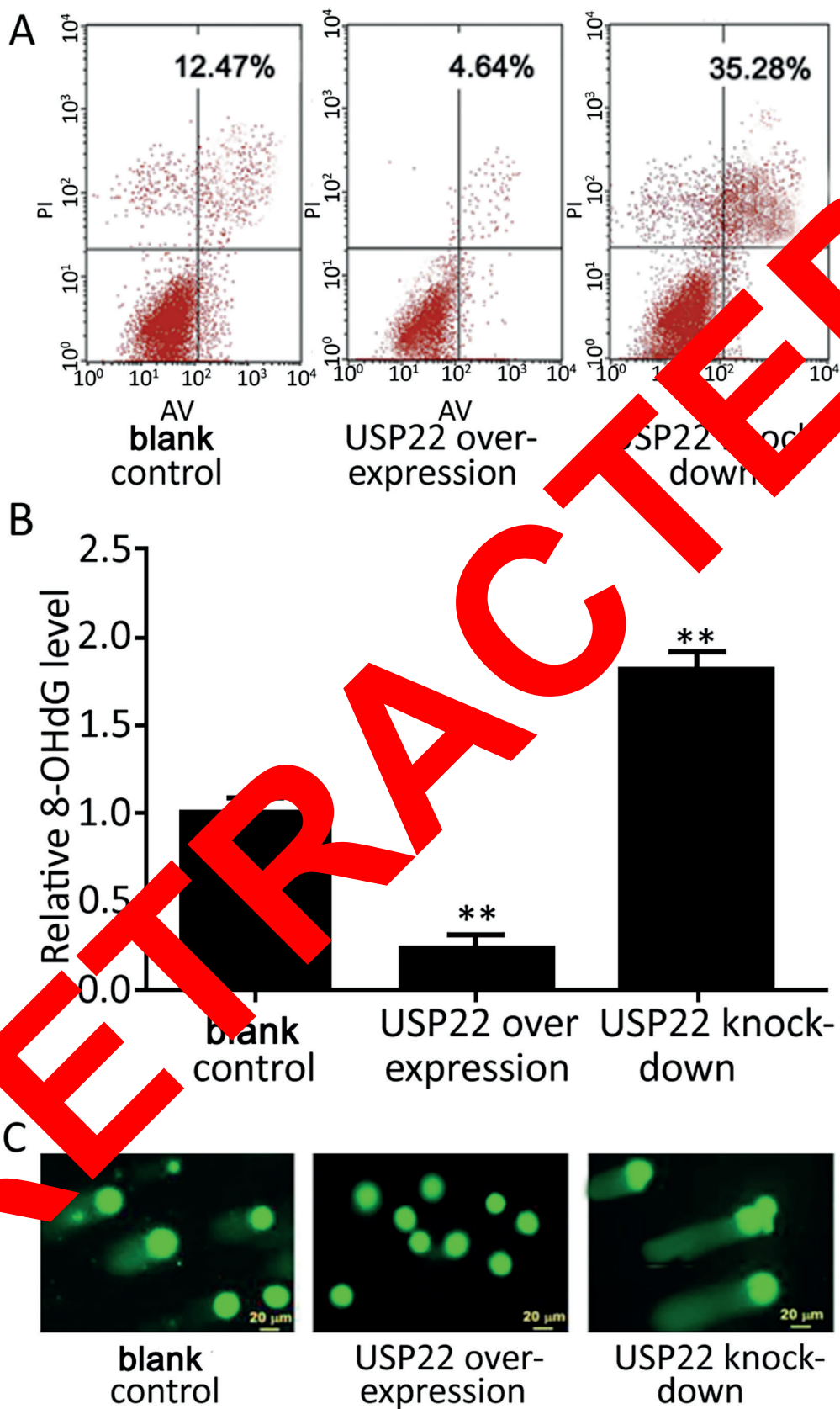


Figure 5. Effects of USP22 knockdown or over-expression lentivirus on cell apoptosis and DNA damage. **, $p < 0.01$ compared to blank control group.



