

A review on irisin, a new protagonist that mediates muscle–adipose–bone–neuron connectivity

B. GRYGIEL-GÓRNIAK, M. PUSZCZEWICZ

Department of Rheumatology and Internal Medicine, Poznan University of Medical Sciences, Poznan, Poland

Abstract. – Physical activity improves the quality of life and decreases the risk of several diseases (i.e. stroke, hypertension, myocardial infarction, obesity, and malignancies). Skeletal muscles are considered as an endocrine organ that produces myokines characterized by a paracrine or endocrine activity. Irisin is a circulating hormone-like myokine and is secreted as a product of fibronectin type III domain-containing protein 5 from skeletal muscle in response to exercise. This molecule regulates the energy metabolism and acts in adipose tissue, bones, and nervous system. As both animal and clinical studies confirmed the action of irisin in muscle and adipocytes, this protein is considered as adipomyokine. In adipose tissue, irisin stimulates the process of browning of beige precursor fat cells, which are present in white fat cells, and promotes energy expenditure. It affects bone metabolism by increasing osteoblast differentiation and reducing osteoclast maturation. In nervous system, irisin influences hippocampal neurogenesis and neural differentiation of embryonic stem cells in mice and is considered as a messenger between exercise and brain function. However, the existence of this protein and its role in humans is a matter of debate. This study presents irisin as a new champion of the molecule, which could be considered as the messenger in the muscle-fat-bone-brain axis.

Key Words:

Irisin, Adipomyokine, Metabolic, Neurologic influence.

Introduction

With aging, increased tendency toward sarcopenic obesity is observed¹. Sarcopenia is a medical condition, which is defined by decreased muscle mass and impaired muscle function, which may be associated with frailty. To maintain a proper functioning of muscles, it is important to have

an adequate physical activity, which is considered to be as the best nonpharmacological treatment for cardiovascular, metabolic, and bone diseases^{2,3}. During exercise, the endocrine activity of muscles releases myokines, which are the crucial molecules in the regulation of energy homeostasis^{4,5}. Boström et al^{4,6} found that physical activity induces transcriptional regulator peroxisome proliferator-activated receptor- γ co-activator 1 α (PGC-1 α) in the skeletal muscle. PGC-1 α is responsible for the synthesis of fibronectin type III domain-containing protein 5 (FNDC5), which is a membrane protein abundantly expressed in skeletal muscle. Structurally, FNDC5 is a protein that contains a signal peptide and fibronectin type III repeats (Figure 1). The C-terminal tail of FNDC5 is in the cytoplasm, whereas the extracellular N-terminal part is released into the circulation as irisin during proteolytic cleavage^{4,6}. The term “irisin” was proposed by Boström et al⁴ after Iris, the courier goddess, to underline the function of irisin as a messenger targeting the endocrine signal from skeletal muscle to adipose tissue. In animal models, increased muscle mRNA expression and enhanced irisin plasma concentrations (65%) are observed^{4,7}. In humans, irisin circulates at concentrations typical of hormones⁸. It induces the browning process of white adipose tissue (the appearance of “bright” cells in white adipose depots) and increases thermogenesis⁹⁻¹². It influences energy homeostasis, and it is one of the regulators of adipocyte metabolism. However, its function depends on the actual doses of irisin secreted by cells or administered synthetically. Browning of adipose tissue occurs when large amounts of recombinant irisin are administered (3,500 $\mu\text{g kg}^{-1}/\text{week}$); but at lower doses, it modulates the skeletal genes expression (*Opn* and *Sost gene*)¹³. Irisin also improves the geometry and strength of cortical bone, specifically when trabecular bone

