

Diagnostic accuracy of a real-time PCR assay for detection of *Helicobacter pylori* and resistance to clarithromycin and levofloxacin directly from stool

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ABSTRACT. – OBJECTIVE: The non-invasive detection of *Helicobacter pylori* (*H. pylori*) and its resistance to clarithromycin and levofloxacin significantly improves the management of infected patients by enabling tailored eradication treatments without the need for endoscopic procedures. This study aimed to assess the effectiveness of real-time PCR (RT-PCR) assays in identifying *H. pylori* infection and antibiotic resistance in stool and gastric biopsy specimens.

PATIENTS AND METHODS: Stool and gastric biopsy samples were collected from patients within three days of post-hospitalization. A total of 115 samples were analyzed for *H. pylori* infection, and an additional 115 samples were evaluated for resistance to clarithromycin and levofloxacin using an RT-PCR-based molecular test. Statistical analyses were performed using (SPSS 26.0 IBM Corp., Armonk, NY, USA).

RESULTS: Among 115 patients (53 males, average age 50.8±13.2 years), *H. pylori* was detected in 93.1% of stool samples and 93.9% of gastric biopsies. The RT-PCR assay demonstrated a sensitivity of 99.1% and a specificity of 100%, with an overall diagnostic accuracy of 99.1%. Clarithromycin resistance was found in 37.3% of stool and 46.9% of gastric biopsy specimens, with the assay showing 79.6% sensitivity and 98.4% specificity. Levofloxacin resistance was identified in 32.1% of stool samples and 31.3% of gastric biopsies, with 86.3% sensitivity and 91.1% specificity of the molecular test.

CONCLUSIONS: The RT-PCR-based detection of *H. pylori* and its resistance to clarithromycin and levofloxacin in stool samples represents a promising approach to enhance eradication therapy outcomes, potentially improving treatment efficacy.

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Key Words:

Helicobacter pylori, Antibiotic resistance, Stool, Molecular diagnosis, Real-time polymerase chain reaction.

Introduction

Helicobacter pylori (*H. pylori*), a spiral-shaped, Gram-negative bacterium, infects approximately 50% of humans worldwide^{1,2}. Despite a declining incidence in developed countries, *H. pylori* infection remains a major cause of morbidity and mortality worldwide³. *H. pylori* infection is associated with a range of serious gastroduodenal diseases, including chronic gastritis, peptic ulcer disease, mucosa-associated lymphoid tissue lymphoma, and gastric cancer, underscoring the need for *H. pylori* eradication worldwide⁴. Given its carcinogenic potential, *H. pylori* has been classified as a class I carcinogen by the World Health Organization (WHO) in 1994 and the U.S. Department of Health and Human Services (HHS) in 2022^{5,6}. Therefore, the eradication of *H. pylori* is considered to be very important for the management of these diseases.

The diagnosis of *H. pylori* infection involves both invasive and non-invasive methods⁷. Detection of *H. pylori* from stool samples is non-invasive and easy to perform in epidemiological studies and diagnosis of infection in children⁸. In recent years, several molecular approaches for the detection of *H. pylori* infection and its antimicrobial resistance have been developed⁹. Notably, the non-invasive molecular testing through real-time polymerase chain reaction (RT-PCR) on stool samples has shown high sensitivity and specificity¹⁰. A meta-analysis¹¹ has identified that the bacterial 23S ribosomal RNA subunit gene as the most dependable marker for diagnosing *H. pylori* infection on stool samples, with a sensitivity and specificity of 82% and 99%, respectively. Therefore, there is an increasing need for non-invasive methods to diagnose *H. pylori* infection and antimicrobial resistance.

The prevalence of *H. pylori* antibiotic resistance has been increasing remarkably worldwide, and the success rate of *H. pylori* eradication is decreasing¹². The main cause of treatment failure has been reported as antimicrobial resistance; the resistance levels for clarithromycin and levofloxacin have reached 20%-50%, in China alone^{13,14}. Although RT-PCR assay is the most sensitive and specific method for detecting *H. pylori* in gastric biopsy specimens, there is a lack of studies comparing the RT-PCR assay using different sample types¹⁵. Therefore, this study aims to evaluate the diagnostic accuracy of the stool-based *H. pylori* RT-PCR test as an alternative approach for detecting *H. pylori* infection and determining antibiotic resistance and compared to the gastric biopsy specimens *H. pylori* RT-PCR test, which serves as the reference standard.

Patients and Methods

Study Design and Assessment

This study was conducted between April 2023 and August 2023, patients were eligible if they were aged 18-75 years and had confirmed *H. pylori* infection *via* the C-urea breath test (UBT)¹⁴. A total of 115 patients from outpatient clinics met the eligibility criteria and were enrolled.

Inclusion Criteria: male or female patients aged 18-75 years; diagnosis of *H. pylori* infection by C-UBT test¹⁴; presence of upper gastrointestinal symptoms (e.g., epigastric pain, acid reflux, heartburn, epigastric distention, nausea); consent to participate *via* signed informed consent form. **Exclusion Criteria:** antibiotic, bismuth, or proton pump inhibitor (PPI) use within the last four weeks; recurrent or long-term use of macrolides and penicillin; serious primary diseases (liver, kidney, heart, brain, lung, endocrine system, hematopoietic system); significant liver or kidney insufficiency; pregnancy, lactation, positive pregnancy test, or intent to become pregnant within six months.

Stool and gastric biopsy samples were collected from patients 1 to 3 days post-hospitalization. The

H. pylori infection and antibiotic resistance (clarithromycin and levofloxacin) were assessed using RT-PCR on both stool and gastric biopsy samples, employing the clarithromycin, quinolone-resistant and non-resistant *Helicobacter pylori* Nucleic Acid Amplification Test kit (Jiangsu Mole Bioscience Co., Ltd, Taizhou, Jiangsu, China).

Ethics Approval

This study followed the Consolidated Standards of Reporting Trials (CONSORT) guidelines. Written informed consent was obtained from all participants involved in the study. The study protocol was approved by the Institutional Ethics Board of the Civil Aviation General Hospital, Beijing, China (No. 2023-L-K-05). The trial was registered in the Chinese Clinical Trials Registration (www.chictr.org.cn) with the registration number ChiCTR2300070267.

Statistical Analysis

Statistical analyses were conducted using SPSS 26.0 (IBM Corp., Armonk, NY, USA). The sensitivity, specificity, positive predictive value (PPV), and negative predictive value (NPV) of each test were investigated. The agreement between stool and gastric biopsy sample PCR test was estimated with the kappa coefficient. A *p*-value lower than 0.05 was considered statistically significant.

Results

In this study, 115 patients were enrolled, comprising 53 males and 62 females, with an average age of 50.8±13.2 years. *H. pylori* infection was detected in 108 patients (93.1%) of stool samples and 109 patients (93.9%) through gastric biopsy samples *via* RT-PCR test. As shown in Table I, the RT-PCR assay achieved a sensitivity of 99.1% and a specificity of 100%, resulting in a diagnostic accuracy of 99.1%. The agreement between RT-PCR results from stool and gastric biopsy samples was excellent, evidenced by a Kappa score of 0.929 (*p*<0.001).

Table I. Performance of RT-PCR detection of *H. pylori* in stool and gastric biopsy samples.

	Sensitivity	Specificity	PPV	NPV	Accuracy	Kappa value	<i>p</i> -value
Analytical performance	99.1%	100%	100%	87.5%	99.1%	0.929	0.001

PPV: positive predictive value; NPV: negative predictive value; RT-PCR: real-time polymerase chain reaction.

Table II. Performance of RT-PCR detection of clarithromycin and levofloxacin resistance in stool and gastric biopsy samples.

Antibiotic resistance	Sensitivity	Specificity	PPV	NPV	Accuracy	Kappa value	p-value
Clarithromycin	79.6%	98.4%	97.7%	84.5%	78.0%	0.788	0.001
Levofloxacin	83.3%	91.1%	81.1%	92.3%	74.4%	0.739	0.001

PPV: positive predictive value; NPV: negative predictive value; RT-PCR: real-time polymerase chain reaction.

Clarithromycin resistance was identified in 43 patients (37.3%) *via* stool samples and 54 patients (46.9%) through gastric biopsy samples. In Table II, the sensitivity and specificity of the RT-PCR test for clarithromycin resistance were 79.6% and 98.4%, respectively, with an overall diagnostic accuracy of 78.0%. The concordance between the RT-PCR results for clarithromycin resistance from stool and gastric biopsy samples was good, with a Kappa value of 0.788 ($p < 0.001$).

Levofloxacin resistance was detected in 37 patients (32.1%) from stool samples and 36 patients (31.3%) from gastric biopsy samples. As shown in Table II, the RT-PCR test for levofloxacin resistance exhibited a sensitivity of 86.3% and a specificity of 91.1%, with a diagnostic accuracy of 74.4%. The concordance for levofloxacin resistance between stool and gastric biopsy samples' RT-PCR test was also good, indicated by a Kappa score of 0.739 ($p < 0.001$).