Figure 6. Effects of USP22 knockdown and overexpression lentivirus on cell aging.

cancer cell behavior. Our results showed that in retinoblastoma cell line, down-regulation of USP22 inhibited TERT/P53 pathway, decreased telomerase activity, weakened cell proliferation potency, and facilitated cell aging, apoptosis or DNA damage. Increasing evidence showed critical functions of USP22 in pathology progression, and can work as markers for cancer treatment or prognosis with promising future.

In this study, we found that in retinoblastoma cells, USP22 expression was correlated with cell proliferation, aging and apoptosis behavior, indicating that down-regulation of USP22 could suppress cancer cell proliferation, and facilitate cell aging or apoptosis. Therefore, USP22 might be served as a diagnostic and a prediction marker for retinoblastoma patients. USP22 is a novel human de-ubiquitinated gene. A previous study¹⁰ indicated that in human ATC cell line 8505G, USP22 knockdown was up-regulated, resulting in G1 phase arrest and lower S or G2/M phase with depletion of USP22, thus it facilitated cell cycle progression in both cell lines. It is reported that the inhibition of USP22 expression effectively impeded the apoptosis of human cancer cells, including carcinoma and thyroid cancer. In the current study, AV-PI staining was used along with flow cytometry assay, and we found the favorable role of USP22 knockdown on apoptosis. Although it is still difficult to illustrate whether USP22 can directly work on apoptosis related proteins via transcriptional and translational regulation, our study still provided evidences for the activity of USP22 in retinoblastoma cell apoptosis mechanism. A previous study has shown that USP22 expression could affect cell proliferation or cycle and led to tumor cell immortalization due to elevated telomerase

activity¹⁸. Another report¹⁹ also showed that telomerase worked as a tumor marker and treatment target. Inhibition of telomerase activity could impede tumor cell immortalization. Moreover, the treatment of cancer cell and normal cell with certain concentration of telomerase decreased drug toxicity or adverse effects²⁰. Previous scholars²¹ demonstrated that activation of TERT/P53 pathway could enhance telomerase activity and decrease expression, thus facilitating cell proliferation via the modulation of cell cycle. However, the exact mechanism of USP22 on signal pathway still requires further researches. Moreover, treatment effects on USP22 or TERT on retinoblastoma are still to be further elucidated.

Conclusions

USP22 played an important role in proliferation, aging and apoptosis of retinoblastoma. The decrease of USP22 expression can facilitate human retinoblastoma cell aging and apoptosis via the inhibition of TERT/P53 signal pathway. USP22 thus may work as a potential treatment target against retinoblastoma.

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Conflict of Interest

The Authors declare that they have no conflict of interests.

