

Causal link between gut microbiota and osteoporosis analyzed *via* Mendelian randomization

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Abstract. – OBJECTIVE: Osteoporosis (OP) is closely associated with gut microbiota (GM), yet the nature of their causal relationship remains elusive. Therefore, this study aims to reverse causality between GM and OP by using population cohorts and two-sample MR (TSMR) analysis.

MATERIALS AND METHODS: In this study, we conducted an extensive genome-wide association study (GWAS) using publicly accessible summary statistics data for GM and OP. Employing rigorous criteria ($p < 1 \times 10^{-5}$), we identified independent genetic loci that exhibited significant associations with GM relative abundances as instrumental variables (IVs). A causal evaluation was primarily carried out using the inverse variance-weighted (IVW) method, supplemented by additional analyses such as MR-Egger, weighted median, simple mode, and weighted mode.

RESULTS: We unveiled that increased abundances of the family *Pasteurellaceae*, order Pasteurellales, and genus *Ruminococcaceae* UCG004 were linked to an increased risk of OP. Conversely, the family Oxalobacteraceae, unknown family id.1000006161, genus *Lachnospiraceae* NK4A136 group, unknown genus id.1000006162, and order *NB1n* were associated with a reduced risk of OP. To ensure the reliability of our findings, we conducted quality assessments through Cochrane's Q test and a leave-one-out analysis. Furthermore, the stability and consistency of the results were confirmed by the MR-Egger intercept test, Mendelian randomization pleiotropy residual sum and outlier (MR-PRESSO) global test, and sensitivity analysis ($p > 0.05$). Our study reveals the causal

relationships between 211 GM taxa and OP, pinpointing specific GM taxa associated with the risk of OP. This research sheds light on the genetic mechanisms that underlie GM-mediated OP and opens up promising avenues for identifying valuable biomarkers and potential therapeutic targets in future OP research.

CONCLUSIONS: This study establishes a substantial GM-OP link with specific taxa being identified, offering biomarkers for early detection, tailored interventions, and improved patient education. These findings enhance OP diagnosis, prevention, and treatment, promising more effective, individualized care and inspiring future research.

Key Words:

Mendelian randomization, Gut microbiota, Osteoporosis, Causal relationship, Genetic analysis.

Introduction

Osteoporosis (OP) is a systemic bone metabolism disease characterized by decreased bone density, disrupted bone microstructure, and increased risk of fragility fractures. It has become one of the major public health issues worldwide¹. OP is closely related to the aging of the population and its prevalence is increasing year by year, with females having a four-fold higher incidence than males. According to the World Health Organization (WHO), more than 200 million people worldwide suffer from OP, which causes 8.9

million fractures annually². In the United States, over 10 million people undergo OP³. In Europe, OP is one of the most common bone diseases among the elderly population and causes over 3 million fractures annually⁴. Fragility fractures caused by OP have brought a significant and increasingly growing economic burden to health-care systems worldwide⁵. The etiology of OP is not yet fully understood, but it may be related to genetic factors, changes in hormone levels, nutrient deficiencies, gut dysbiosis, diseases and drug effects, and unhealthy lifestyles^{6,7}.

The human gastrointestinal tract is colonized by more than 1,013-14 bacteria, and the gut microbiota (GM) contains approximately 150 times more genes than the human genome. The gut microbial ecosystem is the richest one in the human body. The interaction between the microbial population and the host plays a crucial role in regulating the host's metabolism and immune system and is a key factor that influences health⁸. Gut dysbiosis has been associated with various diseases, including diabetes, obesity, Alzheimer's Disease, rheumatoid arthritis, and multiple sclerosis⁹⁻¹¹. Depletion of GM might simultaneously affect the gut and the central nervous system through 5-hydroxytryptamine (5-HT), indicating the existence of a gut-brain-bone-axis¹². In recent years, the study of the relationship between the GM and OP has become a hot topic in scientific research. Investigating the association between GM and bone health, including the gut-bone axis and the role of gut-derived hormones, holds promise for uncovering novel strategies for preventing and treating osteoporosis, particularly in the elderly. This is a field with the potential to improve population health, especially as the aging population grows¹³. GM is a regulator of bone mass, and some scholars¹⁴ have suggested that the inhibitory effect of GM on bone mass is mediated by its impact on the immune system.

Studies¹⁵ have shown that GM regulates bone metabolism through various pathways. GM can affect the production of osteocalcin and cytokines such as interleukins by B cells and Treg cells, and regulate the differentiation of osteoclasts and bone resorption. In addition, GM can also regulate bone metabolism through nucleotide-binding oligomerization domains (NOD), and Wnt/ β -catenin signaling pathways^{16,17}. Specific inactivation of NOD1 or NOD2 in germ-free (GF) mice did not significantly increase cortical bone mass, while tumor necrosis factor and receptor activator of nuclear factor-kappa B ligand (RANKL)

were lower than normal levels, indicating that the regulation of GM on bone mass depends on NOD1 and NOD2 signaling¹⁸. Leptin, low-density lipoprotein receptor-related protein 5 (LRP5) gene, and transcription factor FOXO1 have also been found to be associated with the regulation of 5-HT by the GM in the anabolic therapy of OP¹⁹.

These studies^{18,19} provide some references for relevant human clinical trials. Microbial fermentation of dietary fiber can produce short-chain fatty acids (SCFAs), which can promote the production of serum insulin-like growth factor-1 (IGF-1), thereby regulating bone mass and metabolism²⁰. Recent literature have found that gut dysbiosis may alter Th17/Treg immune responses and promote osteogenesis, changing the migration of mononuclear cells and lymphocytes in tissues. Guo et al²¹ found that *Lactobacillus rhamnosus* GG (LGG) treatment had a protective effect in an ovariectomized (OVX) rat model. Therefore, we speculate that gut dysbiosis may be one of the reasons leading to the occurrence and aggravation of OP. There may be a causation between them. After all, different study designs may lead to different conclusions, and the human gastrointestinal tract is influenced by various factors, including diet and rest. Therefore, it is necessary to establish research on the relationship between GM and OP.

Mendelian randomization (MR) is a statistical method used in epidemiological inference to uncover causal relationships between exposure and outcomes. It utilizes genetic variation associated with the exposure as a proxy to evaluate the association between the exposure factor and the outcome²². Based on the rich harvest of large-scale genome-wide association studies (GWASs) at the genetic and disease levels, MR analysis has been widely applied in various disease studies²³⁻²⁵. This study aims to use population cohorts and two-sample MR analysis to reverse causality between GM and OP. The mechanism of using MR to analyze the causal relationship remains unexplored. It is expected that these approaches will further promote the research and development of GM and OP and provide new insights for microbiology and related fields.

Materials and Methods

Research Methodology

This study investigated the causal link between 211 GM taxa and OP using a TSMR anal-

ysis. We adhered to the MR-STROBE guidelines (**Supplementary Table 1**)²⁶ and relied on existing publicly available summary data, eliminating the need for extra ethical approval. The MR analysis must follow three key assumptions: strong association between instrumental variables (IVs) and GM, lack of IVs' association with confounding factors, and IVs' impact solely through the GM pathway referred to in our previous research methods⁹. Detailed methodology is provided in Figure 1.