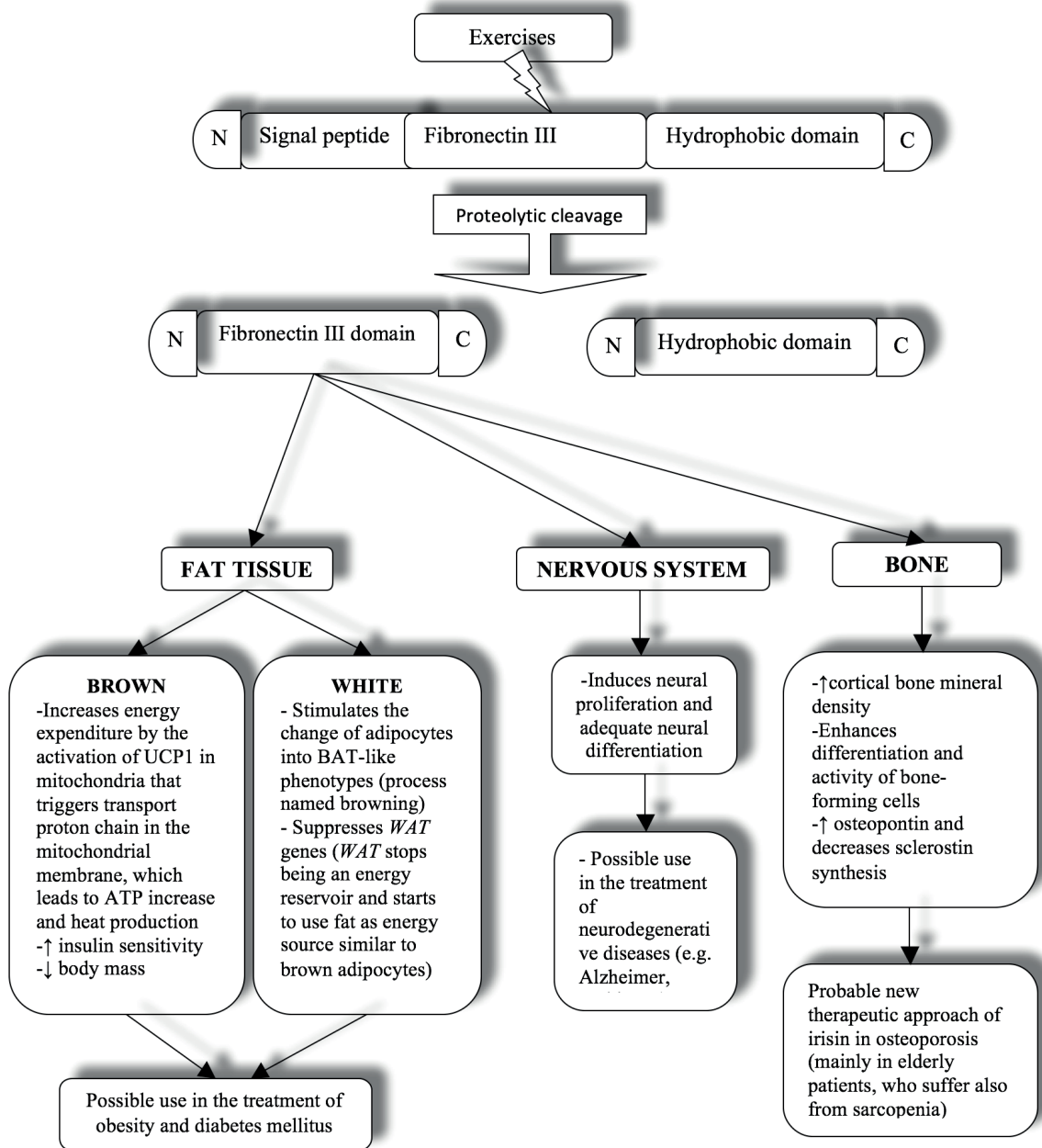


Figure 1. Activation of irisin and its role on the muscle–fat–bone–brain axis (FNDC5 is proteolytically cleaved and contains an amino (N)-terminal part and a hydrophobic domain inserted in the lipidic bilayer at carboxy (C)-terminal domain).

(which is a calcium reservoir) is spared^{14,15}. It is also expressed in brain in low amounts^{16,17}, and its concentration differs with age (inverse correlation) and gender (higher amounts of irisin were reported in young men compared with middle-aged women)¹⁸. This study summarizes the most relevant research findings on the role of irisin in muscles, adipose tissue, bones, and brain. As the exact role of irisin in humans is still a matter of

debate, we decided to present recent data regarding this interesting molecule and describe its activity in different body parts.

Irisin and Skeletal Muscles

Irisin is a hormone secreted by myocytes and is proposed to be a bridge between exercise and metabolic homeostasis^{4,12,18}. The level of irisin positively correlates with the amount of muscle

mass^{18,19}. In animals, plasma irisin levels are increased by 65% after 3 weeks of freewheel running, whereas in healthy humans, irisin concentration is doubled after 10 weeks of endurance exercises⁴. An increased level of irisin is also reported in male athletes, 30 minutes after sprinting, but this effect disappears after 8 weeks of training¹⁸. However, some data did not confirm FNDC5 gene activation by aerobic exercises in younger humans²⁰, and regular exercises were inversely correlated with irisin concentration in adult men²¹. These discrepancies have been explained by the fact that the level of irisin increases only when more energy is needed, and such condition is observed in sedentary lifestyle, in which ATP concentration is strongly decreased in muscles without physical activity¹⁸.

