

Protective effect of 3-aminobenzamide, an inhibitor of poly (ADP-ribose) polymerase in distant liver injury induced by renal ischemia-reperfusion in rats

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Abstract. – BACKGROUND AND AIMS: Renal ischemia followed by reperfusion causes remote liver injury. This research was planned to investigate whether 3-aminobenzamide (3-AB), has any preventive effect against distant liver injury triggered by renal IR.

MATERIALS AND METHODS: Twenty four rats were randomly divided into three different groups Each group has 8 rats. The groups were as follows: (1) Sham operated group; (2) Renal ischemia-reperfusion (IR) group; (3) Renal IR+ 3-AB group. 3-AB (10 mg/kg) was given intraperitoneally 10 minute before reperfusion. At the end of study, the rats were sacrificed. Their liver tissues and serum samples were collected for measurement of malondialdehyde (MDA) levels, total oxidant status (TOS), total antioxidant status (TAS), paraoxonase (PON-1) activity and nitric oxide (NO).

RESULTS: Renal IR injury significantly increased Oxidative stress index (OSI) and MDA, TOS levels and significantly decreased PON-1 activity and TAS, NO levels in serum and liver tissue ($p < 0.05$). Despite that, changes in these biochemical parameters related with IR injury were diminished by 3-AB administration ($p < 0.05$).

CONCLUSIONS: The inhibition of PARP [Poly(ADP-Ribose)Polymerase] by 3-AB showed protective effects against distant liver injury triggered by renal ischemia-reperfusion by the ameliorating effects of 3-AB on oxidative stress.

Key Words:

3-aminobenzamide, Poly (ADP-Ribose) polymerase (PARP), Ischemia- reperfusion, Remote liver injury, Oxidative stress.

Introduction

Acute kidney injury is an ordinary and significant therapeutic contest for clinicians. Researchers emphasized that the frequency of acute kidney injury varies between clinical statements and popu-

lations. According to reports, more than 5000 acute kidney injury patients per million people per year classified as for non-dialysis-requiring, while about 300 sufferers per million people per year require dialysis treatment^{1,2}. Ischaemia-reperfusion (IR) injury induced acute renal injury arises in many clinical circumstances, such as urinary tract surgery, septic shock conditions, organ transplantation. The danger of renal failure remains – a devastating problem³. Latest researches have established that acute kidney injury triggered by IR induces dysfunction in liver^{4,5}. Renal IR injury decreases antioxidant enzyme activities^{3,5,6} and increase oxidative stress and hepatic lipid peroxidation products⁵⁻⁷ in liver tissue.

The poly(adenosine diphosphate-ribose) polymerase (PARP) is an enzyme which modifies proteins and polymerises nucleotides. It is considerably demonstrated in the nucleus⁸. PARP tract is implicated in the pathogenesis of distinct forms of IR injury⁹⁻¹¹. In several organ systems, the PARP inhibitor 3-aminobenzamide (3-AB) has been used successfully to decrease IR injury^{9,12-15}.

This study was planned to research the potential protective effect of 3-AB in distant liver injury induced by renal IR in rats.

Materials and Methods

Animals

Twenty four Wistar albino male rats weighing between 200 and 240 g were used in the present study. In the course of experiment, the rats were kept and maintained in constant laboratory conditions recommended by NIH. The study was initiated after obtaining approval from Dicle University Local Committee on Animal Research

Ethics. We meticulously complied with the principles for the "Protection of Animal Rights" specified by the National Institutes of Health (NIH) during the entire course of the research.