Genome-Wide Association Study (GWAS) Data Sources

We collected data for this OP study from the UK Biobank (<https://gwas.mrcieu.ac.uk/>, id: ukb-a-87, accessed on 2 May 2023), a well-established Biobank in the UK with nearly 500,000 participants. This extensive database includes genetic information, as well as comprehensive data on demographics, socioeconomic factors, lifestyle, and health for its participants²⁷. We chose a more recent GWAS with a large sample size

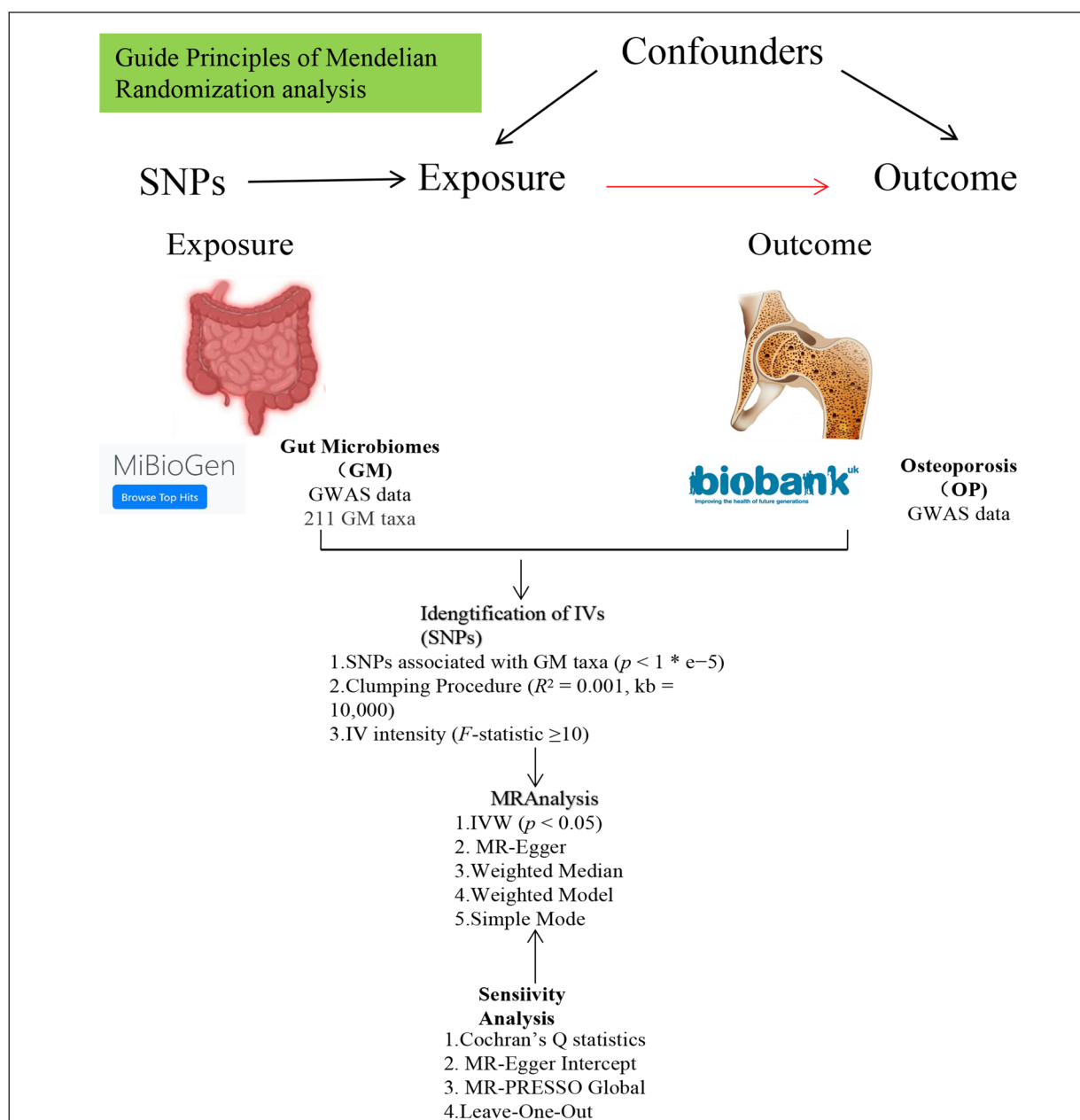


Figure 1. Study flow.

single nucleotide polymorphism (SNPs), and the phenotype “osteoporosis, strict definition” within the UK Biobank on OP cases (5,266) and controls (331,893) identified using the ICD-10 code M81.0 (ID: ukb-a-87).

For the analysis of 16S rRNA gene sequencing profiles and genetic typing data with GM, we utilized data from the latest GWAS summary data provided by the MiBioGen consortium (www.mibiogen.org, accessed on 2 May 2023). This dataset includes 18,340 individuals from 24 European cohorts, spanning 211 taxa and 122,110 variant sites²⁸. Our GM study examined 16S rRNA gene regions (V4, V3-V4, V1-V2), using microbiota quantitative trait loci (mbQTL) mapping to spot genetic variations linked to microbial taxa abundance. In summary, our research drew upon the UK Biobank for OP genetic data and the MiBioGen consortium for GM data, allowing us to perform a comprehensive analysis of both aspects.

Instrumental Variables Identification

We aimed to evaluate the association between IVs and GM with a significance threshold set at $p < 1 \times 10^{-5}$. The methodology employed can be summarized as follows based on previous studies²⁹:

- Relevant Single nucleotide polymorphisms (SNPs) were extracted using the TSMR package in R software (version 4.2.0, The R Foundation for Statistical Computing, Nashville, TN, USA).
- SNPs were subjected to clustering to mitigate the impact of linkage disequilibrium (LD), with LD status assessed using cumulative explained variance (r^2) and kb values. SNPs meeting the criteria of $r^2 < 0.001$ within a 10,000 kb window were selected.
- Echo SNPs were excluded from the analysis.
- The remaining SNPs were derived from a combination of p -value filtering removal of chain imbalances.
- The PhenoScanner database was utilized to retrieve significant results from GWAS data ($p < 1 \times 10^{-5}$), while simultaneously eliminating potential confounding factors (<http://www.phenoscanter.medschl.cam.ac.uk/phenoscanter>, on 2 May 2023)³⁰.
- The IVs strength was estimated for each GM taxon using the F -statistic, calculated as:

$$F = \frac{R^2 \times (N-1-k)}{(1-R^2) \times k}$$

$$R^2 = 2 \times AF \times (1-MAF) \times \left(\frac{\beta}{sd} \right)$$

Here MAF represents the minimum allele frequency, β is derived from the exposed GWAS, sd denotes its variance ($sd = se \times \sqrt{N}$), N signifies the total sample size in the exposed GWAS, and k indicates the number of IVs³¹.

Statistical Analysis

We used the TSMR method to examine the potential causal link between GM and OP. We primarily employed the random-effects inverse variance weighted (IVW) analysis, along with supplementary methods such as the weighted median, MR-Egger, Robust IVW, and Robust MR-Egger, to ensure robust results and account for outliers. To assess horizontal pleiotropy, we conducted MR-Egger intercept tests and Mendelian randomization pleiotropy residual sum and outlier (MR-PRESSO) tests. Heterogeneity was evaluated using Cochran's Q and I^2 values, with IVs having p -values lower than 0.05 considered heterogeneous.

Our rationale for these statistical tests is as follows:

- IVW provides an overall effect estimate by weighted averaging of all IVs, assuming they meet MR assumptions, and it is more powerful without horizontal pleiotropy³².
- The weighted median method is robust and reduces outlier influence³³.
- MR-Egger handles IVs correlation but demands careful IV selection. It detects and corrects horizontal pleiotropy³⁴.
- MR-PRESSO methods identify and correct outliers and anomalies, ensuring robust results³⁵.
- We conducted leave-one-out MR analyses for sensitivity testing³⁶.

We utilized R Studio, with various R packages for statistical analysis, setting a significance threshold of $p < 0.001$. These analyses were conducted in January 2023. Funnel and forest plots were generated to detect pleiotropy.