References

- 1) ROH MR, PARK KH, CHUNG KY, SHIN SJ, RHA SY, TSAO H. Telomerase reverse transcriptase (TERT) promoter mutations in Korean melanoma patients. *Am J Cancer Res* 2017; 7: 134-138.
- 2) YIP BW, MOK HO, PETERSON DR, WAN MT, TANIGUCHI Y, GE W, AU DW. Sex-dependent telomere shortening, telomerase activity and oxidative damage in marine medaka *Oryzias melastigma* during aging. *Mar Pollut Bull* 2017; pii: S0025-326X(17)30032-2. [Epub ahead of print]
- 3) LEE SA, HUANG KC. Epigenetic profiling of human brain differential DNA methylation networks in schizophrenia. *BMC Med Genomics* 2016; 9(Suppl 3): 68.
- 4) LIANG JH, CHEN F, KRSTEW E, COWEN MS, CARROLL FY, CRAWFORD D, BEART PM, LAWRENCE AJ. The GABA(B) receptor allosteric modulator CGP7930, like baclofen, reduces operant self-administration of ethanol in alcohol-preferring rats. *Neuropharmacology* 2006; 50: 632-639.
- 5) HUANG C, ZHANG GF, HAN J, LIAO GJ, ZOU BG. Mechanism of age-related changes of bone marrow mesenchymal stem cells in senile osteoporosis. *Biol Regul Homeost Agents* 2016; 30: 565-568.
- 6) ZHANG Y, YAO L, ZHANG X, JI H, WANG L, SUN Y, PANG D. Elevated expression of USP22 in correlation with poor prognosis in patients with invasive breast cancer. *J Cancer Res Clin Oncol* 2011; 137: 1245-1253.
- 7) LIU YL, YANG YM, XU H, DONG Y. Overexpression of ubiquitin-specific protease 22 can promote cancer progression and predict therapy failure in human colorectal cancer. *J Gastroenterol Hepatol* 2016; 31: 1800-1805.
- 8) WANG H, LIU CH, CHEN JH, YU J, WANG L, ZHANG JL, YAO Q, LIU L, BIAN JF, FAN J, WANG R. Prognostic significance of USP22 as an oncogene in papillary thyroid carcinoma. *Tumour Biol* 2013; 34: 1635-1640.
- 9) LIU YL, XIAO Y, GU Z, LENG FQ, HUANG LQ, JIANG Y. Silencing of USP22 by asymmetric structure of interfering RNA inhibits proliferation and induces cell cycle arrest in bladder cancer cells. *Mol Cell Biochem* 2017; 346: 11-21.
- 10) ZHAO HD, TANG HL, LIU NN, ZHAO YL, LIU QQ, ZHU YC, JIA LT, GAO CF, YANG AG, LI JT. Targeting ubiquitin-specific protease 22 suppresses growth and metastasis of anaplastic thyroid carcinoma. *Oncotarget* 2016; 7: 31191-31203.
- 11) HOELLER D, DIKIC I. Targeting the ubiquitin system in cancer therapy. *Nature* 2009; 458: 438-444.
- 12) LI ZH, YU Y, DU C, FU H, WANG J, TIAN Y. RNA interference-mediated USP22 gene silencing promotes human brain glioma apoptosis and induces cell cycle arrest. *Oncol Lett* 2013; 5: 1289-1294.
- 13) NEVINS JR. The Rb/E2F pathway and cancer. *Annu Rev Mol Genet* 2001; 10: 699-703.
- 14) ZHAO H, TANG H, HUANG Q, QIU Y, LIU X, FAN D, GONG L, GUO H, CHEN C, LIU Y, YANG J, BAO G. MiR-101 targets USP22 to inhibit the proliferation of papillary thyroid carcinoma. *Am J Cancer Res* 2016; 6: 2575-2580.
- 15) ZHOU A, LIN K, ZHANG C, GUO Y, ZHANG N, XUE J, WANG Z, ALDARABSHI D, XIE Y, GOODGETT J, HUANG S. Nuclear GSK-3 β promotes tumorigenesis by phosphorylating KDM1A and blocking its deubiquitination by UBR2. *Nat Cell Biol* 2016; 18: 954-966.
- 16) WILSON WE, SHAY DK, PIATYSZEK MA. Modifications of a telomeric repeat amplification protocol (TRAP) result in increased reliability, linearity and sensitivity. *Nucleic Acids Res* 1995; 23: 3794-3795.
- 17) WU XL, JIAO JX, YANG YM, XU H, LIU JL, WANG XS. Ubiquitin-specific protease 22 as an oncogene by the activation of BMI-1-mediated INK4a/ARF pathway and Akt pathway. *Cell Biochem Biophys* 2012; 62: 229-234.
- 18) XU L, BIAN W, GU XH, SHEN C. Differing expression of cytokines and tumor markers in combined pulmonary fibrosis and emphysema compared to emphysema and pulmonary fibrosis. *COPD* 2017; 14: 245-250.
- 19) DE SIMONE P, VALIANTE M, SILIPO V. Familial melanoma and multiple primary melanoma. *G Ital Dermatol Venereol* 2017; 152: 262-265.
- 20) BLASIAK J. DNA-damaging anticancer drugs - a perspective for DNA repair-oriented therapy. *Curr Med Chem* 2017. [Epub ahead of print]
- 21) XIONG S, PATRUSHEV N, FOROUZANDEH F, HILENSKI L, ALEXANDER RW. PGC-1 α modulates telomere function and DNA damage in protecting against aging-related chronic diseases. *Cell Rep* 2015; 12: 1391-1399.
- 22) MENDELSON AR, LARRICK JW. Telomerase reverse transcriptase and peroxisome proliferator-activated receptor gamma co-activator-1 α cooperate to protect cells from DNA damage and mitochondrial dysfunction in vascular senescence. *Rejuvenation Res* 2015; 18: 479-483.