LncRNA ZEB2-AS1 promotes the proliferation, migration and invasion of esophageal squamous cell carcinoma cell through miR-574-3p/HMGA2 axis

J.-H. XU¹, R.-Z. CHEN¹, L.-Y. LIU¹, X.-M. LI¹, C.-P. WU¹, Y.-T. ZHOU², J.-D. YAN³, Z.-Y. ZHANG¹

¹Department of Pathology, Tangshan Gongren Hospital, Tangshan, Hebei, China ²Department of Center Laboratory, Tangshan Gongren Hospital, Tangshan, Hebei, China ³Department of Chest Surgery, Tangshan Gongren Hospital, Tangshan, Hebei, China

Jinheng Xu and Rongzhen Chen contributed equally to this work

Abstract. – OBJECTIVE: Esophageal squamous cell carcinoma (ESCC) is a common malignant epithelial tumor in the elderly, and the cause is very complicated. Therefore, the study of the pathogenesis of ESCC is conducive to the effective treatment of ESCC. Many studies indicated that IncRNAs were important regulatory factors in tumor formation and disease development. However, the regulatory network of IncRNA in ESCC has not been fully explored.

MATERIALS AND METHODS: The expression of miR-574-3p, ZEB2-AS1, and HMGA2 was measured using qRT-PCR. The protein expression of PCNA, Cleaved-caspase3, MMP9, and HMGA2 was detected through Western blot. Cell proliferation or apoptosis of transfected cells was calculated via CKK-8 assay or flow cytometry. Transwell assay was applied to detect cell migration and invasion of ESCC cells. Luciferase reporter assay and RNA pull-down were used to determine the relationship among miR-574-3p, ZEB2-AS1, and HMGA2 in ESCC. Moreover, the regulatory network of ZEB2-AS1 has been verified *in vivo* in this study.

RESULTS: We found that ZEB2-AS1 was upregulated in ESCC tissues and cells. The knockdown of ZEB2-AS1 could inhibit cell proliferation, invasion, and migration, as well as promoted cell apoptosis in ESCC. Interestingly, miR-574-3p deficiency or HMGA2 promotion could reverse the effects of si-ZEB2-AS1 on ESCC cell progression. Luciferase reporter assay indicated that miR-574-3p was a target miRNA of ZEB2-AS1 and HMGA2 was a target gene of miR-574-3p in ESCC.

CONCLUSIONS: In this paper, we first verified the novel regulatory mechanism of IncRNA ZEB2-AS1 in ESCC cellular process. LncRNA ZEB2-AS1 promoted the proliferation, migration, and invasion of ESCC by modulating miR-574-3p/HMGA2 axis, indicating that ZEB2-AS1 played essential roles in cell progression in ES-CC and providing a new therapeutic target of ESCC.

Key Words: ESCC, Cell progression, LncRNA, Regulatory mechanism.

Introduction

Esophageal squamous cell carcinoma (ESCC) is a type of esophageal cancer, a malignant tumor with high invasion and poor prognosis¹⁻³. The current treatment methods are very limited, mainly including surgery, radiotherapy, and chemotherapy⁴. Because of the inability to eliminate the lesions, ESCC has a poor prognosis and high recurrence rate. Therefore, studying the pathogenesis of ESCC is an important way to effectively treat ESCC.

LncRNA belongs to a non-coding RNA and is more than 200 nt in length⁵. LncRNA is abnormally expressed in a variety of cancers and is widely involved in important cellular physiological processes, such as cancer cell metabolism and tumor formation⁶⁻⁸. LncRNA, as a ceRNA, regulates mRNA expression by binding to miRNAs and is an important regulator of cell progression in cancer⁹. Multiple lncRNAs have been shown to be involved in the metabolic activities of ESCC cells, such as UCA1, HOTAIR, and GAS5¹⁰⁻¹². Wang et al¹⁰ reported that lncRNA UCA1 suppressed cell progression in ESCC by activating the Wnt signaling pathway. Moreover, lncRNA is also closely related to cancer diagnosis and treatment in ESCC¹³⁻¹⁵. ZEB2-AS1 has been shown to be highly expressed in a variety of cancers including bladder cancer, pancreatic cancer, lung cancer and gastric cancer¹⁶⁻¹⁹. However, the regulatory mechanism of ZEB2-AS1 in ES-CC has not been fully elucidated.

Herein, we found that ZEB2-AS1 was highly expressed in ESCC; further, it was predicted by bioinformatic that miR-574-3p was a potential target miRNA of ZEB2-AS1, and HGMA2 was a candidate target gene of miR-574-3p. The purpose of this study was to elucidate the potential mechanism by which ZEB2-AS1 was involved in the regulation of ESCC cell growth.

Materials and Methods

Patients and Samples

ESCC tissues and adjacent tissues were collected from 30 patients who had been diagnosed with ESCC in Tangshan Gongren Hospital (Tangshan, Hebei, China). This experiment was approved by the Ethics Review Board of Tangshan Gongren Hospital. Written informed consent was obtained from all patients involved in the study.

Cell Culture and Cell Transfection

Normal human esophageal squamous cell (HET-1A) and esophageal squamous cell carcinoma (ESCC) cell lines (EC109, EC9706, KYSE30, and KYSEE150) were purchased from American Type Culture Collection (ATCC; Manassas, VA, USA). All cell lines were cultured in Dulbecco's Modified Eagle's medium (DMEM; Thermo Fisher Scientific Inc., Waltham, MA, USA) with 10% fetal bovine serum (FBS, HyClone, South-Logan, UT, USA), 1% penicillin-streptomycin at 37°C in a humidified atmosphere of 5% CO₂.