Results

Screening Selection of IVs and Initial MR Analysis Outcomes

A total of 2,551 SNPs were identified as IVs for 211 GM taxa. These IVs were categorized

into five different levels based on their taxonomic classification (Phylum, Class, Order, Family, and Genus). Eight microbial taxa were isolated based on the IVW method, with both a p -value of < 0.05 and an F -statistics > 10 for these positive IVS, indicating no evidence of weak instrument bias. In the heterogeneity test, the p -values for Cochran's Q statistic were all higher than 0.05, indicating the absence of heterogeneity among the SNPs (**Supplementary Table II** presents the confounder factors.).

Subsequently, we employed the fixed-effects IVW model as the primary analytical approach for MR analysis. The distribution of IVs across these levels was as follows: 31 IVs in 2 orders, 46 IVs in 3 families, and 1,372 IVs in 3 genera. It is worth noting that all IVs exhibited a stronger association with exposure compared to the outcome, as indicated by the p -value for exposure being less than that for the outcome (exposure $<$ outcome). The systematic collection of essential information on SNPs is crucial for subsequent analysis. The essential SNP information, such as effect allele, another allele, SE, and p -value were gathered carefully and systematically to ensure accuracy and consistency in the analysis. It provides more detailed information regarding the identified related IVs.

The Impact of GM on OP From a MR Analysis

Using five MR research methods, including IVW, we causally identified eight microbial taxa associated with OP. Results suggest that family *Pasteurellaceae* [odds ratio (OR) = 1.002, $p = 0.030$], order *Pasteurellales* (OR = 1.002, $p = 0.030$), and genus *Ruminococcaceae UCG004* (OR = 1.003, $p = 0.040$) may increase OP risk. Conversely, family *Oxalobacteraceae* (OR = 0.998, $p = 0.040$), unknown family id.1000006161 (OR = 0.998, $p = 0.015$), genus *Lachnospiraceae NK4A136* group (OR = 0.996, $p = 0.003$), unknown genus id.1000006162 (OR = 0.998, $p = 0.015$), and order *NBIn* (OR = 0.998, $p = 0.015$) are negatively associated with OP, indicating reduced risk (Figure 2, Table I, **Supplementary Table III**).

Quality Control Methods

Scatter plots can assist in identifying outliers or data points that deviate from the norm. As depicted in Figure 3, it is evident that the directions calculated for the family *Lachnospiraceae NK4A136* group, family *Oxalobacteraceae*, and

genus *Ruminococcaceae UCG004* are consistent with the expected values. Moreover, the funnel plot also eliminates the potential influence of outliers. Due to the superior statistical power of the IVW method, our findings primarily rely on IVW estimation. No evidence of horizontal pleiotropy was found in the results.

Mendelian randomization pleiotropy residual sum and outlier (MR-PRESSO) can automatically identify and remove outliers and then recalculate the causal effects. We used MR-PRESSO to further validate the statistical results of the causal relationship between GM and OP. Our results demonstrate that both the MR-Egger intercept test and the MR-PRESSO global test indicate a low possibility of horizontal pleiotropy ($p > 0.05$). Additionally, the leave-one-out analysis further confirms the robustness of the data (Figure 4). This demonstrates that, in the absence of heterogeneity and pleiotropy, the results from the IVW method are reliable (See sensitivity analysis in Table II).

Discussion

To the best of our knowledge, this is the first study to explore the causal relationship between the relative abundance of 211 gut microbiota species and OP using gut microbiota genome-wide association study (GWAS) data from public databases and OP GWAS data. Using TSMR analysis, we assessed the potential causal relationship between transgenic populations and OP from a host genetic perspective, confirming their impact on OP susceptibility. Our findings discovered the causal relationship between GM and OP at the genetic level. Specifically, two families, one order, and two genera were found to be associated with low OP risk, while one family, one order, and one genus were associated with high OP risk. These findings may contribute to the future discovery of novel biomarkers in OP. These results may contribute to the future discovery of novel biomarkers for OP and provide new insights for intervention and treatment approaches by targeting specific GM populations to address ecological imbalances in OP. There is evidence to suggest that GM may have an impact on OP by modulating bone metabolism²⁶. Microbial products such as SCFA can stimulate bone formation while inhibiting bone resorption³⁷. Additionally, GM can influence bone metabolism through mechanisms such as increasing cell proliferation and differen-

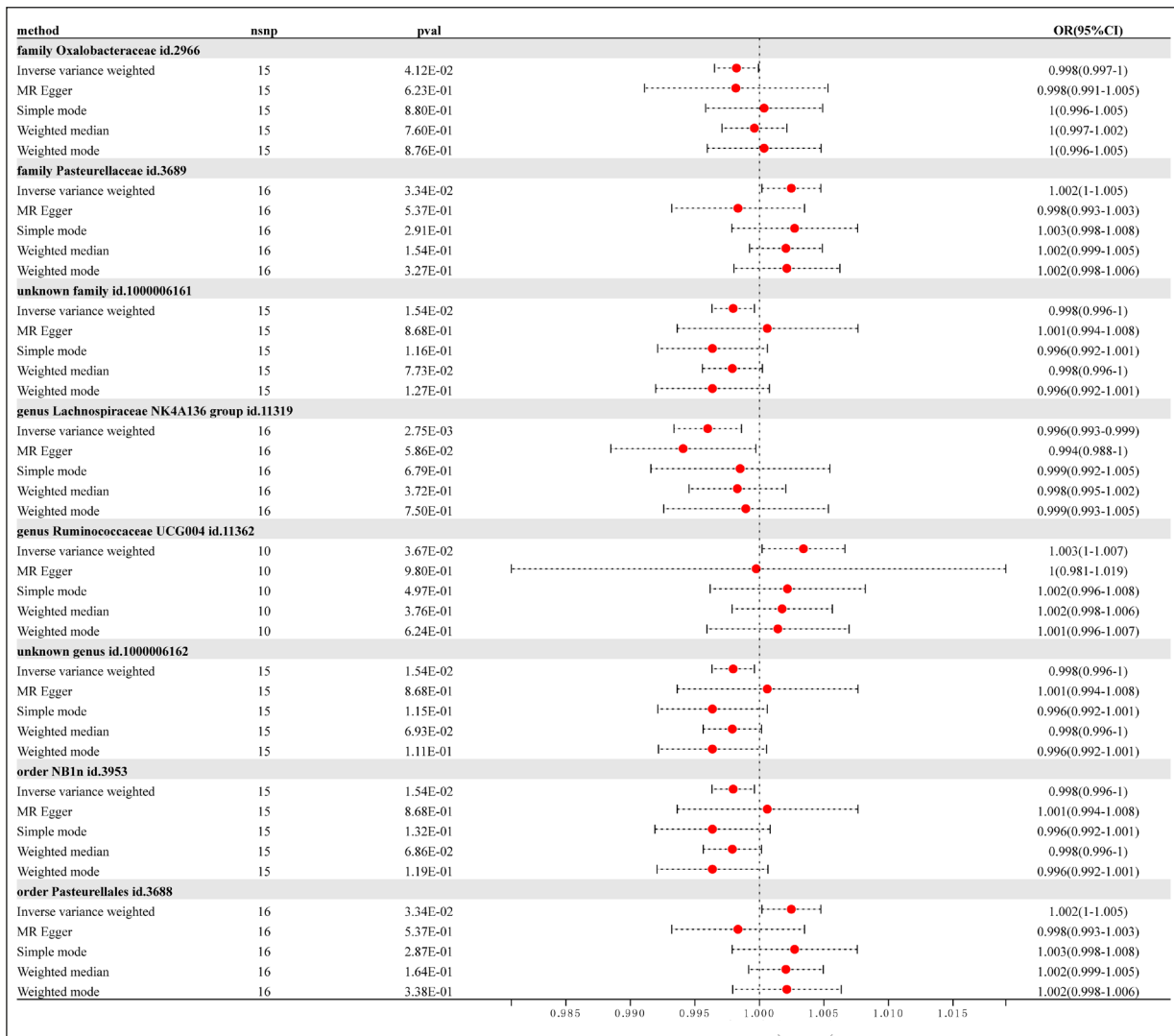


Figure 2. Forest plots depicting the results of five MR methods (Inverse variance-weighted, MR Egger, weighted median, simple mode and weighted mode) to analyze the causal relationship between GM and OP.

tiation, regulating the immune and nervous systems, and affecting hormone levels^{13,38}. Molecular products produced by intestinal bacteria can have both beneficial and harmful effects. It is known that they can influence endocrine cells, the enteric nervous system, intestinal permeability, and the immune system in the gut. The composition of GM undergoes changes as individuals age, with particularly significant variations observed in individuals aged 65 and above, which coincides with a period of high incidence of OP³⁹. GM serves as a potential source of antigens for the host immune system. However, the relationship between GM and OP is not yet fully understood. Recent studies, such as the work conducted by Yu

et al⁴⁰, have indicated that GM can promote the expansion of bone marrow T cells and increase tumor necrosis factor-alpha (TNF- α) production, suggesting that intestinal T cells could be used as targets for bone loss-associated sex steroid deficient mice. GM composition may also be involved in regulating bone mass reduction in postmenopausal women. Furthermore, the use of various types of probiotics has shown promise in preventing bone loss caused by ovariectomy⁴¹. Metabolic cofactor patterns can ameliorate bone loss by modulating gut-bone axis participants, such as probiotics, which may reverse GM-induced inflammation, enhance bone density, and improve bone quality. Probiotics can regulate the

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Table I. Causal analysis of MR results for GM vs. OP ($p < 1 \times 10^{-5}$).

Classification		No. of SNP	MR analysis method	SE	p-value	OR (95% CI)	F	
Family	<i>Oxalobacteraceae</i>	15	Inverse variance weighted	0.0008	0.0412	0.998 (0.997-1)	97.426	
			MR Egger	0.0036	0.6228	0.998 (0.991-1.005)		
			Simple mode	0.0023	0.8795	1 (0.996-1.005)		
			Weighted median	0.0013	0.7602	1 (0.997-1.002)		
			Weighted mode	0.0022	0.8762	1 (0.996-1.005)		
	<i>Pasteurellaceae</i>	16	Inverse variance weighted	0.0012	0.0334	1.002 (1-1.005)		61.793
			MR Egger	0.0026	0.5365	0.998 (0.993-1.003)		
			Simple mode	0.0025	0.2909	1.003 (0.998-1.008)		
			Weighted median	0.0014	0.1544	1.002 (0.999-1.005)		
			Weighted mode	0.0021	0.3274	1.002 (0.998-1.006)		
	<i>unknown family id.1000006161</i>	15	Inverse variance weighted	0.0008	0.0154	0.998 (0.996-1)		92.765
			MR Egger	0.0036	0.8683	1.001 (0.994-1.008)		
			Simple mode	0.0022	0.1158	0.996 (0.992-1.001)		
			Weighted median	0.0012	0.0773	0.998 (0.996-1)		
			Weighted mode	0.0023	0.1275	0.996 (0.992-1.001)		
Genus	<i>Lachnospiraceae NK4A136 group</i>	16	Inverse variance weighted	0.0013	0.0028	0.996 (0.993-0.999)	34.490	
			MR Egger	0.0029	0.0586	0.994 (0.988-1)		
			Simple mode	0.0035	0.6786	0.999 (0.992-1.005)		
			Weighted median	0.0019	0.3720	0.998 (0.995-1.002)		
			Weighted mode	0.0033	0.7501	0.999 (0.993-1.005)		
	<i>Ruminococcaceae UCG004</i>	10	Inverse variance weighted	0.0016	0.0367	1.003 (1-1.007)	47.311	
			MR Egger	0.0098	0.9802	1 (0.9817-1.019)		
			Simple mode	0.0031	0.4968	1.002 (0.996-1.008)		
			Weighted median	0.0020	0.3763	1.002 (0.998-1.006)		
			Weighted mode	0.0028	0.6241	1.001 (0.996-1.007)		
	<i>unknown genus id.1000006162</i>	15	Inverse variance weighted	0.0008	0.0154	0.998 (0.996-1)	92.765	
			MR Egger	0.0036	0.8683	1.001 (0.994-1.008)		
			Simple mode	0.0022	0.1149	0.996 (0.992-1.001)		
			Weighted median	0.0012	0.0693	0.998 (0.996-1)		
			Weighted mode	0.0022	0.1110	0.996 (0.992-1.001)		
Order	<i>NBIn</i>	15	Inverse variance weighted	0.0008	0.0154	0.998 (0.996-1)	92.765	
			MR Egger	0.0036	0.8683	1.001 (0.994-1.008)		
			Simple mode	0.0023	0.1322	0.996 (0.992-1.001)		
			Weighted median	0.0011	0.0686	0.998 (0.996-1)		
			Weighted mode	0.0022	0.1191	0.996 (0.992-1.001)		
	<i>Pasteurellales</i>	16	Inverse variance weighted	0.0012	0.0334	1.002 (1-1.005)	61.793	
			MR Egger	0.0026	0.5365	0.998 (0.993-1.003)		
			Simple mode	0.0025	0.2874	1.003 (0.998-1.008)		
			Weighted median	0.0015	0.1641	1.002 (0.999-1.005)		
			Weighted mode	0.0021	0.3383	1.002 (0.998-1.006)		

Mendelian randomization (MR), gut microbiota (GM), osteoporosis (OP), single nucleotide polymorphism (SNP).

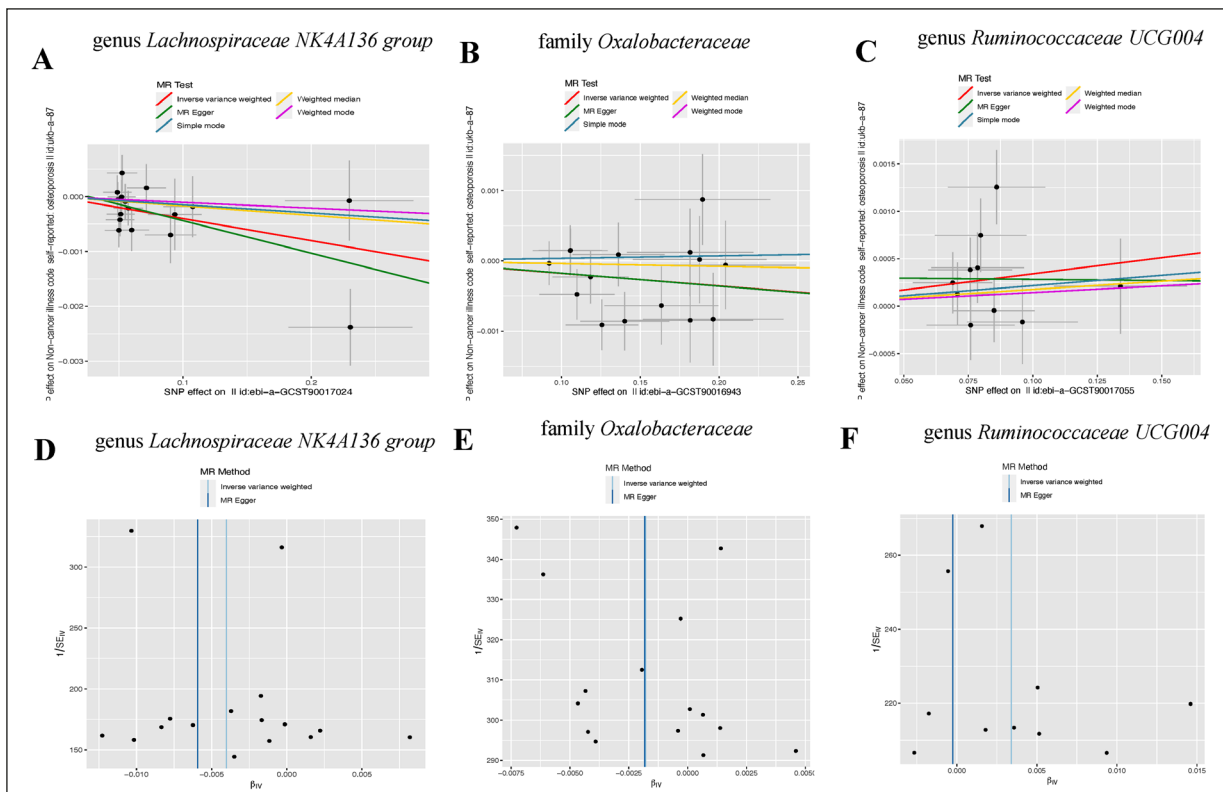


Figure 3. Scatter (A-C) and funnel (D-F) plots of the causal relationship between GM and OP. In scatter plots, the different lines represent various MR statistical methods as follows: The red line represents the Inverse variance-weighted method. The green line represents the MR Egger method. The blue line represents the Simple mode. The yellow line represents the weighted median. The purple line represents the weighted mode. In funnel plots, the different lines represent various statistical methods as follows: the deep blue line represents the Inverse variance-weighted method, while the light blue line represents the MR Egger method.

immune activity, including Insulin-like growth factor-1 (IGF-1), TNF- α , and interleukin 1-B (IL-1 β) while stimulating IL-7 and interferon-gamma (IFN- γ)⁴².

Pasteurellaceae is a Gram-negative bacterial family, including *Haemophilus influenzae*, causing respiratory infections in humans, and pathogenic to animals. It is relevant in bone research, particularly in osteomyelitis⁴³. Research⁴⁴ has found a causal relationship between microbial communities and bone development in twins, including *Pasteurellaceae* and *Lachnospiraceae*, and identified specific bacterial groups that regulate changes in bone mass. Research^{45,46} indicates that *Pasteurellaceae* may promote bone resorption by stimulating osteoclast function. Bacterial components stimulate the formation and activation of osteoclasts, leading to the destruction of bone tissue. Infection with *Pasteurellaceae* triggers an inflammatory response, disrupting normal bone metabolism and increasing the risk

of bone loss. Certain bacterial metabolites induce apoptosis in bone cells and interfere with normal cellular signaling pathways^{45,46}.

Ruminococcaceae is a family of intestinal bacteria that belongs to the anaerobic group and is predominantly found in the intestines of humans and animals⁴⁷. There is currently ongoing debate regarding the relationship between *Ruminococcaceae* and bone health.

Research indicates⁴⁸ that there is a correlation between OP and the β -diversity, taxonomy, and functional composition of the GM. The abundance of *Ruminococcaceae* is negatively correlated with the presence of OP. Additionally, another study⁴⁹ arrived at a similar conclusion, indicating that OP affects the composition of the GM. Treatment with parathyroid hormone can regulate microbial metabolic function, increase the abundance of various GMs, including *Ruminococcaceae*, and thereby reduce bone loss. Interestingly, contrary to these researches, a study on human

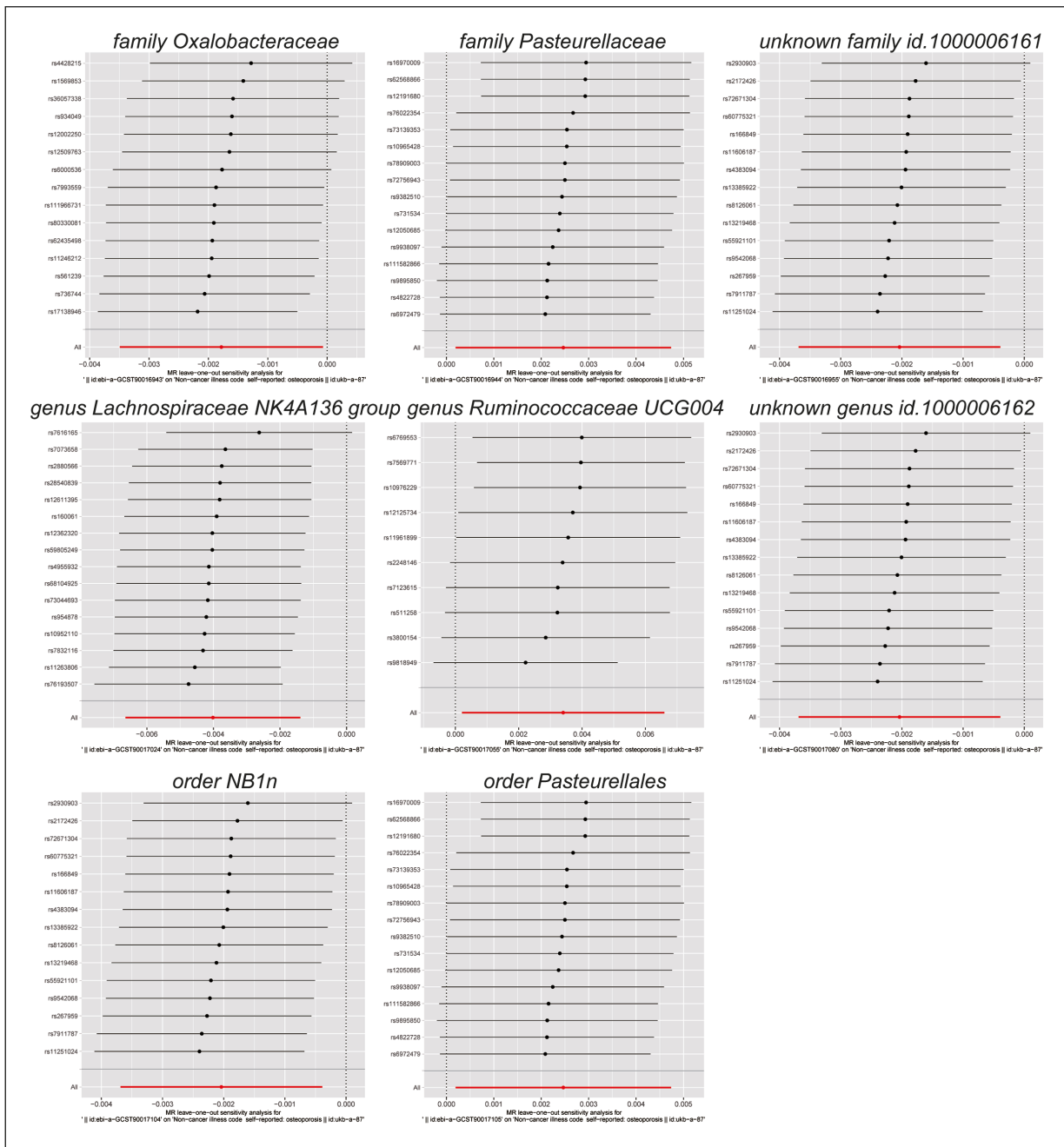


Figure 4. Leave-one-out analysis of GM on OP. Mendelian randomization (MR), gut microbiota (GM), osteoporosis (OP).

immunodeficiency virus (HIV)-infected women found that women with lower bone mineral density (BMD) had a higher relative abundance of *Ruminococcus*, which differed significantly from women with normal BMD⁵⁰. Furthermore, another study using fecal 16S rRNA gene sequencing diversity analysis found a significant increase in the abundance of the genera *Blautia*, *Eubacterium*, and *Ruminococcaceae* family in the OP

group compared to the control group. Population differences and dissimilar experimental design methods may have contributed to the opposite results⁵¹. The results of the latter two studies^{52,53} collectively indicate that *Ruminococcus* may be one of the biomarkers promoting bone loss, which is consistent with our research findings. Further functional studies are needed in the future to validate this result. The mechanisms by which the

Table II. Sensitivity analysis of MR results for GM vs. OP.

	GM exposure	No. of SNP	MR Egger interpreter	Steiger approach	MR-PRESSO global test	Cochran 'Q
			p-value	p-value	p-value	p-value
Family	<i>Oxalobacteraceae</i>	15	0.989	4.79E-67	0.364	0.349
	<i>Pasteurellaceae</i>	16	0.106	4.50E-77	0.151	0.135
	unknown family id.1000006161	15	0.460	7.02E-66	0.566	0.554
Genus	<i>Lachnospiraceae</i>	16	0.463	4.70E-68	0.329	0.365
	<i>NK4A136 group</i>					
	<i>Ruminococcaceae</i>	10	0.714	9.52E-44	0.219	0.2110
	<i>UCG004</i>					
Order	unknown genus id.1000006162	15	0.460	7.02E-66	0.577	0.554
	<i>NBIn</i>	15	0.460	7.02E-66	0.583	0.554
	<i>Pasteurellales</i>	16	0.106	4.50E-77	0.152	0.135

Mendelian randomization (MR), gut microbiota (GM), osteoporosis (OP), single nucleotide polymorphism (SNP), Mendelian randomization pleiotropy residual sum and outlier (MR-PRESSO).

Ruminococcaceae family regulates bone metabolism may be related to the fermentation of dietary fibers, which produce SCFAs. These SCFAs promote the activation of osteoblasts while inhibiting the activity of osteoclasts, thereby exerting a positive regulatory effect on bone metabolism. Furthermore, the synthesis of nutrients such as vitamin K by the *Ruminococcaceae* family may have a beneficial impact on bone metabolism^{52,53}.

Research suggests that a high-fiber diet is associated with a reduction in the levels of *Oxalobacteraceae*⁵⁴. On the other hand, a fiber-rich diet helps reduce the risk of OP. Therefore, we speculate that *Oxalobacteraceae* may regulate bone health through diet pattern⁵⁵.

Lachnospiraceae is an important member of the GM. They have the ability to degrade cellulose and other complex polysaccharides in the diet, producing beneficial SCFAs such as propionic acid, butyric acid, and acetic acid. These SCFAs serve as crucial energy sources for intestinal epithelial cells and also play a regulatory role in the immune system⁵⁶.

GM dysbiosis has been found to be associated with the development of arthritis. Research⁵⁷ has discovered that a high-fiber diet can effectively mitigate arthritis and bone erosion while simultaneously increasing T-cell and IL-10 levels. Moreover, this dietary intervention exerts its impact by inducing changes in the gut microbiota, including the modulation of the *Lachnospiraceae NK4A136 group*, ultimately resulting in elevated levels of SCFAs. In

an OVX rat model experiment, it was found that the abundance of *Lachnospiraceae bacterium 10 I* and *Lachnospiraceae bacterium A4* significantly increased⁵⁸. Another study⁵⁹ revealed that *Lachnospiraceae* were more abundant in individuals with low BMD, and it was positively correlated with BMD and T-score. This phenomenon may be related to pathways associated with lipopolysaccharide (LPS) biosynthesis. Gushudan (GSD), a traditional Chinese herbal formula for the treatment of OP, was studied⁶⁰ regarding its pharmacological mechanisms on bone metabolism. It was found that GSD can increase the abundance of bacteria *g_Lachnospiraceae nk4a136 group*, and lactate-producing bacteria *Lactobacillus*. Therefore, *Lachnospiraceae* might have bone-protective effects, which is consistent with the conclusions of our study.

This study has some highlights: we first delved into the intricate causal relationship between GM and OP using a robust combination of GWAS data and MR analysis. By employing rigorous statistical methodologies, including MR-Egger and sensitivity tests, we uncovered specific GM taxa that either increase or decrease the risk of OP, thereby pinpointing precise genetic mechanisms involved in GM-mediated OP.

Limitations

Firstly, this study only utilized GWAS data from European populations for TSMR analysis, and further research is needed in other populations. Secondly, the quantity of GM abundance

data included in this study is limited. Although the GWAS dataset used is one of the largest and most up-to-date population cohorts, additional GWAS data on GM are required to comprehensively explore the causal relationship between GM and OP. Thirdly, although TSMR is an effective method for causal relationship analysis, future animal experiments are necessary to further validate the potential causal relationship between GM and OP. Lastly, the relationship between GM and OP is not solely a single causal relationship, and the investigation of the etiology and pathogenesis of OP should be approached from multiple perspectives.

Conclusions

In summary, we have confirmed the causal effect of GM on OP through TSMR. We discovered that the family *Pasteurellaceae*, order *Pasteurellales*, genus *Ruminococcaceae* UCG004, family *Oxalobacteraceae*, and genus *Lachnospiraceae* NK4A136 group were causally associated with OP. These strains may serve as potential novel biomarkers. Furthermore, those GMs, such as the *Oxalobacteraceae* family and genus *Lachnospiraceae* NK4A136 group negatively correlated with OP may hold promising prospects for the prevention and treatment of OP. This study provides insights into the genetic level of treatment and prevention of OP.

Conflict of Interest

The authors declare that they have no conflict of interests.

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Availability of Data and Materials

All data used in this study are available in publicly available datasets. The GM data can be found here: MiBioGen con-

sortium (www.mibiogen.org, accessed on 2 May 2023), and the OP data can be found here: UKB consortium (<https://gwas.mrcieu.ac.uk/>, id: ukb-a-87, accessed on 2 May 2023).

Ethics Approval and Informed Consent

All data used in our study were based on existing publicly available summary data; no additional ethical approval was required.

Authors' Contribution

Huiqiong Zeng, Wei Liu, and Ye Zhang are responsible for conception and design. All authors are responsible for data curation. All authors are responsible for investigation. All authors are responsible for methodology. Huiqiong Zeng, Guan Li, and Ye Zhang are responsible for formal analysis. Kaixia Zhou, Aidong Li, and Ye Zhang are responsible for project administration. Aidong Li, Guan Li, and Kaixia Zhou are responsible for data analysis. All authors are responsible for resources. Guan Li, Aidong Li, and Ye Zhang are responsible for supervision. Huiqiong Zeng, and Ye Zhang are responsible for the writing-original draft. Huiqiong Zeng, and Wei Liu are responsible for the writing-review and editing. Approval of final manuscript: all authors.

References

- 1) Compston JE, McClung MR, Leslie WD. Osteoporosis. *Lancet* 2019; 393: 364-376.
- 2) Oryan A, Sahvieh S. Effects of bisphosphonates on osteoporosis: Focus on zoledronate. *Life Sci* 2021; 264: 118681.
- 3) Peng J, Chen J, Liu Y, Lyu J, Zhang B. Association between periodontitis and osteoporosis in United States adults from the National Health and Nutrition Examination Survey: a cross-sectional analysis. *BMC Oral Health* 2023; 23: 254.
- 4) Curtis EM, van der Velde R, Moon RJ, van den Bergh JP, Geusens P, de Vries F, van Staa TP, Cooper C, Harvey NC. Epidemiology of fractures in the United Kingdom 1988-2012: Variation with age, sex, geography, ethnicity and socioeconomic status. *Bone* 2016; 87: 19-26.
- 5) Kanis JA, Harvey NC, Cooper C, Johansson H, Odén A, McCloskey EV; Advisory Board of the National Osteoporosis Guideline Group. A systematic review of intervention thresholds based on FRAX : A report prepared for the National Osteoporosis Guideline Group and the International Osteoporosis Foundation. *Arch Osteoporos* 2016; 11: 25.
- 6) Tański W, Kosiorowska J, Szymańska-Chabowska A. Osteoporosis - risk factors, pharmaceutical and non-pharmaceutical treatment. *Eur Rev Med Pharmacol Sci* 2021; 25: 3557-3566.
- 7) Biver E, Berenbaum F, Valdes AM, Araujo de Carvalho I, Bindels LB, Brandi ML, Calder PC, Castronovo V, Cavalier E, Cherubini A, Cooper C, Dennison E, Franceschi C, Fuggle N, Laslop

- A, Miossec P, Thomas T, Tuzun S, Veronese N, Vaskovska M, Reginster JY, Rizzoli R. Gut microbiota and osteoarthritis management: An expert consensus of the European society for clinical and economic aspects of osteoporosis, osteoarthritis and musculoskeletal diseases (ESCEO). *Ageing Res Rev* 2019; 55: 100946.
- 8) Tu Y, Kuang X, Zhang L, Xu X. The associations of gut microbiota, endocrine system and bone metabolism. *Front Microbiol* 2023; 14: 1124945.
 - 9) Zeng H, Zhou K, Zhuang Y, Li A, Luo B, Zhang Y. Unraveling the connection between gut microbiota and Alzheimer's disease: a two-sample Mendelian randomization analysis. *Front Aging Neurosci* 2023; 15: 1273104.
 - 10) Gomaa EZ. Human gut microbiota/microbiome in health and diseases: a review. *Antonie Van Leeuwenhoek* 2020; 113: 2019-2040.
 - 11) Fan Y, Pedersen O. Gut microbiota in human metabolic health and disease. *Nat Rev Microbiol* 2021; 19: 55-71.
 - 12) Park KR, Kim EC, Hong JT, Yun HM. Dysregulation of 5-hydroxytryptamine 6 receptor accelerates maturation of bone-resorbing osteoclasts and induces bone loss. *Theranostics* 2018; 8: 3087-3098.
 - 13) Inchingolo AM, Gargiulo Isacco C, Inchingolo AD, Nguyen KCD, Cantore S, Santacroce L, Scacco S, Cirulli N, Corriero A, Puntillo F, Dipalma G, Ballini A, Inchingolo F. The human microbiota key role in the bone metabolism activity. *Eur Rev Med Pharmacol Sci* 2023; 27: 2659-2670.
 - 14) Sjögren K, Engdahl C, Henning P, Lerner UH, Tremaroli V, Lagerquist MK, Bäckhed F, Ohlsson C. The gut microbiota regulates bone mass in mice. *J Bone Miner Res* 2012; 27: 1357-1367.
 - 15) de Sire A, de Sire R, Curci C, Castiglione F, Wahli W. Role of Dietary Supplements and Probiotics in Modulating Microbiota and Bone Health: The Gut-Bone Axis. *Cells* 2022; 11: 743.
 - 16) Li J, Yang M, Lu C, Han J, Tang S, Zhou J, Li Y, Ming T, Wang ZJ, Su X. Tuna Bone Powder Alleviates Glucocorticoid-Induced Osteoporosis via Coregulation of the NF- κ B and Wnt/ β -Catenin Signaling Pathways and Modulation of Gut Microbiota Composition and Metabolism. *Mol Nutr Food Res* 2020; 64: e1900861.
 - 17) Kwon Y, Park C, Lee J, Park DH, Jeong S, Yun CH, Park OJ, Han SH. Regulation of Bone Cell Differentiation and Activation by Microbe-Associated Molecular Patterns. *Int J Mol Sci* 2021; 22: 5805.
 - 18) Ohlsson C, Nigro G, Boneca IG, Bäckhed F, Sansonetti P, Sjögren K. Regulation of bone mass by the gut microbiota is dependent on NOD1 and NOD2 signaling. *Cell Immunol* 2017; 317: 55-58.
 - 19) Yadav VK, Ryu JH, Suda N, Tanaka KF, Gingrich JA, Schütz G, Glorieux FH, Chiang CY, Zajac JD, Insogna KL, Mann JJ, Hen R, Ducy P, Karsenty G. Lrp5 controls bone formation by inhibiting serotonin synthesis in the duodenum. *Cell* 2008; 135: 825-837.
 - 20) Yan J, Herzog JW, Tsang K, Brennan CA, Bower MA, Garrett WS, Sartor BR, Aliprantis AO, Charles JF. Gut microbiota induce IGF-1 and promote bone formation and growth. *Proc Natl Acad Sci U S A* 2016; 113: E7554-E7563.
 - 21) Guo M, Liu H, Yu Y, Zhu X, Xie H, Wei C, Mei C, Shi Y, Zhou N, Qin K, Li W. *Lactobacillus rhamnosus* GG ameliorates osteoporosis in ovariectomized rats by regulating the Th17/Treg balance and gut microbiota structure. *Gut Microbes* 2023; 15: 2190304.
 - 22) Ference BA, Holmes MV, Smith GD. Using Mendelian Randomization to Improve the Design of Randomized Trials. *Cold Spring Harb Perspect Med* 2021; 11: a040980.
 - 23) Bowden J, Holmes MV. Meta-analysis and Mendelian randomization: A review. *Res Synth Methods* 2019; 10: 486-496.
 - 24) Li C, Liu C, Li N. Causal associations between gut microbiota and adverse pregnancy outcomes: A two-sample Mendelian randomization study. *Front Microbiol* 2022; 13: 1059281.
 - 25) Xu YS, Liao RY, Huang D, Wang D, Zhang L, Li YZ. Evidence from Mendelian randomization: increased risk of miscarriage in patients with asthma. *Eur Rev Med Pharmacol Sci* 2023; 27: 11587-11596.
 - 26) Skrivankova VW, Richmond RC, Woolf BAR, Yarmolinsky J, Davies NM, Swanson SA, VanderWeele TJ, Higgins JPT, Timpson NJ, Dimou N, Langenberg C, Golub RM, Loder EW, Gallo V, Tybjaerg-Hansen A, Davey Smith G, Egger M, Richards JB. Strengthening the Reporting of Observational Studies in Epidemiology Using Mendelian Randomization: The STROBE-MR Statement. *JAMA* 2021; 326: 1614-1621.
 - 27) Rusk N. The UK Biobank. *Nat Methods* 2018; 15: 1001.
 - 28) van der Velde KJ, Imhann F, Charbon B, Pang C, van Enckevort D, Slofstra M, Barbieri R, Alberts R, Hendriksen D, Kelpin F, de Haan M, de Boer T, Haakma S, Stroomborg C, Scholtens S, van de Geijn GJ, Festen EAM, Weersma RK, Swertz MA. MOLGENIS research: advanced bioinformatics data software for non-bioinformaticians. *Bioinformatics* 2019; 35: 1076-1078.
 - 29) Pierce BL, VanderWeele TJ. The effect of non-differential measurement error on bias, precision and power in Mendelian randomization studies. *Int J Epidemiol* 2012; 41: 1383-1393.
 - 30) Staley JR, Blackshaw J, Kamat MA, Ellis S, Surendran P, Sun BB, Paul DS, Freitag D, Burgess S, Danesh J, Young R, Butterworth AS. PhenoScanner: a database of human genotype-phenotype associations. *Bioinformatics* 2016; 32: 3207-3209.
 - 31) Palmer TM, Lawlor DA, Harbord RM, Sheehan NA, Tobias JH, Timpson NJ, Davey Smith G, Sterne JA. Using multiple genetic variants as instrumental variables for modifiable risk factors. *Stat Methods Med Res* 2012; 21: 223-242.

- 32) Freeman G, Cowling BJ, Schooling CM. Power and sample size calculations for Mendelian randomization studies using one genetic instrument. *Int J Epidemiol* 2013; 42: 1157-1163.
- 33) Burgess S, Thompson SG; CRP CHD Genetics Collaboration. Avoiding bias from weak instruments in Mendelian randomization studies. *Int J Epidemiol* 2011; 40: 755-764.
- 34) Bowden J, Del Greco M F, Minelli C, Davey Smith G, Sheehan NA, Thompson JR. Assessing the suitability of summary data for two-sample Mendelian randomization analyses using MR-Egger regression: the role of the I² statistic. *Int J Epidemiol* 2016; 45: 1961-1974.
- 35) Wu F, Huang Y, Hu J, Shao Z. Mendelian randomization study of inflammatory bowel disease and bone mineral density. *BMC Med* 2020; 18: 312.
- 36) Geroldinger A, Lusa L, Nold M, *Int J Epidemiol*. Leave-one-out cross-validation, penalization, and differential bias of some prediction model performance measures—a simulation study. *Diagn Progn Res* 2023; 7: 9.
- 37) Greenbaum J, Lin X, Su KJ, Gong R, Shen H, Shen J, Xiao HM, Deng HW. Integration of the Human Gut Microbiome and Serum Metabolome Reveals Novel Biological Factors Involved in the Regulation of Bone Mineral Density. *Front Cell Infect Microbiol* 2022; 12: 853499.
- 38) Delzenne NM, Rodriguez J, Olivares M, Neyrinck AM. Microbiome response to diet: focus on obesity and related diseases. *Rev Endocr Metab Disord* 2020; 21: 369-380.
- 39) Ghosh TS, Shanahan F, O'Toole PW. The gut microbiome as a modulator of healthy ageing. *Nat Rev Gastroenterol Hepatol* 2022; 19: 565-584.
- 40) Yu M, Pal S, Paterson CW, Li JY, Tyagi AM, Adams J, Coopsmith CM, Weitzmann MN, Pacifici R. Ovariectomy induces bone loss via microbial-dependent trafficking of intestinal TNF+ T cells and Th17 cells. *J Clin Invest* 2021; 131: e143137.
- 41) Parvaneh K, Ebrahimi M, Sabran MR, Karimi G, Hwei AN, Abdul-Majeed S, Ahmad Z, Ibrahim Z, Jamaluddin R. Probiotics (*Bifidobacterium longum*) Increase Bone Mass Density and Upregulate Sparc and Bmp-2 Genes in Rats with Bone Loss Resulting from Ovariectomy. *Biomed Res Int* 2015; 2015: 897639.
- 42) Jia R, Liu N, Zhu Y, Li Q. Curative Effect of Probiotics/Probiotics Preparations Combined with Zoledronic Acid+Calcitriol Regimen on Patients with Primary Osteoporosis and Their Influences on Bone Metabolism Markers. *Emerg Med Int* 2022; 2022: 3293362.
- 43) von Schroeder HP, Bell RS. *Pasteurella multocida* osteomyelitis: An unusual case presentation. *Can J Infect Dis* 1996; 7: 137-139.
- 44) Ni JJ, Yang XL, Zhang H, Xu Q, Wei XT, Feng GJ, Zhao M, Pei YF, Zhang L. Assessing causal relationship from gut microbiota to heel bone mineral density. *Bone* 2021; 143: 115652.
- 45) Maekawa S, Katagiri S, Takeuchi Y, Komazaki R, Ohtsu A, Udagawa S, Izumi Y. Bone metabolic microarray analysis of ligature-induced periodontitis in streptozotocin-induced diabetic mice. *J Periodontol Res* 2017; 52: 233-245.
- 46) Kloos B, Chakraborty S, Lindner SG, Noack K, Harre U, Schett G, Krämer OH, Kubatzky KF. *Pasteurella multocida* toxin-induced osteoclastogenesis requires mTOR activation. *Cell Commun Signal* 2015; 13: 40.
- 47) Pacios S, Andriankaja O, Kang J, Alnammary M, Bae J, de Brito Bezerra B, Schreiner H, Fine DH, Graves DT. Bacterial infection increases periodontal bone loss in diabetic rats through enhanced apoptosis. *Am J Pathol* 2013; 183: 1928-1935.
- 48) Ling CW, Miao Z, Xiao ML, Zhou H, Jiang Z, Fu Y, Xiong F, Zuo LS, Liu YP, Wu YY, Jing LP, Dong HL, Chen GD, Ding D, Wang C, Zeng FF, Zhu HL, He Y, Zheng JS, Chen YM. The Association of Gut Microbiota With Osteoporosis Is Mediated by Amino Acid Metabolism: Multiomics in a Large Cohort. *J Clin Endocrinol Metab* 2021; 106: e3852-e3864.
- 49) Zhou J, Wang R, Zhao R, Guo X, Gou P, Bai H, Lei P, Xue Y. Intermittent Parathyroid Hormone Alters Gut Microbiota in Ovariectomized Osteoporotic Rats. *Orthop Surg* 2022; 14: 2330-2338.
- 50) Mei Z, Yin MT, Sharma A, Wang Z, Peters BA, Chandran A, Weber KM, Ross RD, Gustafson D, Zheng Y, Kaplan RC, Burk RD, Qi Q. Gut microbiota and plasma metabolites associated with bone mineral density in women with or at risk of HIV infection. *AIDS* 2023; 37: 149-159.
- 51) Wang J, Wang Y, Gao W, Wang B, Zhao H, Zeng Y, Ji Y, Hao D. Diversity analysis of gut microbiota in osteoporosis and osteopenia patients. *PeerJ* 2017; 5: e3450.
- 52) Zhang J, Zhang Q, Liu H, Liu X, Yu Y, Han D, He X, Zeng P, Wang J. Soy-whey dual-protein alleviates osteoporosis of ovariectomized rats via regulating bone fat metabolism through gut-liver-bone axis. *Nutrition* 2022; 103-104: 111723.
- 53) Wang W, Cai H, Zhang A, Chen Z, Chang W, Liu G, Deng X, Bryden WL, Zheng A. *Enterococcus faecium* Modulates the Gut Microbiota of Broilers and Enhances Phosphorus Absorption and Utilization. *Animals (Basel)* 2020; 10: 1232.
- 54) Cao RR, He P, Lei SF. Novel microbiota-related gene set enrichment analysis identified osteoporosis associated gut microbiota from autoimmune diseases. *J Bone Miner Metab* 2021; 39: 984-996.
- 55) Rizzoli R, Biver E, Brennan-Speranza TC. Nutritional intake and bone health. *Lancet Diabetes Endocrinol* 2021; 9: 606-621.
- 56) Vacca M, Celano G, Calabrese FM, Portincasa P, Gobbetti M, De Angelis M. The Controversial

- Role of Human Gut Lachnospiraceae. *Microorganisms* 2020; 8: 573.
- 57) Bai Y, Li Y, Marion T, Tong Y, Zaiss MM, Tang Z, Zhang Q, Liu Y, Luo Y. Resistant starch intake alleviates collagen-induced arthritis in mice by modulating gut microbiota and promoting concomitant propionate production. *J Autoimmun* 2021; 116: 102564.
- 58) Wang N, Meng F, Ma S, Fu L. Species-level gut microbiota analysis in ovariectomized osteoporotic rats by Shallow shotgun sequencing. *Gene* 2022; 817: 146205.
- 59) Li C, Huang Q, Yang R, Dai Y, Zeng Y, Tao L, Li X, Zeng J, Wang Q. Gut microbiota composition and bone mineral loss-epidemiologic evidence from individuals in Wuhan, China. *Osteoporos Int* 2019; 30: 1003-1013.
- 60) Tong L, Feng Q, Lu Q, Zhang J, Xiong Z. Combined ¹H NMR fecal metabolomics and 16S rRNA gene sequencing to reveal the protective effects of Gushudan on kidney-yang-deficiency-syndrome rats via gut-kidney axis. *J Pharm Biomed Anal* 2022; 217: 114843.