Discussion

H. pylori infection is the most popular chronic bacterial infection worldwide, frequently associated with a wide range of gastrointestinal and extraintestinal disorders¹⁶. The role of *H. pylori* in the pathogenesis of chronic atrophic gastritis, peptic ulcer disease, and functional dyspepsia, along with its links to various malignancies, autoimmune conditions, and other conditions, significantly influences our understanding and management approaches for these diseases¹⁷⁻¹⁹. This extensive impact underscores the need for precise diagnostic measures.

To accurately diagnose *H. pylori* infection, healthcare professionals have access to a range of both invasive and non-invasive techniques. However, no single test emerges as the definitive standard²⁰. Invasive diagnostic procedures, such as histology, culture, and rapid urease tests, involve obtaining gastric biopsy specimens during gastro-duodenoscopy, which, while effective, may carry risks and cause discomfort to patients^{21,22}. Con-

versely, non-invasive diagnostic methods include urea breath tests, stool antigen tests, and serological assays¹⁵. Among these, recent advancements have highlighted the efficacy of RT-PCR in fecal samples, noted for its high sensitivity and specificity²³. Specifically, the *23SrRNA* gene is utilized as a precise marker in RT-PCR-based fecal analyses for detecting *H. pylori* infection²⁴. Our research confirmed this by identifying *H. pylori* in 93.1% of stool samples and 93.9% of gastric biopsy samples. The RT-PCR test demonstrated outstanding sensitivity (99.1%), specificity (100%), and diagnostic accuracy (99.1%), emphasizing its reliability. This is further supported by the strong concordance between the stool and gastric biopsy results (Kappa=0.929, $p < 0.001$).

A critical aspect of managing *H. pylori* infection involves monitoring its antibiotic susceptibility, which is important to ensure the efficacy of treatment regimens. However, routine susceptibility testing is not commonly conducted due to its invasive nature, the limited availability of culture facilities, and cost considerations. Clarithromycin, a macrolide antibiotic, plays a pivotal role in *H. pylori* eradication by inhibiting protein synthesis²⁵. Nonetheless, the increasing resistance to clarithromycin, primarily due to mutations in the *23SrRNA* ribosomal component, poses a significant challenge and frequently leads to treatment failures²⁶. Empirical first-line treatments that incorporate clarithromycin have shown disappointing success rates, with only approximately 18% achieving an eradication threshold above 85%, while about 60% fail to reach even 80% effectiveness²⁷. Resistance to clarithromycin, metronidazole, and levofloxacin is highly prevalent, complicating the development of effective treatment strategies^{28,29}. Our findings indicate resistance rates of 37.3% for clarithromycin and 32.1% for levofloxacin in stool samples, figures that closely mirror those observed in gastric samples. The consistency between RT-PCR results from both types of samples validates the stool RT-PCR test as an effective method for assessing resistance to these antibiotics.

Conclusions

Routine culture and antimicrobial susceptibility testing are rarely conducted in clinical practice, highlighting the need for continuous research into *H. pylori* resistance to improve treatment methods. A novel molecular test utilizing RT-PCR technology has emerged as a critical tool in this effort. This non-invasive method provides an accurate diagnosis of *H. pylori* infection and identifies resistance to clarithromycin and levofloxacin. These capabilities are crucial for enhancing patient compliance, ensuring prompt and precise treatment interventions, improving the success rates of *H. pylori* eradication, and thereby reducing the risk of associated diseases.

Conflict of Interest

The authors declare that they have no conflict of interest.

Availability of Data and Materials

The datasets generated and analyzed during the current study are available from the corresponding author upon reasonable request. The datasets are also available in the Chinese Clinical Trial Registry.

Ethics Approval

The study was conducted in accordance with the Helsinki Declaration and has been approved by the Institutional Ethics Board of the Civil Aviation General Hospital (registration number 2023-L-K-05). This study followed the Consolidated Standards of Reporting Trials (CONSORT) guidelines.

Clinical Trial Registration

The trial was registered in the Chinese Clinical Trials Registration (available at: www.chictr.org.cn) with the registration number ChiCTR2300070267.

Informed Consent

We obtained informed consent from patients or their immediate family members.

Authors' Contributions

CJP: study design, data analysis, data interpretation, and manuscript writing. BQX: study design and critical revision of the manuscript. LZ, ZLL, WH, ZXL, XDL, CY, and DHO: material preparation, data collection. FCJ and HK: patient recruitment, study coordination, and study supervision. All authors have read and approved the submitted manuscript.