The cells were transfected with si-ZEB2-AS1, anti-miR-574-3p, HMGA2 OE (HMGA2), and their negative control using Lipofectamine 2000 reagent (Invitrogen, Carlsbad, CA, USA) according to the manufacturer's instructions.

Cell Proliferation

Cell proliferation was calculated using Cell Counting Kit-8 (CCK-8; Dojindo Laboratories, Kumamoto, Japan) according to the manufacturer's instructions. After transfection for 24 h, cells were seeded into 96-well plates and incubated at 37°C. Then 10 ul CCK-8 solutions were added to each well and incubated 1 h at 37°C. The absorbance was detected using a spectrophotometric microplate reader (Beyotime Institute of Biotechnology, Shanghai, China) at 450 nm.

Cell Apoptosis

Flow cytometry was applied to detect cell apoptosis of transfected cells. Briefly, 2×10^5 transfected cells were seeded in 6-well plates and incubated at room temperature with 5% CO₂. After that, the cells were harvested and stained with annexin V-FITC and PI for 30 min in the dark.

Cell Migration and Invasion

Cell migration and invasion of transfected cells was measured using transwell chambers (Corning Inc., Corning, NY, USA). For cell invasion of transfected cells, the upper chamber with Matrigel (BD Biosciences, Franklin Lakes, NJ, USA) was used. For cell migration of transfected cells, the upper chamber without Matrigel was used. The cells were seeded into the upper chamber, and the lower chamber was filled with medium containing 10% FBS. Then, the plates were incubated at 37°C for 24 h. After incubation, the invasive and migrated cells were stained using 0.1% crystal violet and counted using an optical microscope.

ORT-PCR

The total RNA was isolated from tissues and cells with the TRIzol reagent (Invitrogen, Carlsbad, CA, USA) according to the manufacturer instructions. For miR-574-3p expression, the TaqMan® MicroRNA Real Time-PCR Assay reagents (Applied Biosystems; Foster City, CA, USA) and SYBR[®] Premix Ex Taq[™] reagent (Ta-KaRa, Otsu, Shiga, Japan) were applied. For ZEB2-AS1 and HMGA2 expression, High Capacity cDNA Reverse Transcription Kit (Applied Biosystems) and SYBR® Premix Ex TaqTM reagent (TaKaRa, Otsu, Shiga, Japan) were used to detect. GAPDH and snRNA U6 were used as endogenous controls of mRNA and miRNA, respectively. The relative mRNA and miRNA expression were calculated using $2^{-\Delta\Delta Ct}$ methods. The primers are as follows: lncRNA ZEB2-AS1 forward 5'-ATGAAGAACGCGCGAAGTGT-3' and reverse 5'-CACCCACATTATCACATGC-CCT-3'; HMGA2 forward 5'-AGTCCCTCTA- AAG-CAGCTCAAAAG-3' and reverse 5'-GC-CATTTCC-TAGGTCTGCCTC-3' GAPDH forward 5'-GTCAACGGATTTGGTCTGTATT-3' and reverse 5'-AGTCTTCTGGGTGGCAGT-GAAT-3'. MiR-574-3p reverse 5'-CACGCT-CATGCACACACCCCACA-3' U6 forward 5'-CGCTTCGGCAGCACATATACTA-3' and reverse 5'-CGCTTCACGAATTTGCGTGTCA-3'.

Western Blot

Tissues and transfected cells were detached with trypsin and centrifuged. After washed with PBS, cell lysis buffer was added and incubated to extract protein. The BCA Protein Assay Kit (Beyotime Biotechnology, Shanghai, China) was used to measure protein concentration. The proteins were separated using sodium dodecyl sulphate-polyacrylamide gel electrophoresis (SDS-PAGE) and transferred onto a polyvinylidene difluoride (PVDF) membrane (Millipore, Billerica, MA, USA). After being blocked in non-fat milk, the membrane was incubated with primary antibody (PCNA, Cleaved-caspase3, MMP9, GAP-DH, and HMGA2, 1:2000, ProteinTech, Chicago, IL, USA). Then, the membrane was incubated with horseradish peroxidase (HRP)-conjugated secondary antibody. Finally, the blot was detected and analyzed using an enhanced chemiluminescence (ECL) Western blotting kit (Amersham Biosciences, Little Chalfont, Buckinghamshire, UK).

Luciferase Reporter Assay

The wild type of ZEB2-AS1 or the mutated type of ZEB2-AS1 was inserted into Luciferase reporter plasmid (pGL3, Promega, Madison, WI, USA) to construct the vector of ZEB2-AS1-WT or ZEB2-AS1-MUT. Then, ZEB2-AS1-WT or ZEB2-AS1-MUT was cotransfected with miR-574-3p or miR-NC into KYSE30 and KY-SEE150 cells using Lipofectamine 2000 reagent (Invitrogen, Carlsbad, CA, USA). Similarly, the wild type of HMGA2 or the mutated type of HMGA2 was inserted into Luciferase reporter plasmid (pGL3, Promega, Madison, WI, USA) to construct the vector of HMGA2-WT or HMGA2-MUT. Then, HMGA2-WT or HM-GA2-MUT was cotransfected with miR-574-3p or miR-NC into KYSE30 and KYSEE150 cells using Lipofectamine 2000 reagent (Invitrogen, Carlsbad, CA, USA). The Dual-Luciferase Reporter assay system (Promega, Madison, WI, USA) was applied to detect Renilla and firefly Luciferase activities.

RNA Pull-Down

The cDNA sequence of ZEB2-AS was synthesized and inserted into pBluescript II SK(+) and transfected into KYSE30 and KYSEE150 cells. After Biotin-labeled miR-NC and miR-574-3p were transcribed and purified, a 500 µl RIP buffer with cell lysate was added. Then, 50 µl of washed streptavidin agarose beads were added and the beads were washed with RIP buffer 5 times. The extracted protein was detected by immunoblotting or mass spectrometry.

Xenograft Assay

Cells transfected were injected subcutaneously in the right flank regions of nude mice (4-weekold). Tumor volumes were monitored every 7 days. Tumor volume = width \times length \times (width + length) \times 0.5. Mice were euthanized at day 28 after injection and the tumor weight was measured. Animal studies were approved by the Ethics Review Board of the Tangshan Gongren Hospital.

Statistical Analysis

All data were analyzed and performed using GraphPad Prism 7.0 (GraphPad Software, San Diego, CA, USA). Values were presented as means \pm standard deviation (SD).

Comparison groups were calculated using Student's *t*-test to analyze the difference between groups. Three or more sets of groups were calculated using One-way analysis of variance (ANO-VA) followed by Tukey's test. p<0.05 was considered to be statistically significant.

Results

LncRNA ZEB2-AS1 was Upregulated in ESCC Tissues and Cell Lines

First, we measured the expression of ZEB2-AS1 from 30 ESCC tissues and normal tissues. As shown in Figure 1A, the expression of ZEB2-AS1 was significantly higher in ESCC tissues than that in normal tissues. The expression of ZEB2-AS1 in the ESCC cell lines (EC109, EC9706, KYSE30, and KYSEE150) was significantly higher than in the Normal human esophageal squamous cell (HET-1A) (Figure 1B). Notably, the survival analysis found that the high expression of ZEB2-AS1 of patients with ESCC had a poor prognosis (Figure 1C). These results indicated that ZEB2-AS1 played important roles in ESCC formation and treatment.



Figure 1. LncRNA ZEB2-AS1 was upregulated in ESCC tissues and cell lines. **A**, The expression of ZEB2-AS1 was measured in ESCC tissues and normal tissues. **B**, The expression of ZEB2-AS1 was measured in ESCC cell lines (EC109, EC9706, KYSE30, and KYSEE150) and normal human esophageal squamous cell (HET-1A). **C**, Kaplan-Meier analysis of the survival rates of patients with ESCC in relation to ZEB2-AS1. *p<0.05, **p<0.01.

LncRNA ZEB2-AS1 Promoted Proliferation, Migration and Invasion As Well As Inhibited Apoptosis in ESCC Cell Lines

Next, in order to verify the function of ZEB2-AS1 in ESCC, ZEB2-AS1 was knocked out for transfection into KYSE30 and KYSEE150 cell lines and stably expressed (Figure 2A). The analvsis of CKK-8 assay showed that the knockdown of ZEB2-AS1 significantly inhibited the proliferation of KYSE30 and KYSEE150 cells (Figure 2B and 2C). Conversely, the apoptotic rate of si-ZEB2-AS1 transfection was significantly elevated (Figure 2D and 2E). Meanwhile, reducing the expression of ZEB2-AS1 also greatly decreased the migration and invasion of ESCC cells (Figure 2F-2I). The protein expression of PCNA and MMP9 was significantly decreased while the protein expression of Cleaved-caspase3 was remarkably increased by inhibition of ZEB2-AS1 in KYSE30 and KYSEE150 cell lines (Figure 2J-2L). Therefore, inhibition of ZEB2-AS1 could suppress cell proliferation, migration, and invasion, and promoted cell apoptosis in ESCC cells, implying that ZEB2-AS1 promoted growth in ESCC cell lines.

LncRNA ZEB2-AS1 Directly Interacted with MiR-574-3p

QRT-PCR assay was used to detect the expression level of ZEB2-AS1 in the nucleus and cytoplasm. The results showed that the expression level of ZEB2-AS1 in the cytoplasm was high (Figure 3A). Next, we predicted the presence of the miR-574-3p binding site in ZEB2-AS1 by DIANA tool (Figure 3B). Therefore, we performed Luciferase reporter assay in KYSE30 and KYSEE150 cells, respectively, and showed that the overexpression of miR-574-3p combined with ZEB2-AS1-WT significantly reduced Luciferase activity in cells, rather than ZEB2-AS1-MUT (Figure 3C and 3D). Moreover, the results of RNA pull-down showed that the miR-574-3p probe enriched more ZEB2-AS1 expression compared to the NC probe, indicating that miR-574-3p effectively bound ZEB2-AS1 in ESCC cells (Figure 3E). More than that, the knockdown of ZEB2-AS1 significantly induced the expression of miR-574-3p in KYSE30 and KYSE150 cells (Figure 3F). Lowly expressed miR-574-3p in ES-CC tissues was negatively correlated with ZEB2-AS1 expression (Figure 3G and 3H). All results determined that miR-574-3p was a target miRNA of ZEB2-AS1 in ESCC cells.

LncRNA ZEB2-AS1 Regulated ESCC Progression by Sponging MiR-574-3p

To further clarify the regulatory mechanism between ZEB2-AS1 and miR-574-3p, we co-transfected si-ZEB2-AS1 and anti-miR-574-3p into KYSE30 and KYSEE150 cells, respectively. The result showed that the induction of miR-574-3p by knockdown of ZEB2-AS1 was significantly decreased by downregulating miR-574-3p in KYSE30 and KYSEE150 cells (Figure 4A). Otherwise, cell proliferation, invasion, and migration were significantly decreased by si-ZEB2-AS1 transfection, which was induced by anti-mIR-574-3p transfection in KYSE30 and KYSEE150 cells (Figure 4B, 4E and 4F). As shown in Figure 4C, the promotion effect of si-ZEB2-AS1 on cell apoptosis was inhibited by the suppression of miR-574-3p in KYSE30 and KYSEE150 cells. Knocking out ZEB2-AS1 not only reduced PCNA



Figure 2. LncRNA ZEB2-AS1 promotes proliferation, migration and invasion as well as apoptosis in ESCC cell lines. **A**, The expression of ZEB2-AS1 was detected in KYSE30 and KYSE150 cells transfected with si-NC or si-ZEB2-AS1 *via* qRT-PCR. **B**, and **C**, Cell proliferation was measured in KYSE30 and KYSE150 cells transfected with si-NC or si-ZEB2-AS1 using CCK-8 assay. **D**, and **E**, Cell apoptosis rate was calculated in KYSE30 and KYSE150 cells transfected with si-NC or si-ZEB2-AS1 using CCK-8 assay. **D**, and **E**, Cell apoptosis rate was calculated in KYSE30 and KYSE150 cells transfected with si-NC or si-ZEB2-AS1 using transfected with si-NC or si-ZEB2-AS1 with Western blot. **p*<0.05, ***p*<0.01.

and MMP9 protein expression, but also promoted Cleaved-caspase3 protein expression, which was reversed by underexpression of miR-574-3p in KYSE30 and KYSEE150 cells (Figure 4G and 4H). Thus, lncRNA ZEB2-AS1 regulated ESCC progression by sponging miR-574-3p.

LncRNA ZEB2-AS1 Promoted HMGA2 Expression by Sponging MiR-574-3p in ESCC Cells

The predicted results of starBase showed that there was a miR-574-3p binding site in the 3' non-coding region of HMGA2 (Figure 5A). To



Figure 3. LncRNA ZEB2-AS1 directly interacted with miR-574-3p in ESCC cells. **A**, QRT-PCR assay was used to detect the expression level of ZEB2-AS1 in the nucleus and cytoplasm of KYSE30 and KYSE150 cells. **B**, The predicted sites of miR-574-3p binding to ZEB2-AS1 were detected using DIANA tool. **C**, and **D**, Luciferase activity was measured in KYSE30 (**C**) and KYSE150 (**D**) cells cotransfected with ZEB2-AS1-WT or ZEB2-AS1-MUT and miR-574-3p or miR-NC using luciferase reporter assay. **E**, RNA pull-down was used to determine the enrichment level of ZEB2-AS1 in Bio-miR-NC and Bio-miR-574-3p groups. **F**, The expression of miR-574-3p was detected in KYSE30 and KYSE150 cells transfected with si-NC or si-ZEB2-AS1. **G**, The expression of miR-574-3p was detected in ESCC tissues and normal tissues. **H**, The expression of ZEB2-AS1 was significantly correlated with that of miR-574-3p in ESCC tissues through Pearson's correlated analysis.

further investigate the regulatory mechanism of ZEB2-AS1 in ESCC, the Luciferase reporter assay was used to demonstrate the targeting relationship between miR-574-3p and HMGA2. The results showed that the overexpression of miR-574-3p inhibited the Luciferase activity in the cells transfected with HMGA2 3'-UTR-WT in KYSE30 and KYSEE150 cells (Figure 5B and 5C). The mRNA and protein expression of HMGA2 was sharply inhibited by the promotion of miR-574-3p in KYSE30 and KYSEE150 cells (Figure 5D and 5E). In addition, the knockdown of ZEB2-AS1 reduced the expression of HMGA2 mRNA and protein. When co-transfected with miR-574-3p inhibitor, the mRNA and protein expression of HMGA2 was partially increased

(Figure 5F-5G). Of note, HMGA2, which was highly expressed in ESCC tissues, was negatively correlated with miR-574-3p expression but positively correlated with ZEB2-AS1 expression (Figure 5J-5L). All data determined that ZEB2-AS1 promoted HMGA2 expression by sponging miR-574-3p in ESCC cells.

Restoration of HMGA2 Reversed the Effects of LncRNA ZEB2-AS1 Knockdown in ESCC Cells

To investigate the effect of the ZEB2-AS1/ miR-574-3p/HMGA2 regulatory network on ES-CC cell progression, si-ZEB2-AS1 and HMGA2 overexpression (HMGA2) were co-transfected into KYSE30 and KYSEE150 cells. The results



Figure 4. LncRNA ZEB2-AS1 regulated ESCC progression by sponging miR-574-3p. **A**, The expression of miR-574-3p was detected in KYSE30 and KYSE150 cells transfected with si-NC, si-ZEB2-AS1 or si-ZEB2-AS1 or si-ZEB2-AS1+ anti-miR-574-3p wia qRT-PCR. **B**, and **C**, Cell proliferation was measured in KYSE30 and KYSE150 cells transfected with si-NC, si-ZEB2-AS1 or si-ZEB2-AS1+ anti-miR-574-3p using CCK-8 assay. **D**, Cell apoptosis rate was calculated in KYSE30 and KYSE150 cells transfected with si-NC, si-ZEB2-AS1+ anti-miR-574-3p using CCK-8 assay. **D**, Cell apoptosis rate was calculated in KYSE30 and KYSE150 cells transfected with si-NC, si-ZEB2-AS1+ anti-miR-574-3p using transwell assay. **G**, and **H**, The protein level of PCNA, Cleaved-caspase3 and MMP9 was measured in KYSE30 and KYSE150 cells transfected with si-NC, si-ZEB2-AS1+ anti-miR-574-3p using transwell assay. **G**, and **H**, The protein level of PCNA, Cleaved-caspase3 and MMP9 was measured in KYSE30 and KYSE150 cells transfected with si-NC, si-ZEB2-AS1+ anti-miR-574-3p using transwell assay. **G**, and **H**, The protein level of PCNA, Cleaved-caspase3 and MMP9 was measured in KYSE30 and KYSE150 cells transfected with si-NC, si-ZEB2-AS1+ anti-miR-574-3p using transwell assay. **G**, and **H**, The protein level of PCNA, Cleaved-caspase3 and MMP9 was measured in KYSE30 and KYSE150 cells transfected with si-NC, si-ZEB2-AS1+ anti-miR-574-3p using transwell assay. **G**, and **H**, The protein level of PCNA, Cleaved-caspase3 and MMP9 was measured in KYSE30 and KYSE150 cells transfected with si-NC, si-ZEB2-AS1+ anti-miR-574-3p using transwell assay. **G**, and **H**, The protein level of PCNA, Cleaved-caspase3 and MMP9 was measured in KYSE30 and KYSE150 cells transfected with si-NC, si-ZEB2-AS1+ anti-miR-574-3p using transwell assay. **G**, and **H**, The protein level of PCNA, Cleaved-caspase3 and MMP9 was measured in KYSE30 and KYSE150 cells transfected with si-NC, si-ZEB2-AS1+ anti-miR-574-3p using transfected with si-NC, si-ZEB2-AS1+ anti-miR-574-3p using transfected with si-

5397



Figure 5. LncRNA ZEB2-AS1 promoted HMGA2 expression by sponging miR-574-3p in ESCC cells. **A**, The predicted sites of miR-574-3p binding to 3'UTR of HMGA2 were detected using starBase. **B**, and **C**, Luciferase activity was measured in KYSE30 (**B**) and KYSE150 (**C**) cells cotransfected with HMGA2 3'UTR-WT or HMGA2 3'UTR-MUT and miR-574-3p or miR-NC using luciferase reporter assay. **D**, and **E**, The mRNA (**D**) and protein (**E**) expression of HGM was detected in KYSE30 and KYSE150 cells transfected with si-NC or miR-574-3p. **F-H**, The mRNA (**F**) and protein (**G** and **H**) expression of HMGA2 was detected in KYSE30 and KYSE150 cells transfected with si-NC, si-ZEB2-AS1 or si-ZEB2-AS1+ anti-miR-574-3p *via* qRT-PCR and Western blot. **J**, The expression of HMGA2 was detected in ESCC tissues and normal tissues. **K**, The expression of HMGA2 was significantly negative correlated with that of miR-574-3p in ESCC tissues through Pearson's correlated analysis. **L**, The expression of HMGA2 was significantly positive correlated with that of ZEB2-AS1 in ESCC tissues through Pearson's correlated analysis.

indicated that overexpression of HMGA2 could weaken the suppression effect of si-ZEB2-AS1 on HMGA2 mRNA and protein expression in KYSE30 and KYSEE150 cells (Figure 6A and 6B). Knockdown of ZEB2-AS1 significantly inhibited cell proliferation, invasion and migration, and promoted apoptosis, which was rescued by overexpression of HMGA2 (Figure 6C-6G). Furthermore, PCNA and MMP9 protein expression were inhibited and Cleaved-caspase3 protein expression was induced by downregulation of ZEB2-AS1, which was reversed by the promotion of HMGA2 in KYSE30 and KYSEE150 cells (Figure 6H and 6I). Thus, the restoration of HMGA2 reversed the effects of lncRNA ZEB2-AS1 knockdown in ESCC cells.

Knockdown of ZEB2-AS1 Inhibited Tumor Growth In Vivo

To further investigate the effects of ZEB2-AS1 on the formation of ESCC, we performed experimental validation in mice. We injected KYSE30 cells transfected with sh-con and sh-ZEB2-AS1 subcutaneously into the right side of the nude mice (N=5). Tumor volume was measured every 7 days, and tumors were removed after transfection for 35 days. The results showed that ESCC tumor volume and weight were significantly inhibited after KEB2-AS1 was knocked out (Figure 7A and 7B). In ESCC tissues, Sh-ZEB2-AS1 significantly increased the expression of miR-574-3p and decreased the expression of ZEB2-AS1 (Figure 7C and 7D). In addition, the protein expressions of HMGA2, PCNA, and MMP9 were significantly lower in Sh-ZEB2-AS1 group than in the sh-con group, while the protein expression of cleavedcaspase3 was reversed (Figure 7E). Therefore, the knockdown of ZEB2-AS1 inhibited tumor growth in vivo.

Discussion

Due to the complex etiology of esophageal cancer, the recurrence rate is high, and the prognosis is poor²⁰. In order to further clarify the pathogenesis of ESCC and improve the therapeutic effect, lncRNAs, which act as important regulators, needed to further investigation. Increasing reports²¹⁻²⁵ showed that lncRNAs were involved in cellular processes, such as cell proliferation and apoptosis in a variety of cancers. Moreover, lncRNAs were also associated with

immune responses and drug sensitivity^{26,27}. LncRNA PCAT1 was related to poor prognosis and contributed cell proliferation, invasion, migration, and EMT in osteosarcoma²⁸. LncRNA has also been proposed to play roles in ESCC cellular process and metastasis²⁹⁻³¹. LncRNA AFAP1-AS1 was upregulated in ESCC cells and promoted cell growth, as well as reduced cell apoptosis in ESCC³². In this study, consistent with previous research³³, we found that lncRNA ZEB2-AS1 expression was increased in ESCC tissues and cells, indicating that ZEB2-AS1 was associated with ESCC formation. Moreover, Lan et al³⁴ reported that ZEB2-AS1 participated in tumor growth and metastasis of hepatocellular carcinoma. Meanwhile, the similar function of ZEB2-AS1 has been verified in colon cancer, pancreatic cancer, bladder cancer, and gastric cancer^{16,17,19,35}. However, the function of ZEB2-AS1 in ESCC has not been fully clarified. Our study is the first to investigate that the knockdown of ZEB2-AS1 could inhibit ESCC cell proliferation, invasion, and migration while promoting apoptosis.

Furthermore, we also explored the regulatory mechanism of ZEB2-AS1 in ESCC cell proliferation. Gao et al¹⁷ reported that ZEB2-AS1 affected pancreatic cancer formation and cell invasion by modulating miR-204/HMGB1 axis. In this study, Luciferase reporter assay determined that miR-574-3p was a target miRNA of ZEB2-AS1 and was negatively regulated in ZEB2-AS1 expression. MiR-574-3p is also abnormally expressed in many types of cancer³⁶⁻³⁸. Su et al³⁹ indicated that miR-574-3p had important biological significance in gastric cancer and was closely related to the early diagnosis of gastric cancer. Of note, in our study, we found that miR-574-3p was downregulated in ESCC tissues and reduction of miR-574-3p could reverse the suppressive effects of si-ZEB2-AS1 on ESCC cell growth, indicating that ZEB2-AS1 play roles in ESCC progression by targeting miR-574-3p.

In addition, we further explored the regulatory mechanism of ZEB2-AS1 and found that HM-GA2 was a target mRNA of miR-574-3p. Functional experiments showed that HMGA2 overexpression could weaken the repressive effects of knockdown of ZEB2-AS1 on ESCC cell progression. Many studies⁴⁰⁻⁴² determined that HMGA2 was closely related to EMT in human pancreatic cancer cells and gastric cancer. Sun et al⁴³ reported that HMGA2/TET1/HOXA9 axis participated in the regulation of breast cancer growth and



Figure 6. Restoration of HMGA2 reversed the effects of lncRNA ZEB2-AS1 knockdown in ESCC cells. **A**, and **B**, The mRNA (**A**) and protein (**B**) expression of HMGA2 was detected in KYSE30 and KYSE150 cells transfected with si-NC, si-ZEB2-AS1 or si-ZEB2-AS1+ HMGA2 via qRT-PCR. **C**, and **D**, Cell proliferation was measured in KYSE30 (**C**) and KYSE150 (**D**) cells transfected with si-NC, si-ZEB2-AS1 or si-ZEB2-AS1+ HMGA2 using CCK-8 assay. **E**, Cell apoptosis rate was calculated in KYSE30 and KYSE150 cells transfected with si-NC, si-ZEB2-AS1+ HMGA2 via flow cytometry. **F**, and **G**, Cell migration (**F**) and invasion (**G**) was detected in KYSE30 and KYSE150 cells transfected with si-NC, si-ZEB2-AS1+HMGA2 using transwell assay. **H**, and **I**, The protein level of PCNA, Cleaved-caspase3, and MMP9 was measured in KYSE30 (**H**) and KYSE150 (**I**) cells transfected with si-NC, si-ZEB2-AS1 or si-ZEB2-AS1 or si-ZEB2-AS1+HMGA2 using transwell assay. **H**, and **I**, The protein level of PCNA, Cleaved-caspase3, and MMP9 was measured in KYSE30 (**H**) and KYSE150 (**I**) cells transfected with si-NC, si-ZEB2-AS1 or si-ZEB2-AS1+HMGA2 using transwell assay. **H**, and **I**, The protein level of PCNA, Cleaved-caspase3, and MMP9 was measured in KYSE30 (**H**) and KYSE150 (**I**) cells transfected with si-NC, si-ZEB2-AS1 or si

5400



Figure 7. Knockdown of ZEB2-AS1 inhibits tumor growth *in vivo*. **A**, and **B**, Tumor volume (**A**) and weight (**B**) were calculated in sh-con and sh-ZEB2-AS1 groups in mice. **C**, and **D**, The expression of ZEB2-AS1 (**C**) and miR-574-3p (**D**) was measured in sh-con and sh-ZEB2-AS1 groups in mice *via* qRT-PCR. **E**, The protein level of PCNA, Cleaved-caspase3, MMP9, and HMGA2 was measured in sh-con and sh-ZEB2-AS1 groups in mice *via* qRT-PCR. **E**, The protein level of PCNA, Cleaved-caspase3, MMP9, and HMGA2 was measured in sh-con and sh-ZEB2-AS1 groups in mice *via* Western blot. *p < 0.05, **p < 0.01.

metastasis. In ESCC, HMGA2 has been reported to be a target gene of miR-490-3p, miR-125b-5p, and miR-195 and regulated cancer formation, cell proliferation, metastasis, and apoptosis. In our study, we first verified that ZEB2-AS1/miR-574-3p/HMGA2 axis played an essential role in ESCC cell proliferation, invasion, migration, and apoptosis. Taken together, our findings revealed the novel regulatory mechanism of lncRNA in ESCC progression.

Conclusions

In this paper, we first verified the novel regulatory mechanism of lncRNA ZEB2-AS1 in ESCC cellular process. LncRNA ZEB2-AS1 promoted the proliferation, migration, and invasion of ES-CC by modulating miR-574-3p/HMGA2 axis, indicating ZEB2-AS1 played essential roles in cell progression in ESCC and providing a new therapeutic target of ESCC.

Conflict of Interest

The Authors declare that they have no conflict of interests.

References

- 1) GATENBY PIERS AC, PRESTON SHAUN R. Oesophageal cancer. Surgery 2014; 32: 588-593.
- GUO Y, CHEN Z, ZHANG L, ZHOU F, SHI S, FENG X, LI B, MENG X, MA X, LUO M, SHAO K, LI N, QIU B, MITCH-ELSON K, CHENG J, HE J. Distinctive microRNA profiles relating to patient survival in esophageal squamous cell carcinoma. Cancer Res 2008; 68: 26-33.
- Song C, Xing D, Tan W, WEI Q, Lin D. Methylenetetrahydrofolate reductase polymorphisms increase risk of esophageal squamous cell carcinoma in a Chinese population. Cancer Res 2001; 61: 3272-3275.
- NAKAJIMA M, KATO H. Treatment options for esophageal squamous cell carcinoma. Expert Opin Pharmacother 2013; 14: 1345-1354.
- 5) MATTICK JS, MAKUNIN IV. Non-coding RNA. Hum Mol Genet 2006; 15 Spec No 1: R17-R29.
- XIA T, LIAO Q, JIANG X, SHAO Y, XIAO B, XI Y, GUO J. Long noncoding RNA associated-competing endogenous RNAs in gastric cancer. Sci Rep 2015; 4: 6088.
- IM JH, MUSCHEL RJ. New evidence of IncRNA role in tumor progression and metastasis. Hepatobiliary Surg Nutr 2012; 1: 55-56.
- ZHANG YL, WAN XC, FU FK, LI Y. Research progress of non-coding RNA (ncRNA) mechanism in prostate cancer. Scientia Sinica 2016; 46: 385-391.

- 9) YANG S, NING O, ZHANG G, SUN H, WANG Z, LI Y. Construction of differential mRNA-IncRNA crosstalk networks based on ceRNA hypothesis uncover key roles of IncRNAs implicated in esophageal squamous cell carcinoma. Oncotarget 2016; 7: 85728-85740.
- 10) WANG X, GAO Z, LIAO J, SHANG M, LI X, YIN L, PU Y, LIU R. LnCRNA UCA1 inhibits esophageal squamous-cell carcinoma growth by regulating the Wnt signaling pathway. J Toxicol Environ Health A 2016; 79: 407-418.
- SONG W, ZOU SB. Prognostic role of IncRNA HO-TAIR in esophageal squamous cell carcinoma. Clin Chim Acta 2016; 463: 169-173.
- 12) WANG K, LI J, XIONG G, HE G, GUAN X, YANG K, BAI Y. Negative regulation of IncRNA GAS5 by miR-196a inhibits esophageal squamous cell carcinoma growth. Biochem Biophys Res Commun 2018; 495: 1151-1157.
- 13) TONG YS, ZHOU XL, WANG XW, WU QQ, YANG TX, LV J, YANG JS, ZHU B, CAO XF. Association of decreased expression of long non-coding RNA LOC285194 with chemoradiotherapy resistance and poor prognosis in esophageal squamous cell carcinoma. J Transl Med 2014; 12: 233.
- 14) WANG W, WEI C, LI P, WANG L, LI W, CHEN K, ZHANG J, ZHANG W, JIANG G. Integrative analysis of mRNA and IncRNA profiles identified pathogenetic IncRNAs in esophageal squamous cell carcinoma. Gene 2018; 661: 169-175.
- 15) GE XS, MA HJ, ZHENG XH, RUAN HL, LIAO XY, XUE WQ, CHEN YB, ZHANG Y, JIA WH. HOTAIR, a prognostic factor in esophageal squamous cell carcinoma, inhibits WIF-1 expression and activates Wnt pathway. Cancer Sci 2013; 104: 1675-1682.
- 16) WU X, YAN T, WANG Z, WU X, CAO G, ZHANG C. LncRNA ZEB2-AS1 promotes bladder cancer cell proliferation and inhibits apoptosis by regulating miR-27b. Biomed Pharmacother 2017; 96: 299-304.
- 17) GAO H, GONG N, MA Z, MIAO X, CHEN J, CAO Y, ZHANG G. LncRNA ZEB2-AS1 promotes pancreatic cancer cell growth and invasion through regulating the miR-204/HMGB1 axis. Int J Biol Macromol 2018; 116: 545-551.
- Guo Y, Hu Y, Hu M, HE J, Li B. Long non-coding RNA ZEB2-AS1 promotes proliferation and inhibits apoptosis in human lung cancer cells. Oncol Lett 2018; 15: 5220-5226.
- 19) WANG F, ZHU W, YANG R, XIE W, WANG D. LncRNA ZEB2-AS1 contributes to the tumorigenesis of gastric cancer via activating the Wnt/β-catenin pathway. Mol Cell Biochem 2019; 456: 73-83.
- 20) Du XX, Yu R, Wang ZF, Du DC, Liu QY, Wang RM, Kang SR, Yang H. Outcomes and prognostic factors for patients with cervical esophageal cancer undergoing definitive radiotherapy or chemoradiotherapy. Bosn J Basic Med Sci 2019; 19: 186-194.
- 21) ZENG B, YE H, CHEN J, CHENG D, CAI C, CHEN G, CHEN X, XIN H, TANG C, ZENG J. LncRNA TUG1 sponges miR-145 to promote cancer progression and reg-

ulate glutamine metabolism via Sirt3/GDH axis. Oncotarget 2017; 8: 113650-113661.

- 22) Cui Z, AN X, Li J, Liu Q, Liu W. LncRNA MIR22HG negatively regulates miR-141-3p to enhance DAPK1 expression and inhibits endometrial carcinoma cells proliferation. Biomed Pharmacother 2018; 104: 223-228.
- 23) XIAO C, WU CH, HU HZ. LncRNA UCA1 promotes epithelial-mesenchymal transition (EMT) of breast cancer cells via enhancing Wnt/beta-catenin signaling pathway. Eur Rev Med Pharmacol Sci 2016; 20: 2819-2824.
- 24) ZHANG J, LI XY, HU P, DING YS. LncRNA NORAD contributes to colorectal cancer progression by inhibition of miR-202-5p. Oncol Res 2018. doi: 10.3727/0 96504018X15190844870055. [Epub ahead of print].
- 25) ZHOU X, YE F, YIN C, ZHUANG Y, YUE G, ZHANG G. The interaction between miR-141 and IncRNA-H19 in regulating cell proliferation and migration in gastric cancer. Cell Physiol Biochem 2015; 36: 1440-1452.
- 26) ZHANG P, CAO L, ZHOU R, YANG X, WU M. The IncRNA Neat1 promotes activation of inflammasomes in macrophages. Nat Commun 2019; 10: 1495.
- 27) O'CONNELL RM, KAHN D, GIBSON WS, ROUND JL, SCHOLZ RL, CHAUDHURI AA, KAHN ME, RAO DS, BALTI-MORE D. MicroRNA-155 promotes autoimmune inflammation by enhancing inflammatory T cell development. Immunity 2010; 33: 607-619.
- 28) ZHANG X, ZHANG Y, MAO Y, MA X. The IncRNA PCAT1 is correlated with poor prognosis and promotes cell proliferation, invasion, migration and EMT in osteosarcoma. Onco Targets Ther 2018; 11: 629-638.
- 29) LIANG Y, CHEN X, WU Y, LI J, ZHANG S, WANG K, GUAN X, YANG K, BAI Y. LncRNA CASC9 promotes esophageal squamous cell carcinoma metastasis through upregulating LAMC2 expression by interacting with the CREB-binding protein. Cell Death Differ 2018; 25: 1980-1995.
- LIU H, ZHEN Q, FAN Y. LncRNA GHET1 promotes esophageal squamous cell carcinoma cells proliferation and invasion via induction of EMT. Int J Biol Markers 2017; 32: e403-e408.
- 31) YAO J, SHEN X, LI H, XU J, SHAO S, HUANG JX, LIN M. LncRNA-ECM is overexpressed in esophageal squamous cell carcinoma and promotes tumor metastasis. Oncol Lett 2018; 16: 3935-3942.
- 32) Luo HL, HUANG MD, GUO JN, FAN RH, XIA XT, HE JD, CHEN XF. AFAP1-AS1 is upregulated and promotes esophageal squamous cell carcinoma cell proliferation and inhibits cell apoptosis. Cancer Med 2016; 5: 2879-2885.
- 33) WANG YL, BAI Y, YAO WJ, GUO L, WANG ZM. Expression of long non-coding RNA ZEB1-AS1 in esophageal squamous cell carcinoma and its correlation with tumor progression and patient survival. Int J Clin Exp Pathol 2015; 8: 11871-11876.
- 34) LAN T, CHANG L, WU L, YUAN Y. Downregulation of ZEB2-AS1 decreased tumor growth and metastasis in hepatocellular carcinoma. Mol Med Rep 2016; 14: 4606-4612.

- 35) BATLINER J, ABBA M, HAUSER C, ALLGAYER H. Abstract 4186: miR-218 suppresses invasion and migration via inhibition of ZEB2 and N-Cadherin in colon cancer. Cancer Res 2013; 73: 4186.
- 36) Xu H, Liu X, ZHOU J, CHEN X, ZHAO J. MiR-574-3p acts as a tumor promoter in osteosarcoma by targeting SMAD4 signaling pathway. Oncol Lett 2016; 12: 5247-5253.
- 37) UJIHIRA T, IKEDA K, SUZUKI T, YAMAGA R, SATO W, HORIE-INOUE K, SHIGEKAWA T, OSAKI A, SAEKI T, OKA-MOTO K, TAKEDA S, INOUE S. MicroRNA-574-3p, identified by microRNA library-based functional screening, modulates tamoxifen response in breast cancer. Sci Rep 2015; 5: 7641.
- 38) TATARANO S, CHIYOMARU T, KAWAKAMI K, ENOKI-DA H, YOSHINO H, HIDAKA H, NOHATA N, YAMASAKI T, GOTANDA T, TACHIWADA T, SEKI N, NAKAGAWA M. Novel oncogenic function of mesoderm development candidate 1 and its regulation by miR-574-3p in bladder cancer cell lines. Int J Oncol 2012; 40: 951-959.
- 39) SU Y, NI Z, WANG G, CUI J, WEI C, WANG J, YANG Q, XU Y, LI F. Aberrant expression of microRNAs

in gastric cancer and biological significance of miR-574-3p. Int Immunopharmacol 2012; 13: 468-475.

- 40) Ma C, Nong K, Zhu H, Wang W, Huang X, Yuan Z, Ai K. H19 promotes pancreatic cancer metastasis by derepressing let-7's suppression on its target HM-GA2-mediated EMT. Tumour Biol 2014; 35: 9163-9169.
- 41) WATANABE S, UEDA Y, AKABOSHI S, HINO Y, SEKITA Y, NAKAO M. HMGA2 maintains oncogenic RAS-induced epithelial-mesenchymal transition in human pancreatic cancer cells. Am J Pathol 2009; 174: 854-868.
- 42) ZHA L, ZHANG J, TANG W, ZHANG N, HE M, GUO Y, WANG Z. HMGA2 elicits EMT by activating the Wnt/β-catenin pathway in gastric cancer. Dig Dis Sci 2013; 58: 724-733.
- 43) SUN M, SONG CX, HUANG H, FRANKENBERGER CA, SAN-KARASHARMA D, GOMES S, CHEN P, CHEN J, CHADA KK, HE C, ROSNER MR. HMGA2/TET1/HOXA9 signaling pathway regulates breast cancer growth and metastasis. Proc Natl Acad Sci U S A 2013; 110: 9920-9925.