Irisin and Adipose Tissue

Irisin is not only a myokine but also an adipokine, which serves as a mediator for the beneficial metabolic effects of exercises⁴. In adipose tissue, irisin stimulates the browning of white adipocytes and thermogenesis by the activation of uncoupling protein 1 (UCP1)^{4,22}. Brown adipose tissue produces energy through thermogenesis, improving energy homeostasis. Also, irisin inhibits adipogenesis and highlights the potential role of direct signaling from skeletal muscle in brown adipocytes²³. Not only irisin but also brown fat has a positive influence on the metabolic processes and increases insulin sensitivity and total body energy expenditure, which enables the reduction of body mass⁴. Most of the studies describe the positive correlation between irisin and adiposity^{18,24}; however, some researches indicate the negative relation between irisin and the amount of fat tissue²⁵. In obese people, elevated irisin level is positively associated with the resting energy expenditure¹⁹. Decreased concentration of irisin is observed after bariatric surgery¹⁸ or in anorexia nervosa²⁶, but exercise and lifestyle changes for 1 year, result in the increase of plasma irisin level in obese children²⁷. In healthy women, a positive tendency between the circulating irisin and BMI is observed^{18,24}; however, this relation disappears after adjusting for the potential confounding factors¹⁸. Overexpression of irisin causes an increase in glucose tolerance and improvement in insulin sensitivity in high-fat-fed mice²⁸. In patients with type 2 diabetes mellitus, serum irisin was found to be negatively associated with hyperglycemia²⁹, and the level of serum irisin in diabetic patients is lower than that in nondiabetic population³⁰. However, some researchers observed an elevated

level of irisin in obesity, diabetes mellitus, or metabolic syndrome and suggested the physiological response of this molecule to the improvement of glucose tolerance and lipid profile in patients with these diseases^{18,21}. Besides metabolic properties, irisin has a pro-proliferation effect and stimulates the proliferation of endothelial cells of humans³¹. Further studies are needed to elucidate the complexity of irisin interactions with the metabolic endpoints of these facts; however, the role of irisin as a messenger between exercise and metabolic homeostasis may be a reason for the possibility to use it in the therapeutic processes in patients with obesity or diabetes mellitus.

Irisin and Bone Mass

Physical exercise has a beneficial influence on bone metabolism, because cortical bone is highly sensitive to anabolic factors released by muscle^{32,33}. In many studies, positive associations between exercise and increased bone size and bone mass are confirmed³²⁻³⁶. Higher muscle mass is associated with a higher BMD and reduced risk of fracture in postmenopausal women³⁷. Animal and clinical researches have shown some beneficial influence of myokines on bone turnover mediated through myokines⁵. Irisin, released upon physical activity, displays anabolic actions on the skeleton¹⁵. Indeed, in athletes, irisin level correlates positively with BMD and bone strength³⁸, but because of decreased physical activity in former athletes, the irisin level leads to a progressive loss of bone³⁴. Bone tissue is more sensitive to the irisin action than adipose tissue, and the effect of irisin depends on the administered dose¹³. Recombinant irisin (r-irisin), given at low dose (100 $\mu\text{g kg}^{-1}/\text{week}$) in young mice, increases cortical bone mineral density and positively modifies bone geometry, thus pro-osteoblastic genes are upregulated and the activity of osteoblasts increases. Simultaneously, osteoblast inhibitors (e.g. *Spp1* and *Sost* gene) are decreased¹³. These genes are responsible for mechanostimulation¹⁵, and one-month treatment of r-irisin causes increased expression of *Spp1* on cortical surfaces^{39,40}. Irisin enhances the level of osteopontin (an osteocyte-specific protein and a component of the bone extracellular matrix), which is a regulator of biomineralization in bone tissue, and decreases the bone formation⁴¹. Moreover, recombinant irisin injections downregulate sclerostin, a molecule, which decreases the bone formation^{42,43}. Similar data are reported in humans, in which plasma irisin level is inversely

correlated with sclerostin concentration independently of age or sex⁴⁴. In addition, circulating irisin is inversely correlated with sclerostin, which not only influences the bone metabolism but also is related to the adiposity and glucose metabolism⁴⁴. Sclerostin plays a role in the formation of bone and fat cell⁴⁵. It increases with aging and is higher in the elderly compared with younger people. The negative association between irisin and sclerostin⁴⁴ may be because of the influence of irisin on the stimulation of the browning process of adipocytes⁴⁶. Besides dose-dependent activity, irisin influences the bone metabolism directly¹⁴ or indirectly by brown fat-mediated effects. The direct effects are related to increased osteoblastogenesis (increased differentiation of bone marrow stromal cells into mature osteoblasts)¹⁴ via the Wnt/beta-catenin pathway downstream of the BMP receptor signal and decreased differentiation of osteoclasts^{47,48}. Studies *in vivo* showed increased bone trabecular volume, cortical bone mass, geometry, and strength exhibited by irisin in mice^{15,47}. Irisin promotes osteogenesis during lineage-specific differentiation⁴⁷. Similar results are reported in humans, in whom irisin levels in young athletes are predicted to increase bone density and strength³⁸. Indirect effect of irisin in bones is mediated through the stimulation of adipocyte browning. As mentioned earlier, high doses of recombinant irisin (3,500 µg kg⁻¹/week) affect uncoupling protein 1 (UCP-1) expression in white adipose tissue, whose upregulation causes browning of fat¹⁵. Brown adipose tissue has a beneficial influence on the skeleton^{26, 46, 49} and shows anabolic effect on bones⁴⁶. On the contrary, low dose of irisin does not induce browning of white adipose tissue, and it increases the molecular markers of browning (*UCP1*) or adipogenesis (*PPARγ*)⁴⁸. Moreover, brown adipocyte-synthesized factors may induce osteoblast differentiation in bone marrow, for example, insulin-like growth factor binding protein 2 (IGFBP2)⁵⁰. The association between brown fat and bone strength was proved in women⁵¹, as well as in children and adolescents (increased with cortical thickness)⁵². Moreover, in adult women, lower irisin levels were associated with a higher risk of osteoporotic fractures⁵³. Irisin can be used as a therapeutic molecule in obesity and exercise-associated bone formation^{54,55}.

The risk of osteoporosis is related to age and gender (the increase of osteoporosis is observed after menopause), which is partially related to hormonal changes. Estradiol level can influence

serum sclerostin and irisin synthesis^{18,56}, and in postmenopausal overweight women with osteoporosis, an inverse correlation between irisin levels and vertebral fragility and fractures is observed⁵⁷. Positive correlations between irisin and estradiol level, muscle mass, and insulin sensitivity are reported^{18,25}. On the contrary, older age and increased fat mass, as well as insulin resistance and elevated cholesterol, are negatively correlated with irisin^{18,21,25}. Moreover, strong positive correlation between irisin and 24-hour energy expenditure was noted, but only in the group of women with energy expenditure higher than predicted⁵⁸. Recent studies have shown that sclerostin and irisin are partly regulated by estrogen^{18,59}. There is also an inverse correlation between serum irisin levels and vertebral fragility or fractures in postmenopausal women⁵³. As irisin has beneficial effect in sarcopenia and osteoporosis, new therapeutic approach of this molecule should be considered, particularly in older people, who often suffer from both the diseases.

Irisin and Nervous System

Irisin is also suspected to be the molecular link between muscle and brain. It is expressed in cerebellar Purkinje cells of rats and mice⁶⁰ and is necessary for the proper neural differentiation of mice embryonic stem cells¹⁷. It is suspected that there is a neural pathway participating in the signal transduction of irisin, produced in cerebellum, to adipocytes through intermediary synapses in the medulla and spinal cord⁶⁰. Irisin also influences hippocampal neurogenesis in a dose-dependent manner (high concentrations of irisin 50-100 nmol/L increases the proliferation of neurons)¹⁶. Because of the positive effect of exercises on neurogenesis and the fact that hippocampus is considered as the main region affected by neurodegenerative diseases (Alzheimer's or Parkinson's disease), irisin is regarded as the molecular messenger between exercise and brain function⁶¹⁻⁶³. Data also suggest that some medications can affect the irisin concentration. Recently, statins have been reported to exhibit a crucial influence on both lipid metabolism and irisin concentration⁶⁴, and simvastatin has been shown to increase irisin concentration⁶⁵. Beside this, the expression of *FNDC5* can also be induced by nutritional factors including polyunsaturated fatty acids (PUFA). The supplementation of omega 3 fatty acids during 24 and 48h has significantly induced irisin expression in human rhabdomyosarcoma cells⁶⁶.

Conclusions

Irisin shows a strong relation between skeletal muscle, adipocytes, bone, and neurons, which better explains the comprehensive regulation of skeletal homeostasis exerted by physical exercises. This specific crosstalk between fat, bone, muscle, and neurons shows the significant role of this molecule in the metabolic processes of our body. Utilizing the metabolic and the osteogenic potential of irisin might be a therapeutic choice in treating certain diseases caused by inactive lifestyle. Irisin can play a potential role in pharmacological treatment and can be the representative of a new anabolic therapy in high-burden diseases such as osteopenia/osteoporosis, obesity/sarcopenia, and tumor-associated cachexia. These disorders often occur concurrently and contribute to increased morbidity/mortality and impaired quality of life. Thus, the beneficial role of irisin in many metabolic and osteogenic processes confirms that physical exercise is necessary for the development of an efficient load-bearing skeleton, browning process in white adipose tissue, and proper function of neurons. Many studies underline the positive role of irisin in metabolic processes; however, the therapeutic potential of this molecule is temporarily questioned. The animal studies have shown an undeniable thermogenic effect of irisin; but in humans, this effect remains controversial. Additional studies with the larger sample size are needed to systematically identify these interacting links and their causal relations. Also, regular exercises are crucial for everyday health and quality of life.

Conflict of interest

The authors declare no conflicts of interest.

References

- MOLINO S, DOSSENA M, BUONOCORE D, VERRI M. Sarcopenic obesity: an appraisal of the current status of knowledge and management in elderly people. *J Nutr Health Aging* 2016; 20: 780-788.
- DUNSTAN D. Diabetes: exercise and T2DM-move muscles more often! *Nat Rev Endocrinol* 2011; 7: 189-190.
- CREPALDI G, MAGGI S. Sarcopenia and osteoporosis: a hazardous duet. *J Endocrinol Invest* 2005; 28: 66-68.
- BOSTRÖM P, WU J, JEDRYCHOWSKI MP, KORDE A, YE L, LO JC, RASBACH KA, BOSTRÖM EA, CHOI JH, LONG JZ, KAJIMURA S, ZINGARETTI MC, VIND BF, TU H, CINTI S, HØJLUND K, GYGI SP, SPIEGELMAN BM. A PGC1- α -dependent myokine that drives brown-fat-like development of white fat and thermogenesis. *Nature* 2012; 481: 463-468.
- LOMBARDI G, SANCHIS-GOMAR F, PEREGO S, SANSONI V, BANFI G. Implications of exercise-induced adipomyokines in bone metabolism. *Endocrine* 2016; 54: 284-305.
- WRANN CD, WHITE JP, SALOGIANNIS J, LAZNIK-BOGOSLAVSKI D, WU J, MA D, LIN JD, GREENBERG ME, SPIEGELMAN BM. Exercise induces hippocampal BDNF through a PGC1 α /FNDC5 pathway. *Cell Metab* 2013; 18: 649-659.
- DE MATTEIS R, LUCERTINI F, GUESCINI M, POLIDORI E, ZEPPA S, STOCCHI V, CINTI S, CUPPINI R. Exercise as a new physiological stimulus for brown adipose tissue activity. *Nutr Metab Cardiovasc Dis* 2013; 23: 582-590.
- JEDRYCHOWSKI MP, WRANN CD, PAULO JA, GERBER KK, SZPYT J, ROBINSON MM, NAIR KS, GYGI SP, SPIEGELMAN BM. Detection and quantitation of circulating human irisin by tandem mass spectrometry. *Cell Metab* 2015; 22: 734-740.
- BARBATELLI G, MURANO I, MADSEN L, HAO Q, JIMENEZ M, KRISTIANSEN K, GIACOBINO JP, DE MATTEIS R, CINTI S. The emergence of cold-induced brown adipocytes in mouse white fat depots is determined predominantly by white to brown adipocyte transdifferentiation. *Am J Physiol Endocrinol Metab* 2010; 298: E1244-1253.
- ROSEN ED, SPIEGELMAN BM. What we talk about when we talk about fat. *Cell* 2014; 156: 20-44.
- LEE P, LINDERMAN JD, SMITH S, BRYCHTA RJ, WANG J, IDELSON C, PERRON RM, WERNER CD, PHAN GO, KAMMULA US, KEBEBEW E, PACAK K, CHEN KY, CELI FS. Irisin and FGF21 are cold-induced endocrine activators of brown fat function in humans. *Cell Metab* 2014; 19: 302-309.
- LECKER SH, ZAVIN A, CAO P, ARENA R, ALLSUP K, DANIELS KM, JOSEPH J, SCHULZE PC, FORMAN DE. Expression of the irisin precursor FNDC5 in skeletal muscle correlates with aerobic exercise performance in patients with heart failure. *Circ Heart Fail* 2012; 5: 812-818.
- ROBLING AG, NIZIOLEK PJ, BALDRIDGE LA, CONDON KW, ALLEN MR, ALAM I, MANTILA SM, GLUHAK-HEINRICH J, BELLIDO TM, HARRIS SE, TURNER CH. Mechanical stimulation of bone in vivo reduces osteocyte expression of Sost/sclerostin. *J Biol Chem* 2008; 283: 5866-5875.
- COLAIANNI G, CUSCITO C, MONGELLI T, ORANGER A, MORI G, BRUNETTI G, COLUCCI S, CINTI S, GRANO M. Irisin enhances osteoblast differentiation *in vitro*. *Int J Endocrinol* 2014; 2014: 902186.
- COLAIANNI G, GRANO M. Role of Irisin on the bone-muscle functional unit. *Bonekey Rep* 2015; 23; 4: 765.
- MOON HS, DINCER F, MANTZOROS CS. Pharmacological concentrations of irisin increase cell proliferation without influencing markers of neurite outgrowth and synaptogenesis in mouse H19-7 hippocampal cell lines. *Metabolism* 2013; 62: 1131-1136.
- HASHEMI MS, GHAEDI K, SALAMIAN A, KARBALAEI K, EMADI-BAYGI M, TANHAEI S, NASR-ESFAHANI MH, BAHARVAND H. Fndc5 knockdown significantly decreased neu-

- ral differentiation rate of mouse embryonic stem cells. *Neuroscience* 2013; 231: 296-304.
- 18) HUH JY, PANAGIOTOU G, MOUGIOS V, BRINKOETTER M, VAMVINI MT, SCHNEIDER BE, MANTZOROS CS. FNDC5 and irisin in humans: I. Predictors of circulating concentrations in serum and plasma and II. mRNA expression and circulating concentrations in response to weight loss and exercise. *Metabolism* 2012; 61: 1725-1738.
 - 19) STENGEL A, HOFMANN T, GOEBEL-STENGEL M, ELBELT U, KOBELT P, KLAPP BF. Circulating levels of irisin in patients with anorexia nervosa and different stages of obesity-correlation with body mass index. *Peptides* 2013; 39: 125-130.
 - 20) TIMMONS JA, BAAR K, DAVIDSEN PK, ATHERTON PJ. Is irisin a human exercise gene? *Nature* 2012; 488: E9-10.
 - 21) PARK KH, ZAICHENKO L, BRINKOETTER M, THAKKAR B, SAHIN-EFE A, JOUNG KE, TSOUKAS MA, GELADARI EV, HUH JY, DINCER F, DAVIS CR, CROWELL JA, MANTZOROS CS. Circulating irisin in relation to insulin resistance and the metabolic syndrome. *J Clin Endocrinol Metab* 2013; 98: 4899-4907.
 - 22) FRONTINI A, CINTI S. Distribution and development of brown adipocytes in the murine and human adipose organ. *Cell Metabolism* 2010; 11: 253-256.
 - 23) CEREJO R, VILLARROYA J, VILLARROYA F. Non-sympathetic control of brown adipose tissue. *Int J Obes Suppl* 2015; 5: S40-44.
 - 24) CRUJEIRAS AB, PARDO M, ARTURO RR, NAVAS-CARRETERO S, ZULET MA, MARTÍNEZ JA, CASANUEVA FF. Longitudinal variation of circulating irisin after an energy restriction-induced weight loss and following weight regain in obese men and women. *Am J Hum Biol* 2014; 26: 198-207.
 - 25) MORENO-NAVARRETE JM, ORTEGA F, SERRANO M, GUERRA E, PARDO G, TINAHONES F, RICART W, FERNÁNDEZ-REAL JM. Irisin is expressed and produced by human muscle and adipose tissue in association with obesity and insulin resistance. *J Clin Endocrinol Metab* 2013; 98: E769-778.
 - 26) BREDELLA MA, FAZELI PK, FREEDMAN LM, CALDER G, LEE H, ROSEN CJ, KLIBANSKI A. Young women with cold-activated brown adipose tissue have higher bone mineral density and lower Pref-1 than women without brown adipose tissue: a study in women with anorexia nervosa, women recovered from anorexia nervosa, and normal-weight women. *J Clin Endocrinol Metab* 2012; 97: E584-590.
 - 27) BLÜHER S, PANAGIOTOU G, PETROFF D, MARKERT J, WAGNER A, KLEMM T, FILIPPAIOS A, KELLER A, MANTZOROS CS. Effects of a 1-year exercise and lifestyle intervention on irisin, adipokines, and inflammatory markers in obese children. *Obesity (Silver Spring)* 2014; 22: 1701-1708.
 - 28) ZHU G, WANG J, SONG M, ZHOU F, FU D, RUAN G, ZHU X, BAI Y, HUANG L, PANG R, KANG H, PAN X. Irisin increased the number and improved the function of endothelial progenitor cells in diabetes mellitus mice. *J Cardiovasc Pharmacol* 2016; 68: 67-73.
 - 29) KURDIOVA T, BALAZ M, VICIAN M, MADEROVA D, VLCEK M, VALKOVIC L, SRBECKY M, IMRICH R, KYSELOVICOVA O, BELAN V, JELOK I, WOLFRUM C, KLIMES I, KRSSAK M, ZEMKOVA E, GASPERIKOVA D, UKROPEC J, UKROPCOVA B. Effects of obesity, diabetes and exercise on Fndc5 gene expression and irisin release in human skeletal muscle and adipose tissue: in vivo and in vitro studies. *J Physiol* 2014; 592: 1091-1107.
 - 30) LIU JJ, WONG MD, TOY WC, TAN CS, LIU S, NG XW, TAVINTHARAN S, SUM CF, LIM SC. Lower circulating irisin is associated with type 2 diabetes mellitus. *J Diabetes Complications* 2013; 27: 365-369.
 - 31) SONG H, WU F, ZHANG Y, ZHANG Y, WANG F, JIANG M, WANG Z, ZHANG M, LI S, YANG L, WANG XL, CUI T, TANG D. Irisin promotes human umbilical vein endothelial cell proliferation through the ERK signaling pathway and partly suppresses high glucose-induced apoptosis. *PLoS One* 2014; 9: e110273.
 - 32) DUCHER G, BASS SL, SAXON L, DALY RM. Effects of repetitive loading on the growth-induced changes in bone mass and cortical bone geometry: a 12-month study in pre/peri- and postmenarcheal tennis players. *J Bone Miner Res* 2011; 26: 1321-1329.
 - 33) JOHANNESDOTTIR F, ASPELUND T, SIGGEISDOTTIR K, JONSSON BY, MOGENSEN B, SIGURDSSON S, HARRIS TB, GUDNASON VG, LANG TF, SIGURDSSON G. Mid-thigh cortical bone structural parameters, muscle mass and strength, and association with lower limb fractures in older men and women (AGES-Reykjavik Study). *Calcif Tissue Int* 2012; 90: 354-364.
 - 34) ANDREOLI A, CELI M, VOLPE SL, SORGE R, TARANTINO U. Long-term effect of exercise on bone mineral density and body composition in post-menopausal ex-elite athletes: a retrospective study. *Eur J Clin Nutr* 2012; 66: 69-74.
 - 35) MORI T, OKIMOTO N, SAKAI A, OKAZAKI Y, NAKURA N, NOTOMI T, NAKAMURA T. Climbing exercise increases bone mass and trabecular bone turnover through transient regulation of marrow osteogenic and osteoclastogenic potentials in mice. *J Bone Miner Res* 2003; 18: 2002-2009.
 - 36) HUANG TH, LIN SC, CHANG FL, HSIEH SS, LIU SH, YANG RS. Effects of different exercise modes on mineralization, structure, and biomechanical properties of growing bone. *J Appl Physiol* (1985) 2003; 95: 300-307.
 - 37) CLARKE BL, KHOSLA S. Physiology of bone loss. *Radiol Clin North Am* 2010; 48: 483-495.
 - 38) SINGHAL V, LAWSON EA, ACKERMAN KE, FAZELI PK, CLARKE H, LEE H, EDDY K, MARENGI DA, DERRICO NP, BOUXSEIN ML, MISRA M. Irisin levels are lower in young amenorrheic athletes compared with eumenorrheic athletes and non-athletes and are associated with bone density and strength estimates. *PLoS One* 2014; 9: e100218.
 - 39) TOMA CD, ASHKAR S, GRAY ML, SCHAFFER JL, GERSTENFELD LC. Signal transduction of mechanical stimuli is dependent on microfilament integrity: identification of osteopontin as a mechanically induced gene in osteoblasts. *J Bone Miner Res* 1997; 12: 1626-1636.
 - 40) ISHIIJIMA M, TSUJI K, RITTLING SR, YAMASHITA T, KUROSAWA H, DENHARDT DT, NIFUJI A, EZURA Y, NODA M. Osteopontin is required for mechanical stress-depen-

- dent signals to bone marrow cells. *J Endocrinol* 2007; 193: 235-243.
- 41) WESSON JA, JOHNSON RJ, MAZZALI M, BESHENSKY AM, STIETZ S, GIACHELLI C, LIAW L, ALPERS CE, COUSER WG, KLEINMAN JG, HUGHES J. Osteopontin is a critical inhibitor of calcium oxalate crystal formation and retention in renal tubules. *J Am Soc Nephrol* 2003; 14: 139-147.
 - 42) LIN C, JIANG X, DAI Z, GUO X, WENG T, WANG J, LI Y, FENG G, GAO X, HE L. Sclerostin mediates bone response to mechanical unloading through antagonizing Wnt/beta-catenin signaling. *J Bone Miner Res* 2009; 24: 1651-61.
 - 43) PASZTY C, TURNER CH, ROBINSON MK. Sclerostin: a gem from the genome leads to bone-building antibodies. *J Bone Miner Res* 2010; 25: 1897-1904.
 - 44) KLANGJAREONCHAI T, NIMITPHONG H, SAETUNG S, BHIROMMUANG N, SAMITTARUCKSA R, CHANPRASERTYOTHIN S, SUDATIP R, ONGPHIPHADHANAKUL B. Circulating sclerostin and irisin are related and interact with gender to influence adiposity in adults with prediabetes. *Int J Endocrinol* 2014; 2014: 261545.
 - 45) MOESTER MJ, PAPAPOULOS SE, LÖWIK CW, VAN BEZOOIJEN RL. Sclerostin: current knowledge and future perspectives. *Calcified Tissue International* 2010; 87: 99-107.
 - 46) RAHMAN S, LU Y, CZERNIK PJ, ROSEN CJ, ENERBACK S, LECKA-CZERNIK B. Inducible brown adipose tissue, or beige fat, is anabolic for the skeleton. *Endocrinology* 2013; 154: 2687-2701.
 - 47) ZHANG Y, LI R, MENG Y, LI S, DONELAN W, ZHAO Y, QI L, ZHANG M, WANG X, CUI T, YANG LJ, TANG D. Irisin stimulates browning of white adipocytes through mitogen-activated protein kinase p38 MAP kinase and ERK MAP kinase signaling. *Diabetes* 2014; 63: 514-525.
 - 48) ZHANG Y, XIE C, WANG H, FOSS R, CLARE M, GEORGE EV, LI S, KATZ A, CHENG H, DING Y, TANG D, REEVES WH, YANG LJ. Irisin exerts dual effects on browning and adipogenesis of human white adipocytes. *Am J Physiol Endocrinol Metab* 2016; 311: E530-541.
 - 49) MOTYL KJ, BISHOP KA, DEMAMBRO VE, BORNSTEIN SA, LE P, KAWAI M, LOTINUN S, HOROWITZ MC, BARON R, BOUXSEIN ML, ROSEN CJ. Altered thermogenesis and impaired bone remodeling in Misty mice. *J Bone Miner Res* 2013; 28: 1885-1897.
 - 50) CANNON B, NEDERGAARD J. Metabolic consequences of the presence or absence of the thermogenic capacity of brown adipose tissue in mice (and probably in humans). *Int J Obes (Lond)* 2010; 34: S7-16.
 - 51) LEE P, BRYCHTA RJ, COLLINS MT, LINDERMAN J, SMITH S, HERSCOVITCH P, MILLO C, CHEN KY, CELI FS. Cold-activated brown adipose tissue is an independent predictor of higher bone mineral density in women. *Osteoporos Int* 2013; 24: 1513-1518.
 - 52) PONRARTANA S, AGGABAO PC, HU HH, ALDROVANDI GM, WREN TA, GILSANZ V. Brown adipose tissue and its relationship to bone structure in pediatric patients. *J Clin Endocrinol Metab* 2012; 97: 2693-2698.
 - 53) ANASTASILAKIS AD, POLYZOS SA, MAKRAS P, GKIOMISI A, BISBINAS I, KATSAROU A, FILIPPAIOS A, MANTZOROS CS. Circulating irisin is associated with osteoporotic fractures in postmenopausal women with low bone mass but is not affected by either teriparatide or denosumab treatment for 3 months. *Osteoporos Int* 2014; 25: 1633-1642.
 - 54) GOUVEIA MC, VELLA JP, CAFFEO FR, AFFONSO FONSECA FL, BACCI MR. Association between irisin and major chronic diseases: a review. *Eur Rev Med Pharmacol Sci* 2016; 20: 4072-4077.
 - 55) LIU J. Irisin as an exercise-stimulated hormone binding crosstalk between organs. *Eur Rev Med Pharmacol Sci* 2015; 19: 316-321.
 - 56) MIRZA FS, PADHI ID, RAISZ LG, LORENZO JA. Serum sclerostin levels negatively correlate with parathyroid hormone levels and free estrogen index in postmenopausal women. *J Clin Endocrinol Metab* 2010; 95: 1991-1997.
 - 57) PALERMO A, STROLLO R, MADDALONI E, TUCCINARDI D, D'ONOFRIO L, BRIGANTI SI, DEFEUDIS G, DE PASCALIS M, LAZZARO MC, COLLELUORI G, MANFRINI S, POZZILLI P, NAPOLI N. Irisin is associated with osteoporotic fractures independently of bone mineral density, body composition or daily physical activity. *Clin Endocrinol (Oxf)* 2015; 82: 615-619.
 - 58) SWICK AG, ORENA S, O'CONNOR A. Irisin levels correlate with energy expenditure in a subgroup of humans with energy expenditure greater than predicted by fat free mass. *Metabolism* 2013; 62: 1070-1073.
 - 59) AL-DAGHRI NM, ALKHARFY KM, RAHMAN S, AMER OE, VINODSON B, SABICO S, PIYA MK, HARTE AL, McTERNAN PG, ALOKAIL MS, CHROUSOS GP. Irisin as a predictor of glucose metabolism in children: sexually dimorphic effects. *Eur J Clin Invest* 2014; 44: 119-124.
 - 60) DUN SL, LYU RM, CHEN YH, CHANG JK, LUO JJ, DUN NJ. Irisin-immunoreactivity in neural and non-neural cells of the rodent. *Neuroscience* 2013; 240: 155-162.
 - 61) SPIEGELMAN BM. Banting lecture 2012: regulation of adipogenesis: toward new therapeutics for metabolic disease. *Diabetes* 2013; 62: 1774-1782.
 - 62) MATTSON MP. Energy intake and exercise as determinants of brain health and vulnerability to injury and disease. *Cell Metabol* 2012; 16: 706-722.
 - 63) ERICKSON KI, WEINSTEIN AM, LOPEZ OL. Physical activity, brain plasticity, and Alzheimer's disease. *Arch Med Research* 2012; 43: 615-621.
 - 64) KOKKINOS PF, FASELIS C, MYERS J, PANAGIOTAKOS D, DOUMAS M. Interactive effects of fitness and statin treatment on mortality risk in veterans with dyslipidaemia: a cohort study. *Lancet* 2013; 381: 394-199.
 - 65) GOUNI-BERTHOLD I, BERTHOLD HK, HUH JY, BERMAN R, SPENRATH N, KRONE W, MANTZOROS CS. Effects of lipid-lowering drugs on irisin in human subjects in vivo and in human skeletal muscle cells ex vivo. *PLoS One* 2013; 8: e72858.
 - 66) VAUGHAN RA, GARCIA-SMITH R, BISOFFI M, CONN CA, TRUJILLO KA. Conjugated linoleic acid or omega 3 fatty acids increase mitochondrial biosynthesis and metabolism in skeletal muscle cells. *Lipids Health Dis* 2012; 11: 142.