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References

- 1) Ailloud F, Didelot X, Woltemate S, Pfaffinger G, Overmann J, Bader RC, Schulz C, Malfertheiner P, Suerbaum S. Within-host evolution of *Helicobacter pylori* shaped by niche-specific adaptation, intragastric migrations and selective sweeps. *Nat Commun* 2019; 10: 2273.
- 2) Verma V, Kumar A, Nitharwal RG, Alam J, Mukhopadhyay AK, Dasgupta S, Dhar SK. Modulation of the enzymatic activities of replicative helicase (DnaB) by interaction with Hp0897: a possible mechanism for helicase loading in *Helicobacter pylori*. *Nucleic Acids Res* 2016; 44: 3288-3303.
- 3) Codolo G, Toffoletto M, Chemello F, Coletta S, Soler Teixidor G, Battaglia G, Munari G, Fassan M, Cagnin S, de Bernard M. *Helicobacter pylori* dampens HLA-II expression on macrophages via the up-regulation of miRNAs targeting CIITA. *Front Immunol* 2020; 10: 2923.
- 4) Lv YP, Cheng P, Zhang JY, Mao FY, Teng YS, Liu YG, Kong H, Wu XL, Hao CJ, Han B, Ma Q, Yang SM, Chen W, Peng LS, Wang TT, Zou QM, Zhuang Y. *Helicobacter pylori*-induced matrix metalloproteinase-10 promotes gastric bacterial colonization and gastritis. *Sci Adv* 2019; 5: eaau6547.
- 5) Duan Y, Dou Y, Xu D. How Can We Better Understand the Gastric Carcinogenesis of *Helicobacter pylori*? *Gastroenterology* 2022; 163: 1121-1122.
- 6) Bauer M, Nascakova Z, Mihai AI, Cheng PF, Levesque MP, Lampart S, Hurwitz R, Pfannkuch L, Dobrovolna J, Jacobs M, Bartfeld S, Dohlman A, Shen X, Gall AA, Salama NR, Töpfer A, Weber A, Meyer TF, Janscak P, Müller A. The ALPK1/TIFA/NF- κ B axis links a bacterial carcinogen to R-loop-induced replication stress. *Nat Commun* 2020; 11: 5117.
- 7) Keskin M, Yavuz A. A novel rapid and accurate method for detecting *Helicobacter pylori*: the modified antigen test. *Eur Rev Med Pharmacol Sci* 2022; 26: 1148-1155.
- 8) Qiu E, Li Z, Han S. Methods for detection of *Helicobacter pylori* from stool sample: current options and developments. *Braz J Microbiol* 2021; 52: 2057-2062.
- 9) Van den Poel B, Gils S, Micalessi I, Carton S, Christiaens P, Cuyle PJ, Moons V, Van Olmen G, Smismans A, Bourgain C, Bossuyt P, Frans J. Molecular detection of *Helicobacter pylori* and clarithromycin resistance in gastric biopsies: a prospective evaluation of RIDA[®]GENE Helico-

