

Killing two birds with one stone: miR-126 involvement in both cancer and atherosclerosis

Q.-Y. YANG, Q. YU, W.-Y. ZENG, M. ZENG, X.-L. ZHANG, Y.-L. ZHANG, L. GUO, X.-J. JIANG, J.-L. GAN

School of Integrative Medicine, Tianjin University of Traditional Chinese Medicine, Tianjin, China

Q.-Y. Yang and Q. Yu have contributed equally to this work and share first authorship

Abstract. – **OBJECTIVE:** Both cancer and atherosclerosis are the main causes of morbidity and mortality in the world, and some patients even suffer from both of them. Several studies have shown an association between the pathogenesis of cancer and atherosclerosis. It has been reported that miR-126 may participate in the pathological process of cancer and atherosclerosis. Therefore, we aimed to summarize the role of miR-126 in cancer and atherosclerosis respectively, as well as a possible association between them.

MATERIALS AND METHODS: In this paper, “miR-126” and “microRNA-126” are used as the first group of keywords, “atheromatosis” and “atherosclerosis” are used as the second group of keywords, and “tumor” and “cancer” are used as the third group of keywords. In PubMed, the authors selected one of the first group and the second group of keywords to search the literature related to miR-126 and cancer, and one of the first group and the third group of keywords was selected to search the literature on miR-126 and atherosclerosis. All collected articles are from 2021 and before. Irrelevant, withdrawn and review articles were excluded, and the included literature was mainly in the recent five years.

RESULTS: After collection and summary, miR-126 is found involved in cell apoptosis, proliferation, angiogenesis, inflammation, and other processes in both cancer and atherosclerosis by negatively targeting PI3K, VEGF, VCAM-1, EGFL7, CXCL12-CXCR4 axis, and LRP6. Moreover, we briefly review the prospects of miR-126 as a biomarker for the diagnosis and treatment of cancer and atherosclerosis in clinical applications.

CONCLUSIONS: It has been demonstrated that miR-126 can influence cancer and atherosclerosis by affecting the same or different target genes. Therefore, it facilitates our understanding of the common prevention and treatment strategies of cancer and atherosclerosis by regulating the miR-126-target genes network.

Key Words:

MiR-126, Cancer, Atherosclerosis, Target genes.

Introduction

Currently, cancer and atherosclerosis are seriously threatening human health, especially for the middle-aged and the elderly, whose mortality rates are on the rise^{1,2}. However, the pathogenesis of cancer and atherosclerosis has not been fully explained, affecting their diagnosis and treatment. Humans are diagnosed and treated with cancer in their advanced years, during which atherosclerosis is highly prevalent³. As a result, some patients suffer from both atherosclerosis and cancer at the same time. Further studies have found that cell proliferation and apoptosis, angiogenesis, inflammation, and so on are also involved in cancer and atherosclerosis⁴, which suggests a potential association between of them. Are there potential common regulatory mechanisms and therapeutic targets for these pathological and pathophysiological changes?

MicroRNAs (miRNAs) are highly conserved small non-coding RNAs with a length of about 22 nt⁵. Despite accounting for only 1-5% of the human genome, miRNAs can negatively regulate the expression of at least 30% of protein-coding genes by degrading or inhibiting mRNA translation⁶. MiRNAs play an important regulatory role in the progression of cell proliferation, apoptosis, differentiation, migration, and inflammation⁷, which is involved in the pathophysiological mechanisms of both cancer and atherosclerosis⁸. Recently, the role of miRNAs in cancer progression and metastasis has been continuously clarified⁹. Simultaneously, evidence also shows that miRNAs regulate the initiation, development, and prognosis of atherosclerosis¹⁰. MiR-126, as an endothelial-specific miRNA, is thought to be involved in several diseases, such as diabetes^{11,12}, Parkinson's disease¹³, viral myocarditis¹⁴, and ischemic stroke¹⁵. MiR-126 may attenuate diabe-

tes^{11,12}, Parkinson's disease¹³, and ischemic stroke progression¹⁵, while increasing the risk of viral myocarditis¹⁴. Besides the diseases mentioned above, miR-126 is considered closely related to cancer and atherosclerosis. MiR-126 is downregulated in malignant cancer cells such as endocrine glands cancer¹⁶, reproductive system cancer¹⁷, digestive system cancer¹⁸, and respiratory system cancer¹⁹; interestingly, miR-126 is also down-regulated in most atherosclerotic patients, as well as animal and cell models of atherosclerosis²⁰, suggesting that miR-126 has a dual role in anti-cancer and anti-atherosclerosis.

MiR-126 was first discovered in the mouse (*Mus musculus*) heart by sequencing in 2002²¹. Langraf²² found that miR-126 is tissue-specifically expressed in the hematopoietic system, respiratory system, digestive system, and reproductive system, especially in the cardiovascular system. Harris et al²³ found that miR-126 is highly expressed in endothelium-rich lung and heart tissues, primary cultured endothelial cells, but not in vascular smooth muscle cells and leukocyte cell lines. MiR-126 is encoded by the intron of epidermal growth factor-like domain 7 (EGFL7) genes²⁴, which is almost entirely expressed by endothelial cells. Based on the phenomenon above, miR-126 is almost completely derived from endothelial cells. Pre-miR-126 is processed into two mature subtypes, miR-126-3p and miR-126-5p, both of which are abundant in endothelial cells. MiR-126 regulates the expression of many different target genes, and its target genes are also regulated by a variety of miRNAs, which determines the diversity and complexity of the role of miR-126 in different diseases²⁵. Therefore, although miR-126 is generally down-regulated in atherosclerosis and various cancers, its role may have both similarities and differences. The key lies in the difference between the diseased tissue microenvironment and its downstream targets. Both cancer and atherosclerosis belong to complex diseases that share some pathophysiological mechanisms⁴. The multiple targets of miR-126 may better explain the mechanistic basis behind the common mechanism of cancer and atherosclerosis. So, it is necessary to review and pool the available literature on this topic.

This review summarizes the relevant literature in recent years, emphasizes the role of miR-126 in cancer and the target genes of intervention, and tries correlating these with the targets and functions of miR-126 in the cardiovascular system. Understanding the role of miR-126 in cancer may help us understand its key function in cells of the

cardiovascular system, to explore the common prevention and treatment targets.

MiR-126 in the Pathogenesis of Cancer

Cancer originates from normal cells and is the result of cumulative mutations in genes responsible for growth and differentiation. Compared with the normal tissue cells of its origin, cancer has the following characteristics: 1) Uncontrollable growth, indefinite proliferation (continuous division); 2) Local infiltration and distant metastatic spread. Studies found that there are miRNAs disorders in cancer, among which miR-126 is down-regulated in a variety of cancers, such as non-small cell lung cancer^{19,26} breast cancer²⁷, liver cancer²⁸, esophageal cancer²⁹, colorectal cancer³⁰ and prostate cancer²⁵. Further studies found that miR-126 can act as a cancer suppressor^{31,32} by negatively regulating target genes, including PI3K, VEGF, VCAM-1, EGFL7, and LRP6, inhibiting cancer growth and metastasis in multi-step that is closely related to their prognosis.

MiR-126 Inhibits Cancer Progression by Targeting PI3K

PI3K is composed of the regulatory subunit (p85) encoded by PIK3R2 and the catalytic subunit (p110) encoded by PIK3CA³³, which is an intracellular phosphatidylinositol kinase. PI3K can be activated by various mitotic signals and is related to oncogene products, such as v-src and v.ras³⁴. Recent studies found that miR-126 is down-regulated and PIK3R2 is up-regulated in human non-small cell lung cancer A549 cell line²⁶ and prostate cancer tissue²⁵, and miR-126 and PIK3R2 are inversely correlated. Similarly, normal expression of miR-126 in the colon can negatively regulate the regulatory subunit p85 β of PI3K, thus maintaining it at a low level, while miR-126 deletion in colon carcinogenesis reduces the targeted inhibition of p85 β and boosting PI3K signaling³⁵.

Dual-Luciferase reporter results confirm that PIK3R2 is a direct target of miR-126, which negatively regulates the expression of the p85-regulatory subunit of PI3K in various cancer cell lines, such as non-small cell lung cancer cell line A549²⁶, human esophageal cancer cell line EC109¹⁸, liver cancer Hep-G2 and BEL-7402 cell line²⁸, colorectal cancer cell line LS174T and DLD³⁵, breast can-

cer SKBR3/TR³⁶ and prostate cancer cell line²⁵. Transcriptomics analysis also unraveled that miR-126 can target PIK3CA, the gene encoding the catalytic subunit p110 of PI3K³⁷. Growth factor signal PI3K, plays a crucial role in cell proliferation, survival, metabolism, and apoptosis³³, which involves cancer initiation and development, such as apoptosis and proliferation of cancer cells, metastasis and invasion, angiogenesis and drug resistance.

MiR-126/PI3K and Cancer Apoptosis and Proliferation

Apoptosis can inhibit cancer growth, but cancer cells can escape from apoptosis through some mechanisms; excessive proliferation, even continuous proliferation, is a characteristic of cancer cell³⁸. Consequently, insufficient apoptosis and excessive proliferation are both involved in cancer growth. Studies showed that miR-126 can inhibit cancer cell proliferation and induce apoptosis by targeting PIK3R2^{17,18,25,26,39}.

In vitro, miR-126 reduces cell viability of endometrial cancer RL95 and HEC1A cell lines, induces G1/S phase arrest as well as caspase-3-mediated apoptosis; *In vivo*, miR-126 induces regression of cervical cancer that depends on at least in part on PIK3R2 signaling¹⁷. Overexpression of miR-126-3p promotes apoptosis and inhibits the proliferation of cervical cancer HeLa cells via regulation of the PI3K/PDK1/Akt signaling pathway³⁹. Similarly, up-regulation of miR-126 in non-small cell lung cancer A549 cell line can down-regulate PIK3R2, PI3K, and p-Akt protein, and then up-regulate the expression of cancer suppressor gene PTEN, which reduces the proliferation ability of cancer cells²⁶. Further studies showed that miR-126 overexpression inhibited the proliferation of esophageal cancer cell line EC10917 and prostate cancer cell line, by negatively regulating target genes PIK3R2 and then PI3K/Akt signaling pathway²⁵. In summary, miR-126 promotes cancer cell apoptosis, inhibiting proliferation by directly regulating PI3K and then its downstream signal Akt, effector molecules such as PTEN and caspase-3.

MiR-126/PI3K and Cancer Invasion and Migration

Local invasion and distant metastasis to other organs are hallmarks of malignancy. The activation of cancer invasion and migration ability is the key factor of cancer cell dissemination. Studies showed that miR-126 and its target gene PI3K are

involved in the invasion and migration of some cancers^{18,26,39-41}.

Overexpression of miR-126 in esophageal cancer cell line EC109 inhibits cancer cell migration via negative regulating PIK3R2 and then PI3K/Akt signaling pathway¹⁸. Similarly, miR-126-3p overexpression by lentivirus-mediated transfection inhibits the migration of HeLa cells by regulating the PI3K/PDK1/Akt signaling pathway³⁹. The anillin actin-binding protein (ANLN) highly expressed in various cancers, is closely related to cancer metastasis and overall short survival⁴². In three mRNA datasets from GEO, GSE18842, GSE19804, and GSE101929, it was found that ANLN was significantly up-regulated in human non-small cell lung cancer A549 cell line and was inversely correlated with miR-126⁴³. Related studies confirmed that ANLN is regulated by PI3K/Akt signaling pathway⁴¹, suggesting that miR-126 can regulate PI3K/Akt signaling and its downstream effector ANLN to inhibit lung cancer metastasis. Similarly, miR-126 also down-regulates PI3K/Akt signaling in non-small cell lung cancer A549 cell line, thereby up-regulating the expression of cancer suppressor gene PTEN, resulting in decreased migration and invasion²⁶. Epithelial-mesenchymal transition (EMT) is considered the initial step of cancer metastasis. MiR-126 mimics-transfected lung cancer cell lines SPC-A1 and LLC showed an upregulation of p-PDK1 and p-Akt, and a downregulation of Snail and a pro-EMT transcription factor protein, thus EMT induced by TGF- β 1 was inhibited⁴⁰; Snail has been confirmed to be regulated by PI3K/Akt signaling⁴⁴, that suggests that miR-126 inhibits lung cancer EMT and metastasis by regulating PI3K/Akt/Snail signaling pathway. To sum up, miR-126 can inhibit cancer metastasis by targeting PI3K and its downstream signal Akt, PDK1, and effector molecules such as ANLN, PTEN, and Snail.

MiR-126/PI3K and Cancer Chemoresistance

Chemoresistance has become a major obstacle to cancer treatment and a main limiting factor for the cure of cancer patients⁴⁵. MiR-126 can reduce the drug resistance of gastric cancer⁴⁶, breast cancer⁴⁷, and other cancers by targeting PI3K. Resistance to chemotherapy is closely related to the upregulation of multidrug resistance-associated protein (MRP)⁴⁶. Studies found that miR-126 can inhibit PI3K/Akt/MRP1 pathway by targeting PIK3R2, thereby reducing the resistance of gastric cancer to cisplatin⁴⁶. Overexpression of lncRNA HOTAIR weakens the effect of miR-126 on drug

resistance by the directly binding and inhibiting of miR-126 and then activating the PI3K/Akt/MRP1 pathway⁴⁶. These results demonstrate that miR-126 attenuates cisplatin resistance by regulating the PI3K/Akt/MRP1 pathway.

Collectively, miR-126 plays an important role in inhibiting cancer apoptosis and proliferation, invasion, and metastasis, and reducing drug resistance. PIK3R2 is an important target gene for miR-126 in inhibiting cancer development by interfering with PI3K and its different downstream signal and effector molecules.