Experimental Method and Procedure

Rats were put under anaesthesia with ketamine (75 mg/kg i.p.) and xylazine (8 mg/kg i.p.). Body temperature was maintained in every part of surgery at $37 \pm 1^\circ\text{C}$. All rats were submitted to surgical exposure of the left and right renal pedicles via midline incision. To create renal ischemia, both renal pedicles were blocked for 45 min with vascular clamps. After 45 min of blockage, the clamps were taken away and kidneys examined to endure reperfusion for 24 hrs. Rats were randomly divided into three different groups (n= 8). The groups were as follows: (1) Sham operated group; (2) Renal IR group; (3) Renal IR+ 3-AB group. 3-AB (10 mg/kg) (Sigma, St. Louis, MO, USA) was administered intraperitoneally 10 minute before reperfusion. At the end of experimental procedure, the rats were sacrificed. Their liver tissues and serum samples were collected for biochemical analysis. The excised liver tissue samples were weighed and all samples instantly stored at -70°C .

Biochemical Assay

The liver tissues washed with 1.15% ice-cold KCl, minced, then homogenized in five volumes (w/v) of the same solution. Assays were performed on the supernatant of the homogenate. The protein concentrations of the tissue and serum sample were assessed by the method of Lowry et al¹⁶. Lipid peroxidation level was phrased as malondialdehyde (MDA). It was assessed by the method of Ohkawa et al¹⁷. Nitric oxide (NO) levels were assessed with Griess' method¹⁸. Paraoxonase (PON-1) activity was as-

sessed spectrophotometrically by modified Eckerson et al method¹⁹. The total antioxidant status (TAS) of supernatant fractions was appraised by using a Erel method²⁰. TAS results are expressed as nmol Trolox equivalent/mg protein. The total oxidant status (TOS) of supernatant fractions was appraised by using a new method Erel²¹. The results are expressed in terms of nmol H_2O_2 equivalent/mg protein²². The TOS/TAS ratio was considered as the oxidative stress index (OSI). The tissue OSI value was calculated as follows: $\text{OSI} = \text{TOS}/\text{TAS}$ ²³.

Statistical Analysis

Statistical analysis was performed using SPSS (version 11.0; SPSS Inc., Chicago, IL, USA). Data were shown as means \pm standard deviation, and Kruskal-Wallis test was used for analysis. In the event of significant results, the Mann-Whitney U test was used for comparisons of differences between two independent groups. A p value < 0.05 was estimated statistically significant.

Results

Consequences of Renal Ischemia on Blood Biochemical Variables

Serum MDA, TOS, TAS, OSI, NO levels and PON-1 enzyme activities are demonstrated in Table I. There were notable changes in the PON-1 enzyme activities and MDA, TOS, TAS, OSI, NO levels in the serum of the IR group compared to sham operated group ($p < 0.01$). In the 3-AB treated IR group, TAS, NO and PON-1 levels were notably elevated when compared to the IR group ($p < 0.001$). On the other hand, MDA ($p < 0.05$), TOS ($p < 0.01$) and OSI ($p < 0.01$) levels were notably reduced in IR+3-AB group when compared to the IR group ($p < 0.05$).

Table I. Levels of MDA, TOS and TAS, OSI, PON-1 activity and NO in serum samples.

| Groups | Sham (n: 8) | IR (n: 8) | IR+3-AB (n: 8) |
|---|------------------|-------------------------------|-------------------------------|
| MDA (nmol/g protein)* | 259.9 \pm 22.1 | 367.4 \pm 54.4 ^a | 256.4 \pm 42.9 ^c |
| TOS (mmol H_2O_2 Equiv./g protein)* | 57.4 \pm 12.4 | 162.4 \pm 11.1 ^a | 78.0 \pm 21.4 ^b |
| TAS (mmol Trolox Equiv./g protein)* | 1.16 \pm 0.12 | 0.84 \pm 0.08 ^a | 1.28 \pm 0.22 ^b |
| OSI (H_2O_2 /Trolox)* | 49.5 \pm 11.3 | 196.0 \pm 34.3 ^a | 60.9 \pm 13.3 ^b |
| PON-1 activity (U/mg protein)* | 165.7 \pm 29.1 | 124.5 \pm 11.0 ^a | 146.1 \pm 28.3 ^b |
| NO ($\mu\text{mol/g protein}$)* | 365.7 \pm 55.7 | 139.7 \pm 45.3 ^a | 362.3 \pm 77.1 ^b |

Results are presented as means \pm standard deviation. * $p < 0.01$ for Kruskal Wallis test. ^a $p < 0.01$ as compared to the sham operated group, ^b $p < 0.001$ as compared to the IR group. ^c $p < 0.05$ as compared to the IR group. IR: Ischemia-reperfusion, 3-AB: 3-aminobenzamide. MDA: malondialdehyde, TOS: Total oxidant status, TAS: Total antioxidant status, OSI: Oxidative stress index, PON-1: paraoxonase activity, NO: Nitric oxide.

Consequence of Renal Ischemia on Liver Biochemical Parameters

The MDA, TOS, TAS, OSI, NO levels and PON-1 enzyme activities in the liver tissues are demonstrated in Table II. MDA, TOS and OSI levels were notably raised and TAS, NO and PON-1 levels were notably reduced in the IR as compared to the sham operated group ($p < 0.01$). However, TAS ($p < 0.05$), NO ($p < 0.01$) and PON-1 ($p < 0.01$) levels were notably elevated in 3-AB treated IR group compared to the IR group. MDA ($p < 0.01$), TOS ($p < 0.05$) and OSI ($p < 0.05$) levels were notably reduced in IR+3-AB group compared to the IR group ($p < 0.05$).

Discussion

This research was planned to explore whether 3-AB, a PARP inhibitor, has a protective effect on distant liver damage triggered by renal IR by diminishing oxidative stress. PARP is an enzyme that modifies proteins and polymerises nucleotides. It is considerably found in the nucleus. The essential stimulus of PARP activation is DNA single strand break, which can be generated by a variety of environmental factors and free radical and oxidants⁸. The PARP pathway is implicated in the pathogenesis of various forms of IR injury⁹⁻¹¹. Reactive oxygen species (ROS) generated during IR are powerful activators of DNA single-strand cleavage and the consequent activation of the nuclear enzyme PARP²⁴. Inhibition of PARP activation exerts beneficial effects that ameliorate the metabolic alterations but not the occurrence of DNA damage in inflammatory response during IR processes²⁵. Over activation of PARP may cause to cell death due to energy depletion²⁶⁻²⁸. Because of an immediate depletion of intracellular nicotinamide adenine dinu-

cleotide (NAD⁺) and adenosine triphosphate (ATP) energy pools, glycolysis and the mitochondrial respiration rate decelerate, leading to cellular dysfunction and death²⁹. Totally, this activity is called as the poly (ADP-ribose) polymerase suicide hypotheses³⁰. This cell suicide phenomenon is driven by PARP activation and has been shown in several cell types²⁵. PARP inhibition exerts favourable effects against free radical-mediated cell injury³¹. In several organ systems, the PARP inhibitor 3-AB has been used successfully to decrease IR injury¹²⁻¹⁵.

It was demonstrated that IR of the tissue to be associated with lipid peroxidation, which is an autocatalytic process causing to oxidative demolition of the cellular membranes, and their catabolites can cause to form harmful metabolites and cell death^{7,32-34}. Lipid peroxidation, as a free radical-producing system, has been proposed to be tightly linked to IR -induced tissue damage, and MDA is an important parameter of oxidative stress is a good pointer of lipid peroxidation⁷. In this study, in IR group the level of MDA notably elevated in the serum and liver tissue, while exogenously administered 3-AB repressed MDA elevation notably. This finding indicates that renal IR induced lipid peroxidation is likely to be improved with 3-AB administration in liver tissue.

Besides MDA level, assessment of TOS, TAS, and OSI contributes unusual and predictable index of oxidative stress. We assayed oxidative status as TOS and TAS along with the assessment of OSI, a pointer of oxidative stress, which exhibits the redox balance between oxidation and antioxidation³⁵. Since separate measurement of different oxidant molecules such as superoxide radical anion, hydrogen peroxide is not functional and their oxidant effects are linear, we measured TOS in serum as previously described by Ereli²¹. Likewise, we measured TAS, instead of assaying antioxidant mole-

Table II. Levels of MDA, TOS and TAS, OSI, PON-1 activity and NO in liver tissue samples.

| Groups | Sham (n: 8) | IR (n: 8) | IR+3-AB (n: 8) |
|--|--------------|---------------------------|----------------------------|
| MDA (nmol/g protein)* | 237.6 ± 20.2 | 335.8 ± 49.8 ^a | 234.4 ± 39.2 ^b |
| TOS (mmol H ₂ O ₂ Equiv./g protein)* | 44.7 ± 22.1 | 97.1 ± 27.9 ^a | 76.5 ± 31.8 ^c |
| TAS (mmol Trolox Equiv./g protein)* | 1.14 ± 0.24 | 0.70 ± 0.21 ^a | 0.93 ± 0.24 ^c |
| OSI (H ₂ O ₂ /Trolox)* | 39.2 ± 18.3 | 149.5 ± 68.7 ^a | 90.2 ± 49.1 ^c |
| PON-1 activity (U/mg protein)* | 6.08 ± 1.78 | 2.40 ± 0.65 ^a | 5.57 ± 1.90 ^b |
| NO (μmol/g protein)* | 490.1 ± 74.6 | 187.2 ± 60.7 ^a | 485.5 ± 103.3 ^b |

Results are presented as means ± standard deviation. * $p < 0.01$ for Kruskal Wallis test. ^a $p < 0.01$ as compared to the sham operated group, ^b $p < 0.001$ as compared to the IR group. ^c $p < 0.05$ as compared to the IR group. IR: Ischemia-reperfusion, 3-AB: 3-aminobenzamide. MDA: malondialdehyde, TOS: Total oxidant status, TAS: Total antioxidant status, OSI: Oxidative stress index, PON-1: paraoxonase activity, NO: Nitric oxide.

cules separately following the methods of Erel³⁶ and Cikrikcioglu et al³⁷. Lately, it has been published that OSI may exhibit the oxidative status more accurately than TOS or TAS level alone³⁸. In this research, serum and liver tissue TOS levels and OSI values were notably increased and the level of TAS was notably reduced in IR group compared with sham operated group. On the other hand, in 3-AB treated IR rats TAS level was significantly increased compared with IR group. These results showed that renal IR leads to increase of oxidative stress in liver tissue, and this increase was prevented by administration of 3- AB.

PON-1, an enzyme associated with high-density lipoprotein that is mainly secreted by the liver. Although its physiological activity has not been completely clarified, it seems PON-1 is responsible for hydrolyzing lipid peroxides and also to play a major role in the antioxidant system³⁹. PON-1 protects liver against inflammation, liver disease and fibrosis^{40,41}. In this study, we found decreased PON-1 activity in the serum and liver tissues of IR rats compared to sham operated rats. 3-AB treatment was reversed the reduced activity of PON-1 in IR+3-AB group. 3-AB treatment prevented the reduced activity of PON-1 in liver tissue. Our data show that a reduction in PON-1 activity is linked to oxidative damage in the liver tissues caused by renal IR and 3-AB administration reverses the decrease of PON-1 activity in these tissues.

NO in the liver also exerts vasodilatory and cytoprotective effects. Inhibition of NO production in the liver has conventionally been demonstrated to be harmful in reperfusion injury models^{8,42}. Studies show that nitric oxide synthase inhibitors reduce microvascular perfusion and aggravate liver injury during IR^{8,43}. These data were also supported in eNOS gene knockout mice^{8,44,45}. A reduction of NO during IR, generally caused by endothelial dysfunction and reduction of endothelial nitric oxide synthase activity^{46,47}. In this work, renal IR reduced the NO level and 3- AB treatment increased NO content in serum and hepatic tissue. These findings showed that 3-AB can increase renal IR associated endothelial dysfunction and reduction of endothelial nitric oxide synthase activity.

Conclusions

This report is the first to investigate the effects of 3-AB against distant liver injury triggered by renal ischemia-reperfusion in rats. Data revealed that the inhibition of PARP by 3-AB showed

protective effects against distant liver injury triggered by renal ischemia-reperfusion by the ameliorating effects of 3-AB on oxidative stress.

Conflict of Interest

The Authors declare that there are no conflicts of interest.

References

- 1) BELLOMO R, KELLUM JA, RONCO C. Acute kidney injury. *Lancet* 2012; 380(9843): 756-766.
- 2) LIANGOS O, WALD R, O'BELL JW, PRICE L, PEREIRA BJ, JABER BL. Epidemiology and outcomes of acute renal failure in hospitalized patients: a national survey. *Clin J Am Soc Nephrol* 2006; 1: 43-51.
- 3) SERTESER M, KOKEN T, KAHRAMAN A, YILMAZ K, AKBULUT G, DILEK ON. Changes in hepatic TNF-alpha levels, antioxidant status, and oxidation products after renal ischemia/reperfusion injury in mice. *J Surg Res* 2002; 107: 234-240.
- 4) PARK SW, CHEN SW, KIM M, BROWN KM, KOLLS JK, D'AGATI VD, LEE HT. Cytokines induce small intestine and liver injury after renal ischemia or nephrectomy. *Lab Invest* 2011; 91: 63-84.
- 5) WANG B, BAI M, BAI Y, LI Q. Liver injury following renal ischemia reperfusion in rats. *Transplant Proc* 2010; 42: 3422-3426.
- 6) KADKHODAEI M, GOLAB F, ZAHMATKESH M, GHAZNAVI R, HEDAYATI M, ARAB HA, OSTAD SN, SOLEIMANI M. Effects of different periods of renal ischemia on liver as a remote organ. *World J Gastroenterol* 2009; 15: 1113-1118.
- 7) KAÇMAZ A, USER EY, SEHIRLI AO, TILKI M, OZKAN S, SENER G. Protective effect of melatonin against ischemia/reperfusion-induced oxidative remote organ injury in the rat. *Surg Today* 2005; 35: 744-750.
- 8) GERO D, SZABÓ C. Role of the peroxynitrite-poly (ADP-ribose) polymerase pathway in the pathogenesis of liver injury. *Curr Pharm Des* 2006; 12: 2903-2910.
- 9) SZABO C, DAWSON VL. Role of poly(ADP-ribose) synthetase in inflammation and ischaemia-reperfusion. *Trends Pharmacol Sci* 1998; 19: 287-298.
- 10) SHALL S, MURCIA G. Poly(ADP-ribose) polymerase-1: What have we learned from the deficient mouse model? *Mutat Res* 2000; 460: 1-15.
- 11) TENTORI L, PORTARENA I, GRAZIANI G. Potential clinical applications of poly(ADP-ribose) polymerase (PARP) inhibitors. *Pharmacol Res* 2002; 45: 73-85.
- 12) THIEMERMANN C, BOWES J, MYINT FP, VANE JR. Inhibition of the activity of poly(ADP ribose) synthetase reduces ischemia-reperfusion injury in the heart and skeletal muscle. *Proc Natl Acad Sci U S A* 1997; 94: 679-683.
- 13) HEKIMOGLU A, KURCER Z, ARAL F, BABA F, ATESSAHIN A, SAKIN F. Effects of 3-aminobenzamide on unilateral testicular ischemia-reperfusion injury: what is the role of PARP inhibition? *Can J Physiol Pharmacol* 2010; 88: 1123-1129.
- 14) BOWES J, THIEMERMANN C. Effects of inhibitors of the activity of poly (ADP-ribose) synthetase on the liv-

- er injury caused by ischaemia-reperfusion: a comparison with radical scavengers. *Br J Pharmacol* 1998; 124: 1254-1260.
- 15) OZTAS E, GUVEN A, TURK E, UYSAL B, AKGUL EO, CAYCI T, ERSOZ N, KORKMAZ A. 3-aminobenzamide, a poly ADP ribose polymerase inhibitor, attenuates renal ischemia/reperfusion injury. *Ren Fail* 2009; 31: 393-399.
 - 16) LOWRY OH, ROSEBROUGH NJ, FARR AL, RANDALL RJ. Protein measurement with the Folin phenol reagent. *J Biol Chem* 1951; 193: 265-275.
 - 17) OHKAWA H, OHISHI N, YAGI K. Assay for lipid peroxides in animal tissues by thiobarbituric acid reaction. *Anal Biochem* 1979; 95: 351-358.
 - 18) CORTAS NK, WAKID NW. Determination of inorganic nitrate in serum and urine by a kinetic cadmium-reduction method. *Clin Chem* 1990; 36: 1440-1443.
 - 19) ECKERSON HW, WYTE CM, LA DU BN. The human serum paraoxonase/arylesterase polymorphism. *Am J Hum Genet* 1983; 35: 1126-1138.
 - 20) EREL O. A novel automated method to measure total antioxidant response against potent free radical reactions. *Clin Biochem* 2004; 37: 112-119.
 - 21) EREL O. A new automated colorimetric method for measuring total oxidant status. *Clin Biochem* 2005; 38: 1103-1111.
 - 22) OZTURK E, BALAT O, ACILMIS YG, OZCAN C, PENCE S, EREL O. Measurement of the placental total antioxidant status in preeclamptic women using a novel automated method. *J Obstet Gynaecol Res* 2011; 37: 337-342.
 - 23) AYCICEK A, EREL O, KOCYIGIT A. Increased oxidative stress in infants exposed to passive smoking. *Eur J Pediatr* 2005; 164: 775-778.
 - 24) ZINGARELLI B, CUZZOCREA S, ZSENGELLÉR Z, SALZMAN AL, SZABÓ C. Protection against myocardial ischemia and reperfusion injury by 3-aminobenzamide, an inhibitor of poly(-ADP-ribose) synthetase. *Cardiovasc Res* 1997; 36: 205-215.
 - 25) ZINGARELLI B, O'CONNOR M, WONG H, SALZMAN AL, SZABÓ C. Peroxynitrite-mediated DNA strand breakage activates poly-adenosine diphosphate ribosyl synthetase and causes cellular energy depletion in macrophages stimulated with bacterial lipopolysaccharide. *J Immunol* 1996; 156: 350-358.
 - 26) WINTERSBERGER U, WINTERSBERGER E. POLY ADP-ribosylation—a cellular emergency reaction? *FEBS Lett* 1985; 188: 189-191.
 - 27) COCHRANE G. Mechanisms of oxidant injury of cells. *Mol Aspects Med* 1991 12: 137-147.
 - 28) BANASIK M, KOMURA H, SHIMOYAMA M, UEDA K. Specific inhibitors of poly(ADP-ribose) synthetase and mono (ADP-ribose) transferase. *J Biol Chem* 1992; 267: 1569-1575.
 - 29) UEDA K, HAYAISHI O. ADP-ribosylation. *Annu Rev Biochem* 1985; 54: 73-100.
 - 30) BERGER NA. Poly(ADP-ribose) in the cellular response to DNA damage. *Radiat Res* 1985; 101: 4-15.
 - 31) PACHER P, LAUDET L, MABLEY JG, CZIRÁKI A, HASKÓ G, SZABÓ C. Beneficial effects of a novel ultrapotent poly (ADP-ribose) polymerase inhibitor in murine models of heart failure. *Int J Mol Med* 2006; 17: 369-375.
 - 32) ESCHWEGE P, PARADIS V, CONTI M, HOLSTEGE A, RICHET F, DETEVE J, MENAGER P, LEGRAND A, JARDIN A, BEDOS SA P, BENOIT G. In situ detection of lipid peroxidation by-products as markers of renal ischemia injuries in rat kidneys. *J Urol* 1999; 162: 553-557.
 - 33) VAGHASIYA JD, SHETH NR, BHALODIA YS, JIVANI NP. Exaggerated liver injury induced by renal ischemia reperfusion in diabetes: effect of exenatide. *Saudi J Gastroenterol* 2010; 16: 174-180.
 - 34) IERI SO, GEDIK IE, ERZIK C, USLU B, ARBAK S, GEDIK N, YE EN BC. Oxytocin ameliorates skin damage and oxidant gastric injury in rats with thermal trauma. *Burns* 2008; 34: 361-369.
 - 35) DAVIES GR, SIMMONDS NJ, STEVENS TR, GRANDISON A, BLAKE DR, RAMPTON DS. Mucosal reactive oxygen metabolite production in duodenal ulcer disease. *Gut* 1992; 33: 1467-1472.
 - 36) EREL O. A novel automated direct measurement method for total antioxidant capacity using a new generation, more stable ABTS radical cation. *Clin Biochem* 2004; 37: 277-285.
 - 37) CIKRIKCIOGLU MA, HURSI TOGLU M, ERKAL H, KINAS BE, SZTAJZEL J, CAKIRCA M, ARSLAN AG, EREK A, HALAC G, TUKEK T. Oxidative stress and autonomic nervous system functions in restless legs syndrome. *Eur J Clin Invest* 2011; 41: 734-742.
 - 38) HARMA M, HARMA M, EREL O. Increased oxidative stress in patients with hydatidiform mole. *Swiss Med Wkly* 2003; 133: 563-566.
 - 39) AVIRAM M, ROSENBLAT M. Paraoxonases 1, 2, and 3, oxidative stress, and macrophage foam cell formation during atherosclerosis development. *Free Radic Biol Med* 2004; 37: 1304-1316.
 - 40) HASHEMI M, BAHARI A, HASHEMZEHNI N, MOAZENI-ROODI A, SHAFIEIPOUR S, BAKHSHIPOUR A, GHAVAMI S. Serum paraoxonase and arylesterase activities in Iranian patients with nonalcoholic fatty liver disease. *Pathophysiology* 2012; 738: 1-5.
 - 41) MARSILLACH J, BERTRAN N, CAMPS J, FERRE N, RIU F, TOUS M, COLL B, ALONSO-VILLAVERDE C, JOVEN J. The role of circulating monocyte chemoattractant protein-1 as a marker of hepatic inflammation in patients with chronic liver disease. *Clin Biochem* 2005; 38: 1138-1140.
 - 42) JAESCHKE H. Molecular mechanisms of hepatic ischemiareperfusion injury and preconditioning. *Am J Physiol Gastrointest Liver Physiol* 2003; 284: 15-26.
 - 43) WANG Y, MATHEWS WR, GUIDO DM, FARHOOD A, JAESCHKE H. Inhibition of nitric oxide synthesis aggravates reperfusion injury after hepatic ischemia and endotoxemia. *Shock* 1995; 4: 282-288.
 - 44) HINES IN, KAWACHI S, HARADA H, PAVLICK KP, HOFFMAN JM, BHARWANI S, WOLF RA, GRISHAM MB. Role of nitric oxide in liver ischemia and reperfusion injury. *Mol Cell Biochem* 2002; 1: 229-237.
 - 45) KAWACHI S, HINES IN, LAROUX FS, HOFFMAN J, BHARWANI S, GRAY L, LEFFER D, GRISHAM MB. Nitric oxide synthase and postischemic liver injury. *Biochem Biophys Res Commun* 2000; 276: 851-854.
 - 46) KÖKEN T, INAL M. The effect of nitric oxide on ischemia-reperfusion injury in rat liver. *Clin Chim Acta* 1999; 288: 55-62.
 - 47) UHLMANN D, UHLMANN S, SPIEGEL HU. Endothelin/nitric oxide balance influences hepatic ischemiareperfusion injury. *J Cardiovasc Pharmacol* 2000; 36: 212-214.