- bacter pylori assay. *Acta Clin Belg* 2021; 76: 177-183.
- 10) Kovacheva-Slavova M, Valkov H, Angelov T, Tropcheva R, Vladimirov B. Screening for Helicobacter pylori infection and clarithromycin resistance using real-time polymerase chain reaction. *Eur Rev Med Pharmacol Sci* 2021; 25: 5042-5046.
 - 11) Iannone A, Giorgio F, Russo F, Riezzo G, Girardi B, Pricci M, Palmer SC, Barone M, Principi M, Strippoli GF, Di Leo A, Ierardi E. New fecal test for non-invasive Helicobacter pylori detection: a diagnostic accuracy study. *World J Gastroenterol* 2018; 24: 3021-3029.
 - 12) Dogan A, Ekinçi O, Ebinc S. Effect of Helicobacter pylori infection on the first-line treatment outcomes in patients with immune thrombocytopenic purpura. *Eur Rev Med Pharmacol Sci* 2022; 26: 3995-4000.
 - 13) Li Y, Lv T, He C, Wang H, Cram DS, Zhou L, Zhang J, Jiang W. Evaluation of multiplex ARMS-PCR for detection of Helicobacter pylori mutations conferring resistance to clarithromycin and levofloxacin. *Gut Pathog* 2020; 12: 35.
 - 14) Liu WZ, Xie Y, Lu H, Cheng H, Zeng ZR, Zhou LY, Chen Y, Wang JB, Du YQ, Lu NH. Fifth Chinese National Consensus Report on the management of Helicobacter pylori infection. *Helicobacter* 2018; 23: e12475.
 - 15) Chung WC, Jung SH, Oh JH, Kim TH, Cheung DY, Kim BW, Kim SS, Kim JI, Sin E. Dual-priming oligonucleotide-based multiplex PCR using tissue samples in rapid urease test in the detection of Helicobacter pylori infection. *World J Gastroenterol* 2014; 20: 6547-6553.
 - 16) Shin SP, Bang CS, Lee JJ, Baik GH. Helicobacter pylori infection in patients with chronic kidney disease: a systematic review and meta-analysis. *Gut Liver* 2019; 13: 628-641.
 - 17) Chiang TH, Chang WJ, Chen SL, Yen AM, Fann JC, Chiu SY, Chen YR, Chuang SL, Shieh CF, Liu CY, Chiu HM, Chiang H, Shun CT, Lin MW, Wu MS, Lin JT, Chan CC, Graham DY, Chen HH, Lee YC. Mass eradication of Helicobacter pylori to reduce gastric cancer incidence and mortality: a long-term cohort study on Matsu Islands. *Gut* 2021; 70: 243-250.
 - 18) Isobe T, Hashimoto K, Kizaki J, Miyagi M, Aoyagi K, Koufujii K, Shirouzu K. Characteristics and prognosis of gastric cancer in young patients. *Oncol Rep* 2013; 30: 43-49.
 - 19) Ng QX, Foo NX, Loke W, Koh YQ, Seah VJM, Soh AYS, Yeo WS. Is there an association between Helicobacter pylori infection and irritable bowel syndrome? A meta-analysis. *World J Gastroenterol* 2019; 25: 5702-5710.
 - 20) Zhao J, Xu S, Gao Y, Lei Y, Zou B, Zhou M, Chang D, Dong L, Qin B. Accuracy of endoscopic diagnosis of Helicobacter pylori based on the Kyoto classification of gastritis: a multicenter study. *Front Oncol* 2020; 10: 599218.
 - 21) Choi YI, Chung JW, Park DK, Kim KO, Kwon KA, Kim YJ, Seo JY. Tailored eradication vs empirical bismuth-containing quadruple therapy for first-line Helicobacter pylori eradication: a comparative, open trial. *World J Gastroenterol* 2019; 25: 6743-6751.
 - 22) Beckman E, Saracino I, Fiorini G, Clark C, Slepnev V, Patel D, Gomez C, Ponaka R, Elagin V, Vaira D. A novel stool PCR test for Helicobacter pylori may predict clarithromycin resistance and eradication of infection at a high rate. *J Clin Microbiol* 2017; 55: 2400-2405.
 - 23) Jung DH, Kim JH, Jeong SJ, Park SY, Kang IM, Lee KH, Song YG. Peptide nucleic acid probe-based analysis as a new detection method for clarithromycin resistance in Helicobacter pylori. *Gut Liver* 2018; 12: 641-647.
 - 24) Zhang XY, Shen WX, Chen CF, Sheng HH, Cheng H, Li J, Hu F, Lu DR, Gao HJ. Detection of the clarithromycin resistance of Helicobacter pylori in gastric mucosa by the amplification refractory mutation system combined with quantitative real-time PCR. *Cancer Med* 2019; 8: 1633-1640.
 - 25) Zhou Q, Zhu LL, Yan XF, Pan WS, Zeng S. Drug utilization of clarithromycin for gastrointestinal disease treatment. *World J Gastroenterol* 2008; 14: 6065-6071.
 - 26) Kuo CJ, Chen CW, Le PH, Hsu JT, Lin CY, Cheng HT, Su MY, Lin CJ, Chiu CT. Efficacy of dexlansoprazole-based triple therapy for Helicobacter pylori infections. *Therap Adv Gastroenterol* 2019; 12: 1756284819870960.
 - 27) Matta AJ, Zambrano DC, Pazos AJ. Punctual mutations in 23S rRNA gene of clarithromycin-resistant Helicobacter pylori in Colombian populations. *World J Gastroenterol* 2018; 24: 1531-1539.
 - 28) Hu Y, Zhu Y, Lu NH. Novel and effective therapeutic regimens for Helicobacter pylori in an Era of increasing antibiotic resistance. *Front Cell Infect Microbiol* 2017; 7: 168.
 - 29) Saranathan R, Levi MH, Wattam AR, Malek A, Asare E, Behin DS, Pan DH, Jacobs WR, Jr., Szymczak WA. Helicobacter pylori infections in the Bronx, New York: surveying antibiotic susceptibility and strain lineage by whole-genome sequencing. *J Clin Microbiol* 2020; 58: e01591-19.