MiR-126 Inhibits Cancer Process by Targeting VEGF

Vascular endothelial growth factor (VEGF) is a highly specific vascular endothelial cell growth factor. The VEGF family includes VEGF-A—VEGF-E and placenta growth factor (PGF), of which VEGF-A is the most important, so VEGF is referred to as VEGF-A in general⁴⁸. VEGFR1/2/3, three receptor tyrosine kinases, mediates the biological function of the VEGF family. Low expression of miR-126 was accompanied by high expression of VEGF in various cancer tissues^{49,50}. Interestingly, the degree of VEGF overexpression is not the same in different types and stages of progression of the same cancer. For example, VEGF expression is higher in triple-negative breast cancer tissues than that in non-triple-negative breast cancer tissues, implying a close relationship to the occurrence, development, and prognosis of cancers⁵¹. Combining the results from both overexpression and inhibition experiments further revealed that VEGF is significantly inversely related to miR-126 in liver cancer⁵², ovarian cancer⁵³, breast cancer⁵⁴, and gastric cancer⁵⁵. In oral squamous cell carcinoma OSCC cell lines, the role of miR-126 is related to the regulation of VEGF-A, but not VEGF-C and VEGF-D³². The expression of VEGF-A is significantly down-regulated by exogenous miR-126 in some thyroid cancer cell lines¹⁶. MiR-126 inhibits the effect of VEGF-A and VEGFR-2 on lung cancer, which vanishes when miR-126-5p, an active form of miR-126, is inhibited⁵⁰, suggesting that VEGF is downregulated by miR-126.

Targeting VEGF-A by miR-126 was predicted by bioinformatics analysis in ovarian cancer⁵³ and gastric cancer⁵⁵. The dual-luciferase reporter assay further confirmed that VEGF-A is a target gene of miR-126 in gastric cancer⁵⁵. It is well known that VEGF is necessary for cancer angiogenesis, growth, and metastasis, suggesting that

miR-126 is involved in pathological processes, such as apoptosis, proliferation, metastasis, invasion, and angiogenesis of cancer cells by targeting VEGF.

MiR-126/VEGF and Cancer Proliferation and Apoptosis

MiR-126 binds directly to VEGF and negatively regulates its expression, which is another important mechanism of miR-126 by inhibiting proliferation and promoting apoptosis in many cancers^{50,56,57}. Down-regulation of miR-126 and up-regulation of VEGF-A and VEGFR-2 inhibit the apoptosis of NCI-H1299-human non-small cell lung cancer cell line; conversely, miR-126-5p overexpression inactivates VEGF-A and VEGFR-2/ERK signaling pathway and promotes apoptosis⁵⁰. LV-miR-126 mimics induce cell cycle arresting in the G1 phase and down-regulates VEGF in ovarian cancer SKOV3 cell line⁵⁶. LV-has-miR-126 inhibitor transfected ovarian cancer SKOV3 cell line showed upregulation of VEGF and increased number of S phase cells⁵⁶. In summary, miR-126 affects the cell cycle, thus inhibiting the proliferation and promoting apoptosis of cancer cells by targeting and negatively regulating VEGF and its downstream signals.

MiR-126/VEGF and Cancer Invasion and Migration

VEGF overexpression promotes cancer angiogenesis and metastasis, and is involved in the invasion and migration of breast cancer⁵⁸, ovarian cancer⁵³, and non-small cell lung cancer⁵⁰. As mentioned above, miR-126 can bind to and inhibit VEGF, suggesting that miR-126 is expected to inhibit cancer invasion and migration by targeting VEGF.

Studies found that low expression of miR-126 and high expression of VEGF in breast cancer are more prone to metastasis: ectopic expression of miR-126 led to a significant decrease in VEGF expression at 7 days and 14 days after surgery of the primary breast cancer in mice, inhibiting its lung metastasis⁵⁸. At the same time, in NCI-H1299, a human non-small cell lung cancer cell line, miR-126 was down-regulated and its target gene, VEGF-A, and VEGFR-2 were up-regulated, which promoted cancer cell invasion⁵⁰. At the same time, the ectopic expression of VEGF-A can offset the cancer invasion induced by miR-126⁵³. Similarly, LV-miR-126 mimics were found to inhibit cell invasion and down-regulate VEGF expression in the SKOV3-ovarian cancer cell line; while LV-has-miR-126 inhibitors have the oppo-

site effect⁵⁶. In lung cancer cell lines, miR-126 inactivates VEGF-A/VEGFR-2/ERK signaling pathway, and then inhibits metastasis⁵⁰. To sum up, the downregulation of miR-126 leads to abnormal accumulation of VEGF-A and activation of its downstream signals. In contrast, the up-regulation of miR-126 is expected to negatively regulate VEGF-A, thereby inhibiting cancer invasion and migration.

MiR-126/VEGF and Cancer Angiogenesis

Angiogenesis is necessary for cancer growth and progression. VEGF, as the most effective activator of angiogenesis, promotes cancer angiogenesis by binding to VEGF receptors on vascular endothelial cells⁵⁹. Meanwhile, miR-126, specifically expressed in vascular endothelial cells, is also highly related to cancer angiogenesis. MiR-126 inhibits angiogenesis partly by targeting and negatively regulating VEGF in multiple cancers, such as oral squamous cell carcinoma³², liver cancer⁵², gastric cancer⁵⁵, and breast cancer⁵⁷.

In vivo studies found that microvessels density and VEGF-A expression were negatively correlated with miR-126 in gastric cancer tissue⁵⁵. Experiments combining overexpression and inhibition of miR-126 found that miR-126 suppressed hepatocellular carcinoma growth by inhibiting VEGF expression and subsequent angiogenesis both *in vivo* and *in vitro*⁵². Upon being restored or inhibited by lentiviral transfection, miR-126 was confirmed to inhibit the growth and angiogenesis of gastric cancer by targeting VEGF-A and then regulating the activity of its downstream signals, such as Akt, mTOR, and ERK1/2 in gastric cancer cell lines SGC-7901, MKN-28 and MKN-45⁵⁵. Transfection of miR-126 into breast cancer cell line MCF inhibits VEGF-A signaling pathway, reduces cancer angiogenesis, and delay its growth⁵⁷. However, anti-VEGF monotherapy is not effective in improving the survival rate of breast cancer⁵⁷. Resistance to VEGF inhibition may be due to the fact that angiogenic factors such as FGF-2 and IL-6 are upregulated to compensate for VEGF⁶⁰. Therefore, it is speculated that miR-126 will have superior anticancer efficacy compared with VEGF inhibitors alone at similar concentrations since miR-126 has a multi-target anti-cancer effect besides targeting VEGF.

MiR-126/VCAM-1 and Cancer Progression

Vascular cell adhesion molecule-1 (VCAM-1), a member of the immunoglobulin superfamily, plays an important role in the immune surveil-

lance of various diseases⁶¹. VCAM-1 is expressed at low levels in resting endothelial cells but increases dramatically on the luminal surface and intercellular interface of endothelial cell membrane under the strong induction of many factors, especially inflammatory factors⁶². VCAM-1 plays a key role in the inflammatory response by mediating leukocyte adhesion and rolling, as well as recruitment to the inflammatory site⁶². Interestingly, high expression of VCAM-1 was also detected in malignant cancers such as renal carcinoma⁶³, gastric carcinoma⁶⁴, colorectal carcinoma⁶⁵, prostate carcinoma⁶⁶, chondrosarcoma⁶⁷, and leukemia⁶⁸, suggesting a close relationship between VCAM-1 and cancer progression, especially cancer metastasis and angiogenesis⁶⁹. Contrary to the high expression of VCAM-1, miR-126 is expressed at low levels in a variety of cancers. It was revealed that miR-126 and VCAM-1 were negatively correlated by overexpressing or inhibiting miR-126 in malignant cancers such as colon cancer⁶⁵, prostate cancer⁶⁶, and chondroma⁶⁷, and VCAM-1 was one of the target genes of miR-126. In a word, miR-126 can also target VCAM-1 to inhibit cancer metastasis and invasion, angiogenesis, and cancer-related inflammation.

MiR-126/VCAM-1 and Cancer Metastasis and Invasion

Cancer cells release cytokines, and their concurrent inflammation was activated, which promotes the up-regulation of VCAM-1⁷⁰. Combined with animal model and clinical sample analysis, a group of 18 genes was identified as lung metastasis gene characteristics (LMS), among which VCAM-1 was listed⁷¹, suggesting a strong relationship between VCAM-1 and cancer metastasis⁶¹. Similarly, the high expression of VCAM-1 in breast cancer contributes to its ability to metastasize to lungs. Further studies have found that up-regulation of VCAM-1 expression in mice can also promote lung metastasis of vaccinated cells, whereas antibodies against VCAM-1 can significantly reduce this metastasis rate⁷². Upon entering the blood stream, cancer cells will form platelet-cancer cell aggregates that can up-regulate the expression of VCAM-1 and easily get stuck in small blood vessels. Cancer cells were then closely combined with the walls of blood vessels by VCAM-1-mediated, which is a prerequisite for cancer metastasis⁷². Therefore, the region of VCAM-1 overexpression tended to present at the site of cancer cell adhesion and early aggregation in the circulation. Integrin alpha4 expressed by

cancer cells, such as melanoma cells, can interact with VCAM-1 of endothelial cells, triggering the activation of Rac1 (a Rho-like GTPase), leading to cytoskeletal rearrangement⁷³. It is believed to remodel the tight junctions between vascular endothelial cells, thus promoting the transendothelial migration of cancer cells⁷³. Direct evidence shows that miR-126 targets VCAM-1 and is involved in the metastasis and invasion of a variety of cancers, including prostate cancer⁶⁶, leukemia⁶⁸, and chondroma⁶⁷.

Most patients with advanced prostate cancer (PCa) will eventually develop bone metastases that lead to intractable pain⁶⁶. Wnt-1-inducible secretory protein 1 (WISP-1) is a member of the homocysteine protein 61/connective tissue growth factor/nephroblastoma overexpressed gene family and is considered a factor promoting bone metastasis⁷⁴. Osteoblasts transfected with WISP-1 shRNA can promote migration and VCAM-1 expression of human PCa cell lines PC3 and DU145 mediated by the osteogenic medium⁶⁶. Osteogenic-derived WISP-1 inhibits miR-126 expression, while miR-126 mimics reverse WISP-1-promoted prostate cancer bone metastasis and VCAM-1 expression. Overall, miR-126/VCAM-1 is involved in WISP-1-induced prostate cancer bone metastasis⁶⁶. Moreover, by co-culturing LAMA84-chronic myeloid leukemia cells with endothelial cells, it was found that miR-126 can shuttle in endothelial cells, negatively regulating the expression of VCAM-1, motility, and adhesion of LAMA84⁶⁸. It is suggested that miR-126 inhibits the metastasis of leukemia by inhibiting VCAM-1 expression⁶⁸. As a primary malignant cancer of bone, chondrosarcoma is prone to local invasion as well as distant metastasis, especially lung metastasis. Naringin can enhance miR-126 expression, reduce VCAM-1 expression, and inhibit movement and invasion of chondrosarcoma cells, while miR-126 inhibitor weakens the effects of Naringin. It is suggested that miR-126 may be involved in the migration and invasion of chondrosarcoma by down-regulating VCAM-1⁶⁷. To sum up, targeting VCAM-1 is also one of the important mechanisms of miR-126 inhibiting cancer metastasis and invasion.

MiR-126/VCAM-1 and Cancer Angiogenesis

Studies have shown that VCAM-1 is highly related to cancer angiogenesis. Yong-Bin et al⁶⁹ reported that microvessel density in VCAM-1 positive cancer tissues was higher than that in

VCAM-1 negative cancer tissues of gastric cancer. The interaction of $\alpha 4\beta 1$ integrin, the main binding partner of VCAM-1, with the Ig-like domains 1 or 4 of VCAM-1 is critical for cancer angiogenesis⁷⁵.

Garmy-Susini firstly observed that VCAM-1 and $\alpha 4\beta 1$ integrin were expressed on endothelial cells and vascular smooth muscle cells respectively in developing vessels of breast cancer, whereas anti-mouse VCAM-1 antibody (M/K-2) reduced microvessel formation in a mouse model⁷⁵. Additional literature has shown that VCAM-1 antibody blocks IL-4/13-induced angiogenesis *in vitro*, while angiogenesis induction by IL-4 and IL-13 is also inhibited by anti-integrin $\alpha 4$ antibodies *in vivo*⁷⁶. Recently, the Ig-like domain 6 of VCAM-1 (VCAM-1-D6) was identified as a potential angiogenesis target. Low expression of VCAM-1 induced by siRNA-mediated VCAM-1 knockdown was found to reduce TNF- α -induced HUVEC migration and angiogenesis. Competition assays showed that TNF- α -induced HUVEC tube formation was specifically inhibited by VCAM-1-D6 fused to Fc, but not by Fc alone, suggesting that VCAM-1-D6 is a critical domain for TNF- α -induced angiogenesis⁷⁷. These data suggest that VCAM-1 may be a key target for the regulation of cancer angiogenesis. However, although a large piece of literature has confirmed that miR-126 negatively regulates VCAM-1 in cancers⁷⁸, the direct evidence that miR-126 inhibits cancer angiogenesis by negatively regulating VCAM-1 is obviously insufficient, and further research is needed.

MiR-126/VCAM-1 and Cancer-associated Inflammation

In recent years, cancer and inflammation have been found to be closely related, in which inflammatory cells act as a bridge⁷⁹. There are a lot of macrophages around the cancer cells, called cancer-associated macrophages (TAM), which express VCAM-1 during differentiation⁸⁰.

Studies have shown that CCL18 secreted by TAMs promotes the malignant progression of pancreatic cancer and induces glycolytic phenotype transformation, partly due to induction of VCAM-1 paracrine. Conversely, VCAM-1 induces lactate production and enhances aerobic glycolysis in pancreatic cancer cells, activates macrophages, and makes them form a TAM-like phenotype. There is a positive feedback loop between VCAM-1 and TAM. Further study has confirmed that TAM promotes the progression of pancreatic ductal adenocarcinoma

and the Warburg effect by the CCL18/NF- κ B/VCAM-1 pathway⁸¹. In addition, TAMs infiltration increased and VCAM-1 upregulated, cancer cell apoptosis decreased, and proliferation index increased in breast cancer after electronic cigarette exposure that affects breast cancer cell survival by modulating VCAM-1 and integrin α 4 β 1 through direct interaction with infiltrating macrophages⁸². Other studies showed that VCAM-1 expression induced by IL-1 β in glioblastoma may regulate the adhesion between glioblastoma and monocytes⁸³. Therefore, VCAM-1 is closely related to cancer-related inflammatory factors and inflammatory cells⁸³. As mentioned above, VCAM-1 is involved in cancer survival, proliferation, metastasis, and other pathological processes by affecting TAM. MiR-126 has been shown to target negative VCAM-1 in a variety of cancers, however, direct evidence that miR-126 affects cancer growth by targeting VCAM-1 in cancer-associated inflammation is insufficient.

MiR-126/EGFL7 and Cancer Progression

EGFL7 is a protein molecule containing an N-terminal signal peptide, a cysteine-rich EMI domain (named for its discovery in the EMIL protein family), and two epidermal growth factor-like domains⁸⁴, which belongs to the epidermal growth factor (EGF)-like protein family. EGFL7 is secreted specifically by endothelial cells⁸⁵ that is highly expressed in normal embryonic tissues, but barely expressed in mature tissues. Interestingly, EGFL7 is highly expressed in endothelial cells of cancer tissues⁸⁶, suggesting its importance in the occurrence and development of cancer. Although miR-126 expression is higher and EGFL7 is lower in hepatocellular carcinoma tissues (HCC) than that in normal tissues adjacent to HCC⁸⁷; miR-126 also is found to increase the expression of EGFL7 in lung cancer-associated cell line A549⁸⁶, the expression of miR-126 was significantly low and the expression of EGFL7 was high in most cancers, indicating a suggestive inverse association between miR-126 and EGFL7⁸⁸⁻⁹¹. Bioinformatics prediction further revealed that EGFL7 is a potential target of miR-126^{86,92}, which was confirmed by luciferase reports in ovarian cancer tissues⁸⁸. More and more evidence showed that miR-126 plays a crucial role in cancer biology by negative regulation targeting EGFL7, participating in cancer cell proliferation, apoptosis, metastasis, invasion, and angiogenesis to affect the pathological process of cancers.

MiR-126/EGFL7 and Cancer Proliferation and Apoptosis

EGFL7, as a potential biomarker of malignancy, is overexpressed in colorectal cancer⁹³. The transfection of EGFL7 siRNA significantly decreased the proliferation of SW620 and LoVo cells-colorectal cancer cell lines; at the same time, EGFL7 inhibits anoikis by regulating anoikis marker proteins, which is related to PI3K/Akt signaling pathway⁹⁴. A large body of previous research shows that the effects of EGFL7 on proliferation and apoptosis are reversed by miR-126 in ovarian cancer⁸⁸, renal cell carcinoma⁹⁵, liver cancer⁸⁷, oral squamous cell carcinoma⁹⁶, and non-small cell lung cancer⁸⁶.

Overexpression of miR-126 inhibits EGFL7 in HCC cell lines (HepG2, Bet-7402, and smmc-7721), thereby inhibiting cell proliferation and inducing apoptosis⁸⁹. By transplantation of HCC cell lines to establish nude liver cancer mice model, it was shown that enforced miR-126 expression decreased cancer weight 3 weeks after transplantation; once the expression of miR-126 was inhibited, less apoptosis and more proliferation were found⁸⁹. A subsequent study found that miR-126 targets EGFL7 as its main molecular mechanism for down-regulating the ERK signaling pathway⁸⁹. Cell proliferation correlation analysis showed that miR-126 targets EGFL7 to inhibit the proliferation of A549 cells *in vitro* and inhibit cancer growth *in vivo*⁸⁶. To sum up, EGFL7 is one of the important targets for miR-126 to promote cancer cell apoptosis, inhibit proliferation, and ultimately suppress cancer growth.

MiR-126/EGFL7 and Cancer Migration and Invasion

EGFL7 and the β 3 integrin, one of its receptors, signal axis is a key regulator in cancer metastasis⁹⁷. EGFL7 is highly expressed in cancer tissues and serum of HCC patients, and its expression level is closely related to HCC cancer vein invasion⁹⁸. Inhibition of EGFL7 expression can significantly inhibit the invasion and metastasis of HCC cells *in vivo* and *in vitro*, in part due to EGFR-mediated FAK phosphorylation and then changes in cancer cell migration capacity⁹⁸.

EGFL7 was upregulated in colorectal cancer cell lines, whereas transfection of EGFL7 siRNA significantly reduced the invasiveness of SW620 and LoVo colorectal cancer cell lines⁹⁴. When EGFL7 is expressed in cancer cells, it promotes cancer immune evasion by activation of cancer blood vessels⁹⁹. As mentioned above,

miR-126 down-regulation is an important reason for EGFL7 up-regulation, and miR-126 can target EGFL7 to delay the migration and invasion process of ovarian cancer⁸⁸ and renal cell carcinoma⁹⁵. Low expression of miR-126 up-regulates the expression of EGFL7, resulting in an invasive phenotype in ovarian cancer patients and poor prognosis; once restoring miR-126 expression can significantly inhibit the migration and invasion of cancer cells⁸⁸. Similar results were obtained from a renal cell carcinoma mice model⁹⁵. To sum up, miR-126 negative regulation of EGFL7 involves cancer metastasis and invasion in different cancers.

MiR-126/EGFL7 and Cancer Angiogenesis

The collective migration of endothelial cells is a critical step in angiogenic¹⁰⁰. EGFL7 is an essential factor for the formation of the vascular lumen, which plays an important role in maintaining the spatial structure and migration direction of endothelial cells during migration. In the absence of EGFL7, lumen formation is blocked, affecting the improvement of vascular function, while overexpression of EGFL7 is prone to result in vascular dysplasia⁹⁵. Studies have found that, in cancer-associated blood vessel wall, the basement membrane is missing or discontinuous, allowing cancerous tissue to pass through the wall of blood vessels and enter the bloodstream. These characteristics are related to the overexpression of EGFL7, which makes the endothelial cell motility too strong⁹⁵. EGFL7 has recently been identified as an important regulator of angiogenesis and progression in renal cell carcinoma. The overexpression of miR-126 inhibits cancer angiogenesis, at least in part by targeting EGFL7⁹⁵.

Studies found that miR-126 mimics and siRNA-EGFL7 significantly reduced the size, weight, and microvessel density of liver cancer xenografts in nude mice⁸⁷. The rat model of hepatoma was established by transplanting hepatoma cell lines (HepG2, Bet-7402, and smmc-7721), and it was found that overexpression of miR-126 in nude mice reduced cancer angiogenesis, while miR-126 inhibition promoted cancer angiogenesis⁸⁹. Further study has found that miR-126 inhibits angiogenesis in hepatocellular carcinoma mainly by decreasing EGFL7 and then the ERK signaling pathway⁸⁹. In a word, miR-126 can indirectly inhibit cancer growth and metastasis by targeting EGFL7, and thereby inhibiting angiogenesis in renal cell carcinoma and HCC.

MiR-126/CXCL12-CXCR4 axis and Cancer Progression

C-X-C motif chemokine ligand 12 (CXCL12), also known as stromal cell-derived factor-1 (SDF-1), is a homeostatic chemokine secreted mainly by fibroblasts, inflammatory, and endothelial cells^{101,102}. C-X-C Motif Chemokine Receptor 4 (CXCR4) is widely expressed in hematopoietic cells, with CXCL12 as its sole ligand¹⁰³⁻¹⁰⁵. Their interaction forms a coupling molecular pair, the CXCL12-CXCR4 axis, which is closely related to cell signal transduction and migration¹⁰⁶. Because of this, the CXCL12-CXCR4 axis may be involved in several aspects of cancer progression, including angiogenesis, metastasis, and survival. Interestingly, both CXCL12¹⁰⁷ and CXCR4¹⁰⁸ have been found to be the target genes of miR-126, suggesting that miR-126 inhibits cancer progression by targeting CXCL12 and/or CXCR4.

The miR-126/CXCL12-CXCR4 is responsible for inhibiting cancer cell metastasis. There was an increase in IL-6 in patients with inflammatory colorectal cancer, which was related to cancer size, stage, and metastasis¹⁰⁹. It was found that miR-126 affected macrophages function, and subsequently inhibited the proliferation and migration of colon cancer cells through CXCL12 / IL-6¹⁰⁷. Clinical studies showed a high level of CXCR4 and low level of miR-126 in the colorectal cancer tissues, which were related to distant metastasis, clinical TNM stage, and poor prognosis¹¹⁰. Similar results were shown *in vitro* and *in vivo*. For example, miR-126-3p reduced the metastasis of lung cancer *in vivo* and H460 lung cancer cells by targeting blocking CXCR4^{19,111}. In addition, miR-126 also inhibited the invasion of colon cancer cell lines HCT116 and SW480 by CXCR4/RhoA¹¹², and the proliferation of cancer cells in gastric cancer¹¹³, thyroid cancer¹¹⁴, and colon cancer¹¹² by targeting CXCR4. As a consequence, the CXCL12-CXCR4 has become an attractive target of miR-126 for cancer therapies.

MiR-126/LRP6 and Cancer Progression

Lipoprotein receptor-related protein 6 (LRP6) is a co-receptor for Wnt signaling¹¹⁵. When Wnt binds to LRP6 and is phosphorylated, the canonical Wnt- β -catenin signaling pathway is triggered, which ultimately affects cell proliferation, survival, and differentiation¹¹⁶. The microenvironment of cancers is rich in the Wnt ligand family, and abnormal signal transduction of Wnt signaling has been observed in cancer cells. LRP6, as an indispensable co-receptor of Wnt, is overexpressed

in colorectal cancer, liver cancer, and breast cancer of epithelial origin¹¹⁷. Silencing LRP6 reduces Wnt signaling and inhibits cancer cell proliferation in breast cancer cells *in vitro*; consistent with the literature mentioned above, the LRP6 antagonist Mesd significantly inhibited the growth of MMTV-Wnt1 cancers *in vivo*¹¹⁸. It suggests that LRP6 is a potential therapeutic target for breast cancer, especially for Wnt-activated breast cancer subtypes. It is found that LRP6 is significantly up-regulated in liver cancer tissues and cell lines (HepG2, SMMC-7721, BEL-7402), and is negatively correlated with the expression of miR-126-3p²⁸. And then LRP6 was identified as a negative target of miR-126-3p. MiR-126-3p significantly inhibits hepatoma cell migration and invasion of extracellular matrix gel, as well as preventing the formation of endothelial capillaries *in vitro*. Overexpression of miR-126-3p significantly reduced the cancer volume and microvessel density of hepatocellular carcinoma in nude mice²⁸. Similarly silencing LRP6 and repairing miR-126-3p had the same effect. These results suggesting that the inhibitory effect of miR-126 on hepatocellular carcinoma metastasis and angiogenesis is closely related to LRP6²⁸. To sum up, miR-126 was found to target PI3K, VEGF, VCAM-1, EGFL7, CXCL12-CXCR4 axis, and LRP6 in cancers, affecting the downstream signal and effector molecules, inhibit cancer apoptosis and proliferation, migration and invasion and angiogenesis, delay the process of cancer. However, a few contrary opinions suggest that further refinement and in-depth study is needed to fully understand the targeting effects of miR-126 and their impact on cancer.

miR-126 in the Pathogenesis of Atherosclerosis

Atherosclerosis is the underlying pathological process of ischemic cardiovascular and cerebrovascular diseases, which is characterized by the formation of atheroma or fibrous plaque in the intima of large and medium arteries¹¹⁹. Consistent with cancers, miR-126 is down-regulated in most patients with cardiovascular disease and atherosclerosis, and overexpression of miR-126 is beneficial for most cardiovascular disease models²⁰. Vascular endothelial cell apoptosis is an initial step of atherosclerosis, inflammatory response plays a crucial role in the whole progression, and intraplaque angiogenesis is involved in atherosclerotic plaque instability^{120,121}. Interestingly, as

in cancer, miR-126 regulates the above pathological processes in atherosclerosis by targeting PI3K, VEGF, VCAM-1, EGFL7, and LRP6 genes.

miR-126/PIK3R2 and Endothelial Cell Apoptosis

Normal endothelial cells line the luminal surface of blood vessels, which effectively prevent atherosclerosis and its complications. On the contrary, apoptosis of endothelial cells facilitates atherogenesis and its thrombotic complications through multi-links: 1) Damage endothelial function, lead to immune and inflammatory disorders¹²²; 2) Destroy the endothelial barrier function and then promote lipids deposition in the arteries intima¹²³; 3) Exposure the subendothelial matrix proteins to the blood and result in thrombosis¹²⁴.

Oxidized low-density lipoprotein (ox-LDL), a strong pro-atherogenic factor, induces human umbilical vein endothelial cells (HUVECs) apoptosis and down-regulation of miR-126, which can be significantly reversed by miR-126 mimics¹²⁵. Studies have found that downregulation of miR-126 inhibits the PI3K/Akt signaling, while overexpression of miR-126 significantly increased PI3K/Akt signaling as well as its downstream proteins, such as Bcl-2, Bad, and cleavage of caspase-9¹²⁶. According to above results, miR-126 can stimulate Akt phosphorylation through targeting PIK3R2, and then inhibit vascular endothelial cells apoptosis in atherosclerosis-related models. Similarly, apoptosis of UVECs-CRL-1730 Cell Line induced by H₂O₂ is related to miR-126 down-regulation and reduction of PIK3R2 (p85 β) inhibition¹²⁷. MiR-126 is involved in HUVECs apoptosis via regulating the PI3K signal. It has been reported that P-PI3K, p-Akt, and p-mTOR are upregulated, while miR-126, caspase-3 activity, and apoptotic rate are downregulated simultaneously. MiR-126 mimics reverse the above state; however, the 740Y-P (a PI3K activator) inverts miR-126 mimic's effect by inhibiting PI3K/Akt/mTOR signal, suggesting that miR-126 exerts an anti-apoptotic effect by targeting PI3K/Akt signaling¹²⁵.

Despite the fact that most studies have recognized that miR-126 inhibits endothelial cell apoptosis, opinions on miR-126 regulating PI3K are mixed. There are even more conflicting opinions: endothelial cells apoptosis induced by ox-LDL is accompanied by overexpression of miR-126, miR-126 inhibitor transfecting endothelial cells inhibits ox-LDL-induced apoptosis, and the PI3K/Akt signaling pathway is continuously acti-

vated, suggesting that miR-126 inhibits the PI3K/Akt signaling pathway and promote endothelial cell apoptosis¹²⁸. In conclusion, it remains unclear how miR-126 regulates endothelial cell apoptosis by targeting PI3K, and further research is needed.

MiR-126/VCAM-1 and PIK3R2 and Atherosclerosis-related Inflammation

Both basic research and clinical evidence show that chronic vascular inflammation is involved in the development of atherosclerosis¹²⁹. Atherosclerosis is largely influenced by inflammation, from the formation of the atherosclerotic plaques to their eventual rupture^{121,130}. Adhesion of monocytes to vascular endothelium is the key step in the early stages of atherogenesis, followed by the formation of foam cells and early plaque in the arterial intima. VCAM-1 is located on the surface of activated endothelial cells and is considered to be a key mediator of atherosclerosis, by specifically mediating the adhesion of monocytes and T lymphocytes to endothelial cells and promote the migration of leukocytes to the intima of blood vessels¹³¹. The combination of VCAM-1 with integrin triggers the signal cascade in endothelial cells, resulting in an increase of reactive oxygen species (ROS), inducing actin reorganization and destroying endothelial tight junction, which is the main mechanism of leukocyte migration from the vascular lumen to intima¹³². Studies have found that VCAM-1 is overexpressed only at the aortic plaque formation area¹³³, and targeted disruption of VCAM-1 inhibits the formation of early plaques in *Ldlr*^{-/-} mice¹³⁴.

Overexpression of miR-126 significantly alleviated the progression of atherosclerosis and VCAM-1 expression, whereas inhibition of miR-126 is reversed¹³⁵. Similarly, miR-126 expression was significantly downregulated, and VCAM-1 expression was significantly up-regulated in the *ApoE*^{-/-} mouse model of atherosclerosis¹³⁶. *In vivo*, atorvastatin treatment increased miR-126 levels, lowered VCAM-1 levels, and alleviated atherosclerotic lesions¹³⁶. Cigarette smoke (CS) upregulated the expression of VCAM-1 and then stimulated vascular inflammation in *apoE*^{-/-} mice, while downregulating the miR-126¹³⁷. These results showed that miR-126 can inhibit the adhesion of monocytes to endothelial cells by inhibiting VCAM-1 expression, and thereby alleviating arterial walls inflammation. Nevertheless, the opposite has also been found in some studies. For example, patients with coronary heart disease exhibited higher levels of both VCAM-1 and miR-

126 (6.72 times) than healthy individuals¹²⁰. It suggests that miR-126 and VCAM-1 expression are affected by multiple factors, which need to be refined. Overall, miR-126 targeting VCAM-1 is a new idea for treating atherosclerosis despite differing opinions.

Additionally, miR-126 inhibits inflammation by targeting PI3K in atherosclerosis. *In vitro* studies, paeonol promotes miR-126 expression in the rat thoracic aortic endothelial cells stimulated by ox-LDL, blocks the PI3K/Akt/NF- κ B pathway, and inhibits the adhesion of monocytes to vascular endothelial cells¹³⁸. In the absence of more support data, PI3K regulation of inflammation by miR-126 is evidently unstudied in atherosclerosis.

MiR-126/VEGF, EGFL7, PI3K, and Intra-Plaque Angiogenesis

Atherosclerotic plaques become increasingly vulnerable and rupture, resulting in luminal thrombosis, increasing the risk of cardiac and cerebrovascular events, such as stroke, dementia, and myocardial infarction. Intra-plaque angiogenesis, driven by local hypoxia¹³⁹, promotes atherosclerotic plaque development and plaque destabilization^{140,141}. Physiologically, there is no neovascularization in the intima of arteries. With the accumulation of foam cells and proliferation of smooth muscle cells, plaques are gradually increasing, and hypoxia in the plaques is aggravating, thereby exacerbating angiogenesis in the plaques by stimulating medial nourishing vessels to enter the intima. VEGF and its receptor (VEGFR) have been identified as the major pathway involved in intra-plaques angiogenesis¹⁴². There was evidence that endothelial cells of intra-plaque neovascularization express higher concentrations of VEGF and VEGFR than those in the lumen of blood vessels¹⁴³. Studies demonstrated that ox-LDL induced angiogenesis of HUVECs *in vitro*, in which VEGF-A induced endothelial cell proliferation and migration¹⁴⁴. Furthermore, Li¹⁴⁵ found that overexpression of VEGF promotes endothelial cell migration, proliferation, and angiogenesis by activating the VEGF/VEGFR2 pathway. In addition, VEGF-A can induce MMP-2, MMP-9, and urokinase-type plasminogen by activating NF- κ B. MMP-2/9 can degrade the basement membrane and extracellular matrix to allow migration of new endothelial cells, and then format capillary sprouts¹⁴⁶. All these suggest that VEGF/VEGFR2 and its induced NF- κ B signal participate in angiogenesis in atherosclerosis, stimulate plaque progression, and cause plaque unstable.

In general, the expression of miR-126 is high, and yet VEGF is low in endothelial cells, and there is a negative correlation between them. The over-expression of miR-126 down-regulates VEGF-A expression in cells, especially in non-endothelial cells, thus indirectly suppressing quiescent endothelial cell activation. As is well known, the enlargement of plaque volume will lead to local hypoxia of plaque and promote intra-plaque angiogenesis. Atherosclerotic patients showed an increase in VEGF-A with the increase of intima-media thickness (IMT) and plaque area, but a decrease in miR-126 expression, and indicating a significant negative correlation between miR-126 and VEGF-A¹⁴⁷. To sum up, miR-126 down-regulation and VEGF-A up-regulation is the possible mechanism of intra-plaque angiogenesis. However, the direct evidence that miR-126 targeting VEGF inhibits intra-plaque angiogenesis is relatively insufficient at present and further research is needed.

EGFL7 is a secretory protein produced by vascular endothelial cells that participates in angiogenesis. There was an upregulation of EGFL7 in 11 of the 14 human atherosclerotic plaque samples compared with matched control samples (plaque adjacent areas), suggesting that elevated EGFL7 may lead to atherosclerosis¹⁴⁸. In line with this, the EGFL7 was detected in both endothelial cells and vascular smooth muscle cells of atherosclerotic plaques, with the highest expression in proliferative endothelial cells¹⁴⁹. Further research shows that transfecting recombinant plasmid p_{gfp-N1}/miR-126 into endothelial cell line ECV-304 caused a reduction of 67% in EGFL7 protein expression compared to empty vector transfection group (only 6.5%) after 48h transfection, suggesting that miR-126 can reduce EGFL7 protein expression in ECV-304 cells¹⁵⁰. In addition, miR-126 inhibits angiogenesis in HUVECs, as well as down-regulate EGFL7 mRNA and protein expression significantly¹⁵¹. In summary, miR-126 inhibits intra-plaque angiogenesis, at least in part, by targeting EGFL7.

Recently, PI3K has been found to be closely related to atherosclerosis-related angiogenesis. As a risk factor of atherosclerosis, low concentration ox-LDL can promote angiogenesis of human coronary artery endothelial cells (HCAECs) *in vitro* and activate the synthesis of nitric oxide via the PI3K/Akt/eNOS pathway¹⁵². It is known that eNOS can promote angiogenesis through Akt/PKA¹⁵³. Therefore, PI3K and downstream pathway Akt/eNOS are important promoting factors of angiogenesis¹⁵³. In addition, ox-LDL promotes the

process of angiogenesis by relying on the nucleocytoplasmic shuttle of Id1, which was controlled by the PI3K pathway, and the inhibition of PI3K blocks the angiogenesis induced by ox-LDL¹⁵⁴. It follows that PI3K participates in the angiogenesis of atherosclerosis through various mechanisms. Although it has been found that PI3K is regulated by miR-126 to participate in the angiogenesis of endothelial cells^{155,156}, the direct evidence of miR-126 regulates PI3K on the angiogenesis of atherosclerotic models is not established.

MiR-126/LRP6 and Vascular Smooth Muscle Proliferation

Low-density lipoprotein receptor-related protein 6 (LRP6), as a member of the low-density lipoprotein receptor family, is a co-receptor of the Wnt signaling pathway and plays an important role in atherosclerosis¹⁵⁷. Excessive PDGF signaling detected in LRP-R611C mice is associated with activation of the Wnt signaling pathway and upregulation of Sp1 (a transcription factor known to target PDGF and PDGFR- β gene expression)¹⁵⁷. Keramati et al¹⁵⁸ reported that human atherosclerotic coronary arteries overexpress and colocalize LRP6 and PDGFR- β . Wild-type LRP6 forms a complex with PDGFR- β , triggering its lysosomal degradation. This effect reduces the proliferation of vascular smooth muscle cells, which has a protective effect against atherosclerosis¹⁵⁸. More studies have found that LRP6R611C mutation significantly activates PDGF signaling and increases smooth muscle proliferation, which promotes the occurrence and development of atherosclerosis¹⁵⁹. The above studies suggest that LRP6 inhibits the proliferation of vascular smooth muscle induced by PDGFR- β , delaying atherosclerosis progression. Previous studies have found that miR-126 targets LRP6, suggesting that miR-126 is involved in atherosclerosis by targeting LRP6²⁸. After miR-126 was transferred from endothelial microparticles into VSMCs, the proliferation of VSMCs and the formation of new intima, as well as LRP6 expression were inhibited¹⁶⁰, which is different from the previous conclusion. Consequently, miR-126 targeting LRP6 participates in the process of atherosclerosis by mediating the proliferation of smooth muscle cells, which needs further study.

MiR-126/CXCL12-CXCR4 Axis and Vascular Insult, Neovascularization

CXCL12 plays important, complex, and even contradictory role in atherosclerosis, from proatherogenic, proinflammatory, and prothrombotic to

atheroprotective, plaque stabilizer, and dyslipidemia rectifier, mainly through its classical receptor CXCR4^{161,162}. CXCR4 is widely expressed in various cell types that play a role in CVDs, such as endothelial progenitor cells (EPCs), endothelial cells, macrophages, platelets, and smooth muscle cells¹⁶². Similarly, high expression of CXCR4 and CXCL12 mRNA was also found in patients with carotid plaques¹⁶³. Furthermore, they are also implicated in intraplaque neovascularization and thrombus formation in the advanced atherosclerotic plaques¹⁶⁴⁻¹⁶⁷. CXCL12 levels are also associated with hyperlipidemia and inflammation. For example, CXCL12 overexpression in ApoE^{-/-} mice increases macrophage infiltration, inhibits reverse cholesterol transport, decreases plasma HDL-C levels, and then enlarges atherosclerotic lesions¹⁶⁸. Through CXCL12/CXCR4 signaling, platelets involved in atherothrombosis¹⁶⁹ that is related to increased dense granule secretion and thromboxane A2 production¹⁷⁰. In addition, CXCL12-CXCR4 axis controls the proliferation and migration of smooth muscle cells¹⁷¹, promoting plaque formation, stabilization, and re-stenosis¹⁷². As mentioned above, the role of the CXCL12/CXCR4 axis in atherosclerosis is still controversial.

CXCL12 has been identified as a direct target of miR-126 by miRNA prediction in EPCs and miR-126 improves EPCs migration by targeting CXCL12^{173,174}. Therefore, miR-126 may affect atherosclerosis by targeting CXCL12 and/or CXCR4. There are only a few studies suggesting that miR-126 negatively regulates CXCL12 to inhibit EPCs-mediated angiogenesis¹⁷³, but others have yielded contradictory results. For example, CXCL12 was found to promote angiogenesis in the aorta of SD rats with the overexpression of miR-126-3p, and its effect was eliminated by inhibiting of miR-126-3p¹⁷⁵. This paradoxical phenomenon may be related to the fact that miR-126 also targets SPRED-1 and then affects the function of CXCL12¹⁷⁵. So, research is needed to determine how miR-126 affects atherosclerosis by directly targeting the CXCL12-CXCR4 axis.

Association Between the Effects of MiR-126 on Atherosclerosis and Cancer

MiR-126 and Proliferation and Apoptosis Both of Cancer and Atherosclerosis

Cell proliferation and apoptosis are two opposite states of cells. In cancer lesions, cancer

growth results from unchecked cell proliferation and inhibition of apoptosis; in atherosclerotic lesions, the apoptosis of endothelial cells coated on the lumen surface of arterial intima is the early initial event of atherosclerosis, while the proliferation of smooth muscle cells promotes the progression of atherosclerotic plaques. As mentioned above, miR-126 mainly targets inhibiting PI3K, VEGF, and EGFL7, participating in cell proliferation and apoptosis, and taking part in the process of cancer and atherosclerosis.

MiR-126 inhibits cancer cells proliferation and promote apoptosis by targeting PI3K/Akt signaling pathway, and participate in the progression of endometrial cancer¹⁷, cervical cancer³⁹, non-small cell lung cancer²⁶, esophageal squamous cell carcinoma¹⁸ and prostate cancer²⁵. At the same time, in atherosclerosis-related models, miR-126 inhibits PI3K/Akt (p85-β) regulating PI3K/Akt or PI3K/Akt/mTOR signaling pathway, further reduce the apoptosis rate of endothelial cells^{125,126}, and restrain the proliferation of VSMCs induced by ox-LDL, ultimately inhibiting atherosclerosis¹⁷⁶. As mentioned above, PTEN acts as cancer-suppressor and an atherosclerosis-promoter. In non-small cell lung cancer A549 cells, miR-126 was found to inhibit PI3K/Akt signaling pathway by targeting PIK3R2, increasing the expression of PTEN and inhibiting the proliferation of cancer cells²⁶. Interestingly, PTEN decreases VSMCs proliferation and migration, accelerating atherosclerosis, suggesting that PTEN is a potential target for miR-126 inhibiting smooth muscle proliferation¹⁷⁷.

As well as PI3K, VEGF and EGFL7 are also miR-126 regulatory signals, inhibit cancer proliferation, promote apoptosis, and participate in the process of cancer. Overexpression of miR-126-5p can inactivate VEGF-A/VEGFR2/ERK signaling pathway and promote apoptosis of non-small cell lung cancer H1299 cell line⁵⁰. In ovarian cancer⁸⁸, renal cell carcinoma⁹⁵, liver cancer⁸⁷, oral squamous cell carcinoma⁹⁶, and non-small cell lung cancer⁸⁶, miR-126 can also inhibit cell proliferation and promote apoptosis by targeting EGFL7 signaling. Further studies have shown that the downstream signal of miR-126 targeting EGFL7 is ERK in hepatoma cell lines (HepG2, Bet-7402, and smmc-7721)⁸⁹. In atherosclerosis-related cell models, ERK was found to mediate curcumin to inhibit HIF-1α-induced apoptosis of macrophages. It suggests that miR-126 can target VEGF or EGFL7 to affect ERK-mediated apoptosis of cancer cells and ath-

therosclerosis-related cells¹⁷⁸. To sum up, despite the existence of different opinions, miR-126 regulates proliferation and apoptosis of cancer cells and atherosclerosis-related cells by targeting PI3K, VEGF, and EGFL7 at the same time.

MiR-126 Influences Angiogenesis in Both Cancers and Atherosclerosis

Angiogenesis is not only a necessary condition for cancer progression but also a factor in atherosclerotic plaque destabilization and rupture. Therefore, inhibition of angiogenesis is also an important point in the control of cancer and atherosclerosis. According to the above, miR-126 participates in angiogenesis in cancers and intra-plaques by targeting VEGF, PI3K, and EGFL7.

VEGF binding to VEGFRs on vascular endothelial cells promotes cancer angiogenesis. MiR-126 inhibits cancer angiogenesis in oral squamous cell carcinoma³², liver cancer⁵², gastric cancer⁵⁵, and breast cancer⁵⁷ by targeting VEGF expression. Down-regulation of miR-126 and up-regulation of VEGF are related to intra-plaque angiogenesis¹⁴⁷, suggesting that miR-126 may also reduce angiogenesis, and thereby stabilizing plaques by inhibiting VEGF expression. In gastric cancer cell lines, such as SGC-7901, MKN-28, and MKN-45, miR-126 could regulate the activity of Akt, mTOR, and ERK1/2 by targeting VEGF-A, and ultimately inhibit the growth and angiogenesis of gastric cancer⁵⁵. Similarly, MMPs were induced by VEGF-A via activating NF- κ B¹⁴⁶, which degrade the basement membrane and extracellular matrix to enable new endothelial cells to migrate, resulting in capillary sprouts in atherosclerosis¹⁴⁶. MiR-126 negatively regulated VEGF/VEGFR2 and then NF- κ B signaling to participate in angiogenesis of atherosclerosis and thereby causes plaque instability¹⁷⁹. These suggests that miR-126 can inhibit cancer and atherosclerosis related angiogenesis by down-regulating VEGF.

In renal cancer⁹⁵ and liver cancer⁸⁷, miR-126 has been shown to affect cancer angiogenesis by regulating EGFL7; even more than that, miR-126 has also been found to inhibit EGFL7 in atherosclerosis, thereby inhibiting intra-plaques angiogenesis^{150,151}. In the study of liver cancer, it was found that miR-126/EGFL7 partially inhibited angiogenesis by down-regulating the ERK signaling pathway⁸⁹, but there has been no further evidence of atherosclerosis.

MiR-126 Influences Inflammatory Responses in Cancers and Atherosclerosis

Inflammation is involved in the occurrence and development of both cancer and atherosclerosis, and miR-126 targets VCAM-1 to do so. The downregulation of VCAM-1 by MiR-126 reduces monocyte adhesion to endothelial cells caused by cigarette smoke, and alleviate the arterial wall inflammation¹³⁷. Macrophages modulate VCAM-1 and integrin α 4 β 1 and involve in the stimulation of breast cancer cell survival by electronic smoke⁸³. It appears that the mechanisms of smoke-induced cancers and atherosclerosis have something in common, and miR-126 can prevent and treat them by inhibiting these mechanisms. MiR-126 can significantly inhibit the adhesion of monocytes to ox-LDL-activated rat thoracic aortic endothelial cells by down-regulating VCAM-1, thereby delaying the process of atherosclerosis¹³⁶. Similarly, IL-1 β -induced VCAM-1 may also modulate adhesion between glioblastomas cells and monocytes⁸⁴. Accordingly, VCAM-1 is involved in monocyte adhesion and is inhibited by miR-126 in both cancer and atherosclerosis. In addition, cancer-associated macrophages contribute to the progression and Warburg effect of pancreatic ductal adenocarcinoma via the CCL18/NF- κ B/VCAM-1 pathway⁸². According to these findings, miR-126 targeting VCAM-1 is involved in the inhibition of cancer and atherosclerotic disease development through multiple mechanisms.

MiR-126 participates in the pathological and pathophysiological processes of cancer and atherosclerosis by inhibiting PIK3R2, VEGF-A, VCAM-1, LRP6, and EGFL7. 1) In terms of cancer, miR-126 inhibits PIK3R2/PI3K/Akt pathway and regulates its downstream targets MRP1, mTOR, PTEN, ANLN, Snail, and caspase-3, and thereby affects drug resistance, proliferation, migration and invasion, apoptosis of cancer cells, delaying cancer progression. In terms of atherosclerosis, miR-126 inhibits PIK3R2/PI3K/Akt pathway and regulates downstream targets Caspase-3, mTOR and NF- κ B, eNOS and Id1, and then affects endothelial cell apoptosis, inflammation and angiogenesis, slowing down the progress of atherosclerosis. 2) On one hand, miR-126 inhibits VEGF-A/VEGFR pathway and regulates its downstream target ERK, and then affects cancer migration, invasion, and apoptosis; on the other hand, MiR-126 inhibits VEGF-A/VEGFR pathway and regulates its downstream

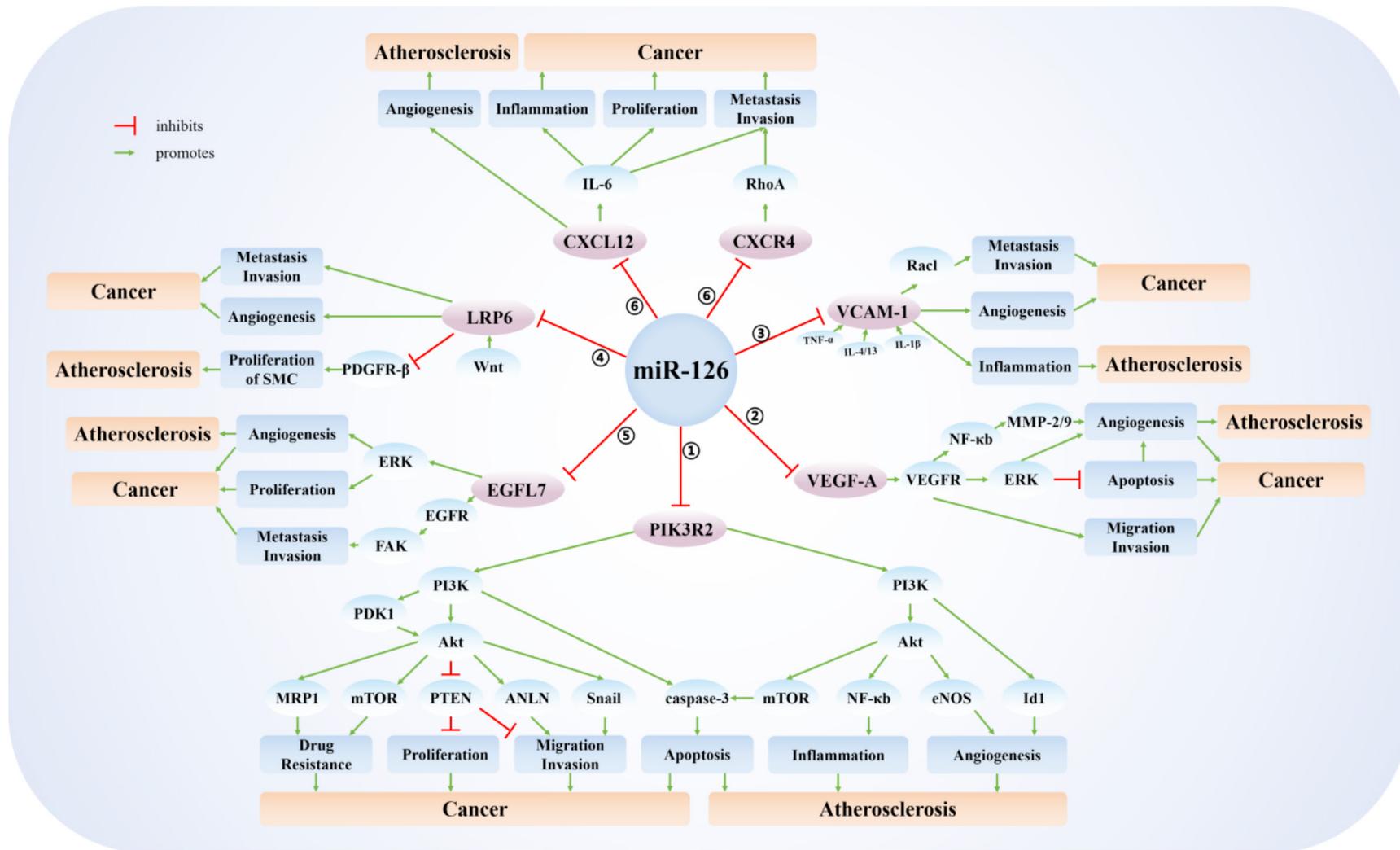


Figure 1. MiR-126 targets PI3K, VEGF, VCAM-1, EGFL7, CXCL12-CXCR4 axis, and LRP6 to involve in both cancer and atherosclerosis.

Table I. The role of miR-126 in cancer and atherosclerosis through common target genes.

Targets of miR-126	Effect on targets	Signaling pathways/effectors/molecule	Effect on pathways/molecule	Disease	Expression of miR-126 in disease	Research object	Role of miR-126	Reference	
PI3K	PIK3R2	-	Caspase-3	Promote	Endometrial carcinoma	Downregulated	RL95 and HEC1A cell	Inhibit proliferation and induce apoptosis	17
	PIK3R2	Inhibit	PTEN	Promote	Non-small cell lung cancer	Downregulated	A549 cell	Inhibit proliferation, metastasis and invasion	26
	-	-	PI3K/PDK1/Akt	-	Cervical cancer	Downregulated	HeLa cell	Promote apoptosis, inhibit proliferation and metastasis	39
	PIK3R2	Inhibit	PI3K/Akt	Inhibit	Esophageal squamous cell carcinoma	Downregulated	EC109 cell	Inhibit proliferation and metastasis	18
	PIK3R2	Inhibit	PI3K/Akt	Inhibit	Prostate cancer	Downregulated	DU145, PC-3 and 293T cell	Inhibit proliferation	25
	-	-	PI3K/Akt/Snail	Inhibit	Lung cancer	Downregulated	SPC-A1 and LLC cell	Inhibit metastasis	40
	PIK3R2	Inhibit	PI3K/Akt/MRP1	Inhibit	Gastric cancer	Downregulated	SGC-7901, BGC-823, SGC-7901/DDP and BGC-823/DDP cell	Reduce drug resistance	46
	PIK3R2	Inhibit	PIK3R2/PI3K/Akt/mTOR	Inhibit	Breast cancer	Downregulated	SKBR3/TR cell	Reduce drug resistance	36
	PIK3R2	Inhibit	Akt	Promote		Downregulated	CRL-1730 cell	Inhibit apoptosis	127
	-	-	PI3K/Akt	Promote		Downregulated	Human umbilical vein endothelial cells	Inhibit apoptosis	126
	-	-	PI3K/Akt/mTOR	Inhibit	Atherosclerosis	Downregulated	Human umbilical vein endothelial cells	Inhibit apoptosis	125
	-	-	PI3K/Akt	Inhibit		Upregulated	Human umbilical vein endothelial cells	Promote apoptosis	128
	-	-	PI3K/Akt/NF-κB	Inhibit		Downregulated	Endothelial cells of rat thoracic aorta	Inhibit inflammation	138
VEGF	VEGFA	Inhibit	VEGF-A/VEGFR-2/ERK	Inhibit	Non-small cell lung cancer	Downregulated	NCI-H1299 cell	Promote apoptosis, inhibit metastasis and invasion	50
	VEGF	Inhibit	-	-	Ovarian cancer	Downregulated	SKOV3 cell	Inhibit proliferation, metastasis and invasion	56
	VEGF	Inhibit	-	-	Breast cancer	Downregulated	Breast cancer mice	Inhibit metastasis	58
	VEGF-A	Inhibit	-	-	Ovarian cancer	Downregulated	SKOV3 and ES2 cell	Inhibit invasion and metastasis	53
	VEGF-A	Inhibit	Akt, mTOR and ERK1/2	-	Gastric cancer	Downregulated	SGC-7901, MKN-28 and MKN-45 cell	Inhibit angiogenesis	55
	VEGF-A	Inhibit	-	-	Liver cancer	Downregulated	HepG2 cell	Inhibit angiogenesis	52
	VEGF-A	Inhibit	-	-	Breast cancer	Downregulated	MCF cell	Promote apoptosis, inhibit proliferation and angiogenesis	57
VCAM-1	VEGF-A	Inhibit	-	-	Atherosclerosis	Downregulated	Atherosclerotic patients	Reduce intima-media thickness and plaque area	147
	VCAM-1	Inhibit	-	-	Prostate cancer	Downregulated	PC3 and DU145 cell	Inhibit metastasis	66
	VCAM-1	Inhibit	-	-	Chronic myelogenous leukemia	Downregulated	LAMA84 cell	Inhibit metastasis and invasion	68
	VCAM-1	Inhibit	-	-	Chondrosarcoma	Downregulated	JJ012 cell	Inhibit metastasis and invasion	67
	VCAM-1	Inhibit	-	-	Atherosclerosis	Downregulated	ApoE ^{-/-} mice; rat thoracic aorta endothelial cell	Inhibit inflammation	136
EGFL7	VCAM-1	Inhibit	-	-		Downregulated	ApoE ^{-/-} mice	Inhibit inflammation	137
	EGFL7	Inhibit	ERK	Inhibit	Liver cancer	Downregulated	Nude mouse liver cancer model; hepG2, Bet-7402 and smmc-7721 cell	Promote apoptosis, inhibit proliferation and angiogenesis	8
	EGFL7	Inhibit	-	-	Non-small cell lung cancer	Downregulated	A549 cell	Inhibit proliferation and promote apoptosis	86
	EGFL7	Inhibit	-	-	Ovarian cancer	Downregulated	Ovarian cancer patients	Inhibit metastasis and invasion	88
	EGFL7	Inhibit	-	-	Renal cell carcinoma	Downregulated	Renal carcinoma mouse model	Inhibit metastasis and angiogenesis	95
	EGFL7	Inhibit	-	-	Liver cancer	Downregulated	Nude mouse liver cancer model	Inhibit angiogenesis	87
	EGFL7	Inhibit	-	-	Atherosclerosis	Downregulated	ECV-304 cell	Inhibit angiogenesis	150
LRP6	EGFL7	Inhibit	-	-		Downregulated	Human umbilical vein endothelial cells	Inhibit angiogenesis	151
	LRP6	Inhibit	-	-	Liver cancer	Downregulated	Clinical liver cancer tissue and normal liver tissue; L02 cell, HepG2 cell, SMMC-7721 cell, BEL-7402 cell	Inhibit migration, invasion and angiogenesis	28
	LRP6	Inhibit	-	-	Atherosclerosis	-	Smooth muscle cells	Inhibit proliferation	160

target NF- κ B and MMP-2/9, and then affects angiogenesis, participating in the progress of atherosclerosis. 3) MiR-126 inhibits VCAM-1 and regulates downstream target Racl, and then affects angiogenesis, cancer cell migration and invasion, participating in cancer progression. MiR-126 inhibits VCAM-1 to affect endothelial inflammation, participating in the progression of atherosclerosis. 4) MiR-126 inhibits LRP6 and then affects cancer cell migration and invasion, angiogenesis, participating in cancer progression. MiR-126 inhibits LRP6- β and regulates PDGFR, and then affects the proliferation of vascular smooth muscle cells, involving in the progression of atherosclerosis. 5) MiR-126 inhibits EGFL7 and regulates downstream targets EGFR, FAK, and ERK, and then affects cancer cell migration and invasion, proliferation and angiogenesis, participating in cancer progression. MiR-126 inhibits EGFL7/ERK, and then affects angiogenesis, participating in the progression of atherosclerosis. 6) MiR-126 inhibits CXCL12/CXCR4 and regulates downstream targets IL-6 and RhoA, and then affects cancer cell inflammation, proliferation, migration and invasion, participating in cancer progression. MiR-126 inhibits CXCL12, and then affects angiogenesis, participating in the progression of atherosclerosis. α 4 β 1, α 4 β 1 integrin.

Clinical Application of MiR-126 in Cancer and Atherosclerosis

MiR-126 has aroused great interest as a new biomarker for the diagnosis and treatment of cancers and atherosclerosis and related cardio-cerebrovascular diseases. Several studies have confirmed the diagnostic and prognostic value of miR-126 in lung cancer¹⁸⁰⁻¹⁸⁴. In addition, miR-126 is also identified as a specific diagnostic marker and new therapeutic target for ovarian cancer¹⁸⁵, colorectal cancer¹⁸⁶, gastric cancer¹⁸⁷, hepatocellular carcinoma¹⁸⁸, esophageal squamous cell carcinoma¹⁸⁹, prostate cancer¹⁹⁰, glioma¹⁹¹ and breast cancer^{27,192,193}. Similarly, recent studies have started to unveil the potentially important function of miR-126 in atherosclerosis and related cardio-cerebrovascular diseases^{25,194}. In addition, miR-126 may be considered as a diagnosis biomarker by using blood-based non-invasive methods, in atherosclerosis¹⁹⁵, acute myocardial infarction¹⁹⁶, ischemic stroke¹⁹⁷ and diabetic vasculopathy¹⁹⁸.

Conclusions

MiR-126 is a small RNA highly expressed in endothelial cells, which is closely related to cancer and atherosclerosis. Thus, miR-126 is regarded as a potentially important target for the intervention of cancer and atherosclerosis. But how miR-126 can kill two birds with one stone: benefit both malignant cancers and atherosclerosis remains a mystery. At present, the main point of view is that the over-expression of miR-126 is beneficial to atherosclerosis and cancerogenesis, which is due to different mechanisms of miR-126 function. The underlying mechanisms of miR-126 are involved in multiple effects and multiple targets (Figure 1 and Table I): 1) miR-126 targets PIK3R2, VEGF, and EGFL7 genes to promote cancer cell apoptosis and inhibit apoptosis of endothelial cell and proliferation of VSMCs related to atherosclerosis. 2) miR-126 targets PIK3R2, VEGF, VCAM-1, and EGFL7, inhibits cancer angiogenesis, delays cancer progression, inhibits intra-plaques angiogenesis, stabilizes vulnerable plaques, and reduces the occurrence of complications such as thrombosis. 3) miR-126 inhibits the inflammatory response of cancers and atherosclerosis by targeting VCAM-1 and PIK3R2. In conclusion, miR-126 affects cancer and atherosclerosis via inhibiting cell proliferation, promoting apoptosis, inhibiting angiogenesis, and inhibiting inflammatory response, which provides a new idea for clinical treatment of cancer and atherosclerosis. However, there are still contradictory results related to the characteristics of environmental dependence and target diversity of miR-126. Further intensive studies are needed to explore the molecular and biological functions of miR-126 in cancer and atherosclerosis.

Conflict of Interest

The authors declare that the paper was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Authors' Contributions

Qiuyue Yang and Qun Yu drafted the manuscript. Xijuan Jiang and Jiali Gan designed and supervise manuscript. Wenyun Zeng verified the contents and revised the manuscript. Miao Zeng, Xiaolu Zhang, Yilin Zhang, Lin Guo critically revised the manuscript. All authors reviewed and approved the final manuscript.

Funding

2020 Annual Graduate Students Innovation Fund, Grant/Award Number: ZXYCXLX202007, ZXYCXLX202008; National Natural Science Foundation of China, Grant/Award Numbers: 81873130, 82074211; Graduate Research

Innovation Project of Tianjin University of Traditional Chinese Medicine, Grant/Award Number: YJSKC-20201017.

Acknowledgments

This study was supported by the National Natural Science Foundation of China (Nos. 82074211, 81873130), 2020 Annual Graduate Students Innovation Fund (School of Integrative Medicine, Tianjin University of Traditional Chinese Medicine, Tianjin, China; No. ZXYCXLX202007, ZXYCXLX202008) and Graduate Research Innovation Project of Tianjin University of Traditional Chinese Medicine (No. YJSKC-20201017).

References

- 1) Santucci C, Carioli G, Bertuccio P, Malvezzi M, Pastorino U, Boffetta P, Negri E, Bosetti C, La Vecchia C. Progress in cancer mortality, incidence, and survival: a global overview. *Eur J Cancer Prev* 2020; 29: 367-381.
- 2) Torres N, Guevara-Cruz M, Velazquez-Villegas LA, Tovar AR. Nutrition and Atherosclerosis. *Arch Med Res* 2015; 46: 408-426.
- 3) Klement H, St CB, Milsom C, May L, Guo Q, Yu JL, Klement P, Rak J. Atherosclerosis and vascular aging as modifiers of tumor progression, angiogenesis, and responsiveness to therapy. *Am J Pathol* 2007; 171: 1342-1351.
- 4) Tapia-Vieyra JV, Delgado-Coello B, Mas-Oliva J. Atherosclerosis and Cancer; A Resemblance with Far-reaching Implications. *Arch Med Res* 2017; 48: 12-26.
- 5) Bartel DP. MicroRNAs: genomics, biogenesis, mechanism, and function. *Cell* 2004; 116: 281-297.
- 6) Staszek T, Zapala B, Polus A, Sadakierska-Chudy A, Kiec-Wilk B, Stepień E, Wybranska I, Chojnacka M, Dembinska-Kiec A. Role of microRNAs in endothelial cell pathophysiology. *Pol Arch Med Wewn* 2011; 121: 361-366.
- 7) Wang X, Meng K, Wang H, Wang Y, Zhao Y, Kang J, Zhang Y, Quan F. Identification of small extracellular vesicle subtypes in follicular fluid: Insights into the function and miRNA profiles. *J Cell Physiol* 2021; 236: 5633-5645.
- 8) Ginckels P, Holvoet P. Oxidative Stress and Inflammation in Cardiovascular Diseases and Cancer: Role of Non-coding RNAs. *Yale J Biol Med* 2022; 95: 129-152.
- 9) Uzuner E, Ulu GT, Gürler SB, Baran Y. The Role of MiRNA in Cancer: Pathogenesis, Diagnosis, and Treatment. *Methods Mol Biol* 2022; 2257: 375-422.
- 10) Lou X, Wang D, Gu Z, Li T, Ren L. Mechanism of microRNA regulating the progress of atherosclerosis in apoE-deficient mice. *Bioengineered* 2021; 12: 10994-11006.
- 11) Olivieri F, Spazzafumo L, Bonafe M, Recchioni R, Prattichizzo F, Marcheselli F, Micolucci L, Mensa E, Giuliani A, Santini G, Gobbi M, Lazzarini R, Boemi M, Testa R, Antonicelli R, Procopio AD, Bonfigli AR. MiR-21-5p and miR-126a-3p levels in plasma and circulating angiogenic cells: relationship with type 2 diabetes complications. *Oncotarget* 2015; 6: 35372-35382.
- 12) Olivieri F, Procopio AD, Montgomery RR. Effect of aging on microRNAs and regulation of pathogen recognition receptors. *Curr Opin Immunol* 2014; 29: 29-37.
- 13) Lin Q, Hou S, Dai Y, Jiang N, Lin Y. LncRNA HO-TAIR targets miR-126-5p to promote the progression of Parkinson's disease through RAB31P. *Biol Chem* 2019; 400: 1217-1228.
- 14) Ye X, Hemida MG, Qiu Y, Hanson PJ, Zhang HM, Yang D. MiR-126 promotes coxsackievirus replication by mediating cross-talk of ERK1/2 and Wnt/beta-catenin signal pathways. *Cell Mol Life Sci* 2013; 70: 4631-4644.
- 15) Gai HY, Wu C, Zhang Y, Wang D. Long non-coding RNA CHRF modulates the progression of cerebral ischemia/reperfusion injury via miR-126/SOX6 signaling pathway. *Biochem Biophys Res Commun* 2019; 514: 550-557.
- 16) Salajegheh A, Vosgha H, Rahman MA, Amin M, Smith RA, Lam AK. Interactive role of miR-126 on VEGF-A and progression of papillary and undifferentiated thyroid carcinoma. *Hum Pathol* 2016; 51: 75-85.
- 17) Zheng X, Liu M, Song Y, Feng C. Long Noncoding RNA-ATB Impairs the Function of Tumor Suppressor miR-126-Mediated Signals in Endometrial Cancer for Tumor Growth and Metastasis. *Cancer Biother Radiopharm* 2019; 34: 47-55.
- 18) Nie ZC, Weng WH, Shang YS, Long Y, Li J, Xu YT, Li Z. MicroRNA-126 is down-regulated in human esophageal squamous cell carcinoma and inhibits the proliferation and migration in EC109 cell via PI3K/AKT signaling pathway. *Int J Clin Exp Pathol* 2015; 8: 4745-4754.
- 19) Di Paolo D, Pontis F, Moro M, Centonze G, Bertolini G, Milione M, Mensah M, Segale M, Petrarola I, Borzi C, Suatoni P, Brignole C, Perri P, Ponzoni M, Pastorino U, Sozzi G, Fortunato O. Cotargeting of miR-126-3p and miR-221-3p inhibits PIK3R2 and PTEN, reducing lung cancer growth and metastasis by blocking AKT and CXCR4 signalling. *Mol Oncol* 2021; 15: 2969-2988.
- 20) Yu B, Jiang Y, Wang X, Wang S. An integrated hypothesis for miR-126 in vascular disease. *Med Res Arch* 2020; 8: 2133.
- 21) Lagos-Quintana M, Rauhut R, Yalcin A, Meyer J, Lendeckel W, Tuschl T. Identification of tissue-specific microRNAs from mouse. *Curr Biol* 2002; 12: 735-739.
- 22) Landgraf P, Rusu M, Sheridan R, Sewer A, Iovino N, Aravin A, Pfeffer S, Rice A, Kamphorst AO, Landthaler M, Lin C, Socci ND, Hermida L, Fulci V, Chiaretti S, Foa R, Schliwka J, Fuchs U, Novosel A, Muller RU, Schermer B, Bissels U, Inman J, Phan Q, Chien M, Weir DB, Choksi R, De Vita G, Frezzetti D, Trompeter H, Hornung V, Teng G, Hartmann G, Palkovits M, Di Lauro R, Wernet P, Macino G, Rogler CE, Nagle JW, Ju J, Papavasiliou FN, Benzing T, Lichter P, Tam W, Brownstein MJ, Bosio A, Borkhardt A, Russo JJ, Sander C, Zavolan M, Tuschl T. A mammalian microRNA expression atlas based on small RNA library sequencing. *Cell* 2007; 129: 1401-1414.

- 23) Harris TA, Yamakuchi M, Kondo M, Oettgen P, Lowenstein CJ. Ets-1 and Ets-2 regulate the expression of microRNA-126 in endothelial cells. *Arterioscler Thromb Vasc Biol* 2010; 30: 1990-1997.
- 24) Tu J, Cheung HH, Lu G, Chan CL, Chen Z, Chan WY. microRNA-126 Is a Tumor Suppressor of Granulosa Cell Tumor Mediated by Its Host Gene EGFL7. *Front Oncol* 2019; 9: 486.
- 25) Song L, Xie X, Yu S, Peng F, Peng L. MicroRNA126 inhibits proliferation and metastasis by targeting pik3r2 in prostate cancer. *Mol Med Rep* 2016; 13: 1204-1210.
- 26) Song L, Li D, Gu Y, Wen ZM, Jie J, Zhao D, Peng LP. MicroRNA-126 Targeting PIK3R2 Inhibits NSCLC A549 Cell Proliferation, Migration, and Invasion by Regulation of PTEN/PI3K/AKT Pathway. *Clin Lung Cancer* 2016; 17: e65-e75.
- 27) Li F. Expression and correlation of miR-124 and miR-126 in breast cancer. *Oncol Lett* 2019; 17: 5115-5119.
- 28) Du C, Lv Z, Cao L, Ding C, Gyabaah OA, Xie H, Zhou L, Wu J, Zheng S. MiR-126-3p suppresses tumor metastasis and angiogenesis of hepatocellular carcinoma by targeting LRP6 and PIK3R2. *J Transl Med* 2014; 12: 259.
- 29) Kong R, Ma Y, Feng J, Li S, Zhang W, Jiang J, Zhang J, Qiao Z, Yang X, Zhou B. The crucial role of miR-126 on suppressing progression of esophageal cancer by targeting VEGF-A. *Cell Mol Biol Lett* 2016; 21: 3.
- 30) Ebrahimi F, Gopalan V, Wahab R, Lu CT, Smith RA, Lam AK. Deregulation of miR-126 expression in colorectal cancer pathogenesis and its clinical significance. *Exp Cell Res* 2015; 339: 333-341.
- 31) Dong Y, Fu C, Guan H, Zhang Z, Zhou T, Li B. Prognostic significance of miR-126 in various cancers: a meta-analysis. *Onco Targets Ther* 2016; 9: 2547-2555.
- 32) Sasahira T, Kurihara M, Bhawal UK, Ueda N, Shimomoto T, Yamamoto K, Kirita T, Kuniyasu H. Downregulation of miR-126 induces angiogenesis and lymphangiogenesis by activation of VEGF-A in oral cancer. *Br J Cancer* 2012; 107: 700-706.
- 33) Dornan GL, Burke JE. Molecular Mechanisms of Human Disease Mediated by Oncogenic and Primary Immunodeficiency Mutations in Class IA Phosphoinositide 3-Kinases. *Front Immunol* 2018; 9: 575.
- 34) Datta K, Bellacosa A, Chan TO, Tsichlis PN. Akt is a direct target of the phosphatidylinositol 3-kinase. Activation by growth factors, v-src and v-Ha-ras, in Sf9 and mammalian cells. *J Biol Chem* 1996; 271: 30835-30839.
- 35) Guo C, Sah JF, Beard L, Willson JK, Markowitz SD, Guda K. The noncoding RNA, miR-126, suppresses the growth of neoplastic cells by targeting phosphatidylinositol 3-kinase signaling and is frequently lost in colon cancers. *Genes Chromosomes Cancer* 2008; 47: 939-946.
- 36) Fu R, Tong JS. miR-126 reduces trastuzumab resistance by targeting PIK3R2 and regulating AKT/mTOR pathway in breast cancer cells. *J Cell Mol Med* 2020; 24: 7600-7608.
- 37) Baldassari F, Zerbinati C, Galasso M, Corra F, Minotti L, Agnoletto C, Previati M, Croce CM, Volinia S. Screen for MicroRNA and Drug Interactions in Breast Cancer Cell Lines Points to miR-126 as a Modulator of CDK4/6 and PIK3CA Inhibitors. *Front Genet* 2018; 9: 174.
- 38) Wong RS. Apoptosis in cancer: from pathogenesis to treatment. *J Exp Clin Cancer Res* 2011; 30: 87.
- 39) Ichikawa R, Kawasaki R, Iwata A, Otani S, Nishio E, Nomura H, Fujii T. MicroRNA1263p suppresses HeLa cell proliferation, migration and invasion, and increases apoptosis via the PI3K/PDK1/AKT pathway. *Oncol Rep* 2020; 43: 1300-1308.
- 40) Jia Z, Zhang Y, Xu Q, Guo W, Guo A. miR-126 suppresses epithelial-to-mesenchymal transition and metastasis by targeting PI3K/AKT/Snail signaling of lung cancer cells. *Oncol Lett* 2018; 15: 7369-7375.
- 41) Xu J, Zheng H, Yuan S, Zhou B, Zhao W, Pan Y, Qi D. Overexpression of ANLN in lung adenocarcinoma is associated with metastasis. *Thorac Cancer* 2019; 10: 1702-1709.
- 42) Lian YF, Huang YL, Wang JL, Deng MH, Xia TL, Zeng MS, Chen MS, Wang HB, Huang YH. Anillin is required for tumor growth and regulated by miR-15a/miR-16-1 in HBV-related hepatocellular carcinoma. *Aging (Albany NY)* 2018; 10: 1884-1901.
- 43) Jiao Z, Yu A, He X, Xuan Y, Zhang H, Wang G, Shi M, Wang T. Bioinformatics analysis to determine the prognostic value and prospective pathway signaling of miR-126 in non-small cell lung cancer. *Ann Transl Med* 2020; 8: 1639.
- 44) Li J, Xu H, Wang Q, Wang S, Xiong N. 14-3-3zeta promotes gliomas cells invasion by regulating Snail through the PI3K/AKT signaling. *Cancer Med* 2019; 8: 783-794.
- 45) Vasan N, Baselga J, Hyman DM. A view on drug resistance in cancer. *Nature* 2019; 575: 299-309.
- 46) Yan J, Dang Y, Liu S, Zhang Y, Zhang G. LncRNA HOTAIR promotes cisplatin resistance in gastric cancer by targeting miR-126 to activate the PI3K/AKT/MRP1 genes. *Tumour Biol* 2016; 8: 266-289.
- 47) Di-Leva G, Piovan C, Gasparini P, Ngankeu A, Taccioli C, Briskin D, Cheung DG, Bolon B, Anderlucchi L, Alder H, Nuovo G, Li M, Iorio MV, Galasso M, Santhanam R, Marcucci G, Perrotti D, Powell KA, Bratasz A, Garofalo M, Nephew KP, Croce CM. Estrogen mediated-activation of miR-191/425 cluster modulates tumorigenicity of breast cancer cells depending on estrogen receptor status. *PLoS Genet* 2013; 9.
- 48) Zhang Y, Yong L, Luo Y, Ding X, Xu D, Gao X, Yan S, Wang Q, Luo J, Pu D, Zou J. Enhancement of HIFU ablation by sonosensitizer-loading liquid fluorocarbon nanoparticles with pre-targeting in a mouse model. *Sci Rep* 2019; 9: 6982.
- 49) Fu Y, Lin L, Xia L. MiR-107 function as a tumor suppressor gene in colorectal cancer by targeting transferrin receptor 1. *Cell Mol Biol Lett* 2019; 24: 31.
- 50) Shi H, Bi H, Sun X, Dong H, Jiang Y, Mu H, Liu G, Kong W, Gao R, Su J. Antitumor effects of Tuberimoside-1 in NCI-H1299 cells are mediated by microRNA-126-5p-induced inactivation of VEGF-A/VEGFR-2/ERK signaling pathway. *Mol Med Rep* 2018; 17: 4327-4336.
- 51) Kristensen TB, Knutsson ML, Wehland M, Laursen BE, Grimm D, Warnke E, Magnusson NE. Anti-vas-

- cular endothelial growth factor therapy in breast cancer. *Int J Mol Sci* 2014; 15: 23024-23041.
- 52) Jing BQ, Ou Y, Zhao L, Xie Q, Zhang YX. Experimental study on the prevention of liver cancer angiogenesis via miR-126. *Eur Rev Med Pharmacol Sci* 2017; 21: 5096-5100.
 - 53) Liu L, Yuan L, Huang D, Han Q, Cai J, Wang S, Cao J. miR126 regulates the progression of epithelial ovarian cancer in vitro and in vivo by targeting VEGFA. *Int J Oncol* 2020; 57: 825-834.
 - 54) Zhu N, Zhang D, Xie H, Zhou Z, Chen H, Hu T, Bai Y, Shen Y, Yuan W, Jing Q, Qin Y. Endothelial-specific intron-derived miR-126 is down-regulated in human breast cancer and targets both VEGFA and PIK3R2. *Mol Cell Biochem* 2011; 351: 157-164.
 - 55) Chen H, Li L, Wang S, Lei Y, Ge Q, Lv N, Zhou X, Chen C. Reduced miR-126 expression facilitates angiogenesis of gastric cancer through its regulation on VEGF-A. *Oncotarget* 2014; 5: 11873-11885.
 - 56) Luo J, Zhu C, Wang H, Yu L, Zhou J. MicroRNA-126 affects ovarian cancer cell differentiation and invasion by modulating expression of vascular endothelial growth factor. *Oncol Lett* 2018; 15: 5803-5808.
 - 57) Alhasan L. MiR-126 Modulates Angiogenesis in Breast Cancer by Targeting VEGF-A mRNA. *Asian Pac J Cancer Prev* 2019; 20: 193-197.
 - 58) Zhao Z, Li Y, Shukla R, Liu H, Jain A, Barve A, Cheng K. Development of a Biocompatible Copolymer Nanocomplex to Deliver VEGF siRNA for Triple Negative Breast Cancer. *Theranostics* 2019; 9: 4508-4524.
 - 59) Wang L, Liu X, Wang H, Wang S. Correlation of the expression of vascular endothelial growth factor and its receptors with microvessel density in ovarian cancer. *Oncol Lett* 2013; 6: 175-180.
 - 60) Incio J, Ligibel JA, Mcmanus DT, Suboj P, Jung K, Kawaguchi K, Pinter M, Babykutty S, Chin SM, Vardam TD, Huang Y, Rahbari NN, Roberge S, Wang D, Gomes-Santos IL, Puchner SB, Schlett CL, Hoffmann U, Ancukiewicz M, Tolaney SM, Krop IE, Duda DG, Boucher Y, Fukumura D, Jain RK. Obesity promotes resistance to anti-VEGF therapy in breast cancer by up-regulating IL-6 and potentially FGF-2. *Sci Transl Med* 2018; 10.
 - 61) Kim MR, Jang JH, Park CS, Kim TK, Kim YJ, Chung J, Shim H, Nam IH, Han JM, Lee S. A Human Antibody That Binds to the Sixth Ig-Like Domain of VCAM-1 Blocks Lung Cancer Cell Migration In Vitro. *Int J Mol Sci* 2017; 18: 566.
 - 62) Rijcken E, Krieglstein CF, Anthoni C, Laukoetter MG, Mennigen R, Spiegel HU, Senninger N, Bennett CF, Schuermann G. ICAM-1 and VCAM-1 antisense oligonucleotides attenuate in vivo leucocyte adherence and inflammation in rat inflammatory bowel disease. *Gut* 2002; 51: 529-535.
 - 63) Chaves KC, Peron JP, Chammas R, Turaca LT, Pesquero JB, Braga MS, Foguer K, Schor N, Bellini MH. Endostatin gene therapy stimulates up-regulation of ICAM-1 and VCAM-1 in a metastatic renal cell carcinoma model. *Cancer Gene Ther* 2012; 19: 558-565.
 - 64) Shen J, Zhai J, You Q, Zhang G, He M, Yao X, Shen L. Cancer-associated fibroblasts-derived VCAM1 induced by H. pylori infection facilitates tumor invasion in gastric cancer. *Oncogene* 2020; 39: 2961-2974.
 - 65) Banerjee N, Kim H, Talcott S, Mertens-Talcott S. Pomegranate polyphenolics suppressed azoxymethane-induced colorectal aberrant crypt foci and inflammation: possible role of miR-126/VCAM-1 and miR-126/PI3K/AKT/mTOR. *Carcinogenesis* 2013; 34: 2814-2822.
 - 66) Tai HC, Chang AC, Yu HJ, Huang CY, Tsai YC, Lai YW, Sun HL, Tang CH, Wang SW. Osteoblast-derived WNT-induced secreted protein 1 increases VCAM-1 expression and enhances prostate cancer metastasis by down-regulating miR-126. *Oncotarget* 2014; 5: 7589-7598.
 - 67) Tan TW, Chou YE, Yang WH, Hsu CJ, Fong YC, Tang CH. Naringin suppress chondrosarcoma migration through inhibition vascular adhesion molecule-1 expression by modulating miR-126. *Int Immunopharmacol* 2014; 22: 107-114.
 - 68) Taverna S, Amodeo V, Saieva L, Russo A, Giallombardo M, De-Leo G, Alessandro R. Exosomal shuttling of miR-126 in endothelial cells modulates adhesive and migratory abilities of chronic myelogenous leukemia cells. *Mol Cancer* 2014; 13: 169.
 - 69) Ding YB, Chen GY, Xia JG, Zang XW, Yang HY, Yang L. Association of VCAM-1 overexpression with oncogenesis, tumor angiogenesis and metastasis of gastric carcinoma. *World J Gastroenterol* 2003; 9: 1409-1414.
 - 70) Kong DH, Kim YK, Kim MR, Jang JH, Lee S. Emerging Roles of Vascular Cell Adhesion Molecule-1 (VCAM-1) in Immunological Disorders and Cancer. *Int J Mol Sci* 2018; 19: 1057.
 - 71) Chen Q, Zhang XH, Massague J. Macrophage binding to receptor VCAM-1 transmits survival signals in breast cancer cells that invade the lungs. *Cancer Cell* 2011; 20: 538-549.
 - 72) Sarvi S, Patel H, Li J, Dodd GL, Creedon H, Muir M, Ward J, Dawson JC, Lee M, Culley J, Salter DM, Sims AH, Byron A, Brunton VG. Kindlin-1 Promotes Pulmonary Breast Cancer Metastasis. *Cancer Res* 2018; 78: 1484-1496.
 - 73) Cook-Mills JM, Johnson JD, Deem TL, Ochi A, Wang L, Zheng Y. Calcium mobilization and Rac1 activation are required for VCAM-1 (vascular cell adhesion molecule-1) stimulation of NADPH oxidase activity. *Biochem J* 2004; 378: 539-547.
 - 74) Pennica D, Swanson TA, Welsh JW, Roy MA, Lawrence DA, Lee J, Brush J, Taneyhill LA, Deuel B, Lew M, Watanabe C, Cohen RL, Melhem MF, Finley GG, Quirke P, Goddard AD, Hillan KJ, Gurney AL, Botstein D, Levine AJ. WISP genes are members of the connective tissue growth factor family that are up-regulated in wnt-1-transformed cells and aberrantly expressed in human colon tumors. *Proc Natl Acad Sci U S A* 1998; 95: 14717-14722.
 - 75) Byrne GJ, Bundred NJ. Surrogate markers of tumoral angiogenesis. *Int J Biol Markers* 2000; 15: 334-339.
 - 76) Garmy-Susini B, Jin H, Zhu Y, Sung RJ, Hwang R, Varner J. Integrin alpha4beta1-VCAM-1-mediated adhesion between endothelial and mural cells is required for blood vessel maturation. *J Clin Invest* 2005; 115: 1542-1551.

- 77) Fukushi J, Ono M, Morikawa W, Iwamoto Y, Kuwano M. The activity of soluble VCAM-1 in angiogenesis stimulated by IL-4 and IL-13. *J Immunol* 2000; 165: 2818-2823.
- 78) Kim TK, Park CS, Na HJ, Lee K, Yoon A, Chung J, Lee S. Ig-like domain 6 of VCAM-1 is a potential therapeutic target in TNF α -induced angiogenesis. *Exp Mol Med* 2017; 49: e294.
- 79) Chen M, Peng W, Hu S, Deng J. miR-126/VCAM-1 regulation by naringin suppresses cell growth of human non-small cell lung cancer. *Oncol Lett* 2018; 16: 4754-4760.
- 80) Liu Y, Li L, Li Y, Zhao X. Research Progress on Tumor-Associated Macrophages and Inflammation in Cervical Cancer. *Biomed Res Int* 2020; 2020: 6842963.
- 81) Franklin RA, Liao W, Sarkar A, Kim MV, Bivona MR, Liu K, Pamer EG, Li MO. The cellular and molecular origin of tumor-associated macrophages. *Science* 2014; 344: 921-925.
- 82) Ye H, Zhou Q, Zheng S, Li G, Lin Q, Wei L, Fu Z, Zhang B, Liu Y, Li Z, Chen R. Tumor-associated macrophages promote progression and the Warburg effect via CCL18/NF- κ B/VCAM-1 pathway in pancreatic ductal adenocarcinoma. *Cell Death Dis* 2018; 9: 453.
- 83) Pham K, Huynh D, Le L, Delitto D, Yang L, Huang J, Kang Y, Steinberg MB, Li J, Zhang L, Liu D, Tang MS, Liu C, Wang H. E-cigarette promotes breast carcinoma progression and lung metastasis: Macrophage-tumor cells crosstalk and the role of CCL5 and VCAM-1. *Cancer Lett* 2020; 491: 132-145.
- 84) Shen CK, Huang BR, Yeh WL, Chen CW, Liu YS, Lai SW, Tseng WP, Lu DY, Tsai CF. Regulatory effects of IL-1 β in the interaction of GBM and tumor-associated monocyte through VCAM-1 and ICAM-1. *Eur J Pharmacol* 2021; 905: 174216.
- 85) Nichol D, Stuhlmann H. EGFL7: a unique angiogenic signaling factor in vascular development and disease. *Blood* 2012; 119: 1345-1352.
- 86) Sun Y, Bai Y, Zhang F, Wang Y, Guo Y, Guo L. miR-126 inhibits non-small cell lung cancer cells proliferation by targeting EGFL7. *Biochem Biophys Res Commun* 2010; 391: 1483-1489.
- 87) Hu MH, Ma CY, Wang XM, Ye CD, Zhang GX, Chen L, Wang JG. MicroRNA-126 inhibits tumor proliferation and angiogenesis of hepatocellular carcinoma by down-regulating EGFL7 expression. *Oncotarget* 2016; 7: 66922-66934.
- 88) Zhang Y, Qin X, Jiang J, Zhao W. MicroRNA-126 exerts antitumor functions in ovarian cancer by targeting EGFL7 and affecting epithelial-to-mesenchymal transition and ERK/MAPK signaling pathway. *Oncol Lett* 2020; 20: 1327-1335.
- 89) Gong C, Fang J, Li G, Liu HH, Liu ZS. Effects of microRNA-126 on cell proliferation, apoptosis and tumor angiogenesis via the down-regulating ERK signaling pathway by targeting EGFL7 in hepatocellular carcinoma. *Oncotarget* 2017; 8: 52527-52542.
- 90) Andersen M, Trapani D, Ravn J, Sorensen JB, Andersen CB, Grauslund M, Santoni-Rugiu E. Methylation-associated Silencing of microRNA-126 and its Host Gene EGFL7 in Malignant Pleural Mesothelioma. *Anticancer Res* 2015; 35: 6223-6229.
- 91) Yang X, Wu H, Ling T. Suppressive effect of microRNA-126 on oral squamous cell carcinoma in vitro. *Mol Med Rep* 2014; 10: 125-130.
- 92) Rouigari M, Dehbashi M, Ghaedi K, Pourhossein M. Targetome Analysis Revealed Involvement of MiR-126 in Neurotrophin Signaling Pathway: A Possible Role in Prevention of Glioma Development. *Cell J* 2018; 20: 150-156.
- 93) Hansen TF, Andersen RF, Olsen DA, Sorensen FB, Jakobsen A. Prognostic importance of circulating epidermal growth factor-like domain 7 in patients with metastatic colorectal cancer treated with chemotherapy and bevacizumab. *Sci Rep* 2017; 7: 2388.
- 94) Juan Z, Dake C, Tanaka K, Shuixiang H. EGFL7 as a novel therapeutic candidate regulates cell invasion and anoikis in colorectal cancer through PI3K/AKT signaling pathway. *Int J Clin Oncol* 2021; 26: 1099-1108.
- 95) Guo YP, Wang ZF, Li N, Lei QQ, Cheng Q, Shi LG, Zhou SL, Wang XH, Sun Y, Kong LF. Suppression of lncRNA HOTAIR alleviates RCC angiogenesis through regulating miR-126/EGFL7 axis. *Am J Physiol Cell Physiol* 2021; 320: C880-C891.
- 96) Chen CM, Chu TH, Chou CC, Chien CY, Wang JS, Huang CC. Exosome-derived microRNAs in oral squamous cell carcinomas impact disease prognosis. *Oral Oncol* 2021; 120: 105402.
- 97) Salama Y, Heida AH, Yokoyama K, Takahashi S, Hattori K, Heissig B. The EGFL7-ITGB3-KLF2 axis enhances survival of multiple myeloma in preclinical models. *Blood Adv* 2020; 4: 1021-1037.
- 98) Wu F, Yang LY, Li YF, Ou DP, Chen DP, Fan C. Novel role for epidermal growth factor-like domain 7 in metastasis of human hepatocellular carcinoma. *Hepatology* 2009; 50: 1839-1850.
- 99) Delfortrie S, Pinte S, Mattot V, Samson C, Villain G, Caetano B, Lauridant-Philippin G, Baranzelli MC, Bonnetterre J, Trottein F, Faveeuw C, Soncin F. Eglf7 promotes tumor escape from immunity by repressing endothelial cell activation. *Cancer Res* 2011; 71: 7176-7186.
- 100) Lamalice L, Le-Boeuf F, Huot J. Endothelial cell migration during angiogenesis. *Circ Res* 2007; 100: 782-794.
- 101) Teicher BA, Fricker SP. CXCL12 (SDF-1)/CXCR4 pathway in cancer. *Clin Cancer Res* 2010; 16: 2927-2931.
- 102) Lu L, Lu M, Pei Y, Chen J, Qin L, Zhu W, Jia H. Down-regulation of SDF1- α expression in tumor microenvironment is associated with aspirin-mediated suppression of the pro-metastasis effect of sorafenib in hepatocellular carcinoma. *Acta Biochim Biophys Sin (Shanghai)* 2015; 47: 988-996.
- 103) Kryczek I, Wei S, Keller E, Liu R, Zou W. Stroma-derived factor (SDF-1/CXCL12) and human tumor pathogenesis. *Am J Physiol Cell Physiol* 2007; 292: C987-C995.
- 104) Meier R, Muhlethaler-Mottet A, Flahaut M, Coulon A, Fusco C, Louache F, Auderset K, Bourlout KB, Daudigeos E, Ruegg C, Vassal G, Gross N, Joseph JM. The chemokine receptor CXCR4 strongly promotes neuroblastoma primary tumour and

- metastatic growth, but not invasion. *PLoS One* 2007; 2: e1016.
- 105) Barbieri F, Bajetto A, Stumm R, Pattarozzi A, Porcile C, Zona G, Dorcaratto A, Ravetti JL, Minuto F, Spaziante R, Schettini G, Ferone D, Florio T. Overexpression of stromal cell-derived factor 1 and its receptor CXCR4 induces autocrine/paracrine cell proliferation in human pituitary adenomas. *Clin Cancer Res* 2008; 14: 5022-5032.
 - 106) O'Dwyer J, Cullen M, Fattah S, Murphy R, Stefanovic S, Kovarova L, Pravda M, Velebny V, Heise A, Duffy GP, Cryan SA. Development of a Sustained Release Nano-In-Gel Delivery System for the Chemotactic and Angiogenic Growth Factor Stromal-Derived Factor 1alpha. *Pharmaceutics* 2020; 12: 513.
 - 107) Wu S, Yuan W, Luo W, Nie K, Wu X, Meng X, Shen Z, Wang X. MiR-126 downregulates CXCL12 expression in intestinal epithelial cells to suppress the recruitment and function of macrophages and tumorigenesis in a murine model of colitis-associated colorectal cancer. *Mol Oncol* 2022; 513.
 - 108) Li Z, Li N, Wu M, Li X, Luo Z, Wang X. Expression of miR-126 suppresses migration and invasion of colon cancer cells by targeting CXCR4. *Mol Cell Biochem* 2013; 381: 233-242.
 - 109) Knufter H, Preiss R. Serum interleukin-6 levels in colorectal cancer patients--a summary of published results. *Int J Colorectal Dis* 2010; 25: 135-140.
 - 110) Liu Y, Zhou Y, Feng X, Yang P, Yang J, An P, Wang H, Ye S, Yu C, He Y, Luo H. Low expression of microRNA-126 is associated with poor prognosis in colorectal cancer. *Genes Chromosomes Cancer* 2014; 53: 358-365.
 - 111) Bertolini G, D'Amico L, Moro M, Landoni E, Perego P, Miceli R, Gatti L, Andriani F, Wong D, Caserini R, Tortoreto M, Milione M, Ferracini R, Mariani L, Pastorino U, Roato I, Sozzi G, Roz L. Microenvironment-Modulated Metastatic CD133+/CXCR4+/EpCAM- Lung Cancer-Initiating Cells Sustain Tumor Dissemination and Correlate with Poor Prognosis. *Cancer Res* 2015; 75: 3636-3649.
 - 112) Yuan W, Guo YQ, Li XY, Deng MZ, Shen ZH, Bo CB, Dai YF, Huang MY, Yang ZY, Quan YS, Tian L, Wang X. MicroRNA-126 inhibits colon cancer cell proliferation and invasion by targeting the chemokine (C-X-C motif) receptor 4 and Ras homolog gene family, member A, signaling pathway. *Oncotarget* 2016; 7: 60230-60244.
 - 113) Xiao J, Lai H, Wei SH, Ye ZS, Gong FS, Chen LC. lncRNA HOTAIR promotes gastric cancer proliferation and metastasis via targeting miR-126 to active CXCR4 and RhoA signaling pathway. *Cancer Med* 2019; 8: 6768-6779.
 - 114) Qian Y, Wang X, Lv Z, Guo C, Yang Y, Zhang J, Wang X. MicroRNA126 is downregulated in thyroid cancer cells, and regulates proliferation, migration and invasion by targeting CXCR4. *Mol Med Rep* 2016; 14: 453-459.
 - 115) Hou X, Shen Z, Li N, Kong X, Sheng K, Wang J, Wang Y. A novel fungal beta-propeller phytase from nematophagous *Arthrobotrys oligospora*: characterization and potential application in phosporus and mineral release for feed processing. *Microb Cell Fact* 2020; 19: 84.
 - 116) Kim M, Kim S, Lee SH, Kim W, Sohn MJ, Kim HS, Kim J, Jho EH. Merlin inhibits Wnt/beta-catenin signaling by blocking LRP6 phosphorylation. *Cell Death Differ* 2016; 23: 1638-1647.
 - 117) Raisch J, Cote-Biron A, Rivard N. A Role for the WNT Co-Receptor LRP6 in Pathogenesis and Therapy of Epithelial Cancers. *Cancers (Basel)* 2019; 11: 1162.
 - 118) Ma J, Lu W, Chen D, Xu B, Li Y. Role of Wnt Co-Receptor LRP6 in Triple Negative Breast Cancer Cell Migration and Invasion. *J Cell Biochem* 2017; 118: 2968-2976.
 - 119) Fan J, Liu L, Liu Q, Cui Y, Yao B, Zhang M, Gao Y, Fu Y, Dai H, Pan J, Qiu Y, Liu CH, He F, Wang Y, Zhang L. CKIP-1 limits foam cell formation and inhibits atherosclerosis by promoting degradation of Oct-1 by REGgamma. *Nat Commun* 2019; 10: 425.
 - 120) Fan JL, Zhang L, Bo XH. MiR-126 on mice with coronary artery disease by targeting S1PR2. *Eur Rev Med Pharmacol Sci* 2020; 24: 893-904.
 - 121) Joshi AA, Lerman JB, Dey AK, Sajja AP, Belur AD, Elnabawi YA, Rodante JA, Aberra TM, Chung J, Salahuddin T, Natarajan B, Dave J, Goyal A, Groenendyk JW, Rivers JP, Baumer Y, Teague HL, Playford MP, Bluemke DA, Ahlman MA, Chen MY, Gelfand JM, Mehta NN. Association Between Aortic Vascular Inflammation and Coronary Artery Plaque Characteristics in Psoriasis. *JAMA Cardiol* 2018; 3: 949-956.
 - 122) Mao H, Li L, Fan Q, Angelini A, Saha PK, Wu H, Ballantyne CM, Hartig SM, Xie L, Pi X. Loss of bone morphogenetic protein-binding endothelial regulator causes insulin resistance. *Nat Commun* 2021; 12: 1927.
 - 123) Gimbrone MJ, Garcia-Cardena G. Endothelial Cell Dysfunction and the Pathobiology of Atherosclerosis. *Circ Res* 2016; 118: 620-636.
 - 124) Liu M, Lutz H, Zhu D, Huang K, Li Z, Dinh PC, Gao J, Zhang Y, Cheng K. Bispecific Antibody Inhalation Therapy for Redirecting Stem Cells from the Lungs to Repair Heart Injury. *Adv Sci (Weinh)* 2020; 8: 2002127.
 - 125) Tang F, Yang TL. MicroRNA-126 alleviates endothelial cells injury in atherosclerosis by restoring autophagic flux via inhibiting of PI3K/Akt/mTOR pathway. *Biochem Biophys Res Commun* 2018; 495: 1482-1489.
 - 126) Chen L, Wang J, Wang B, Yang J, Gong Z, Zhao X, Zhang C, Du K. MiR-126 inhibits vascular endothelial cell apoptosis through targeting PI3K/Akt signaling. *Ann Hematol* 2016; 95: 365-374.
 - 127) Sui XQ, Xu ZM, Xie MB, Pei DA. Resveratrol inhibits hydrogen peroxide-induced apoptosis in endothelial cells via the activation of PI3K/Akt by miR-126. *J Atheroscler Thromb* 2014; 21: 108-118.
 - 128) Li WM, Yue JN, Guo DQ, Fu WG. MiR-126 promotes endothelial cell apoptosis by targeting PI3K/Akt in rats with lower limb arteriosclerosis obliterans. *Eur Rev Med Pharmacol Sci* 2019; 23: 327-333.
 - 129) Wolf D, Ley K. Immunity and Inflammation in Atherosclerosis. *Circ Res* 2019; 124: 315-327.

- 130) Liao L, Yang Q, Li H, Meng R, Li Y. miR-454-3p prevents ox-LDL-induced apoptosis in HAECs by targeting TRPC3. *Exp Ther Med* 2021; 21: 323.
- 131) Kim JS, Kim JH, Palaniyandi SA, Lee CC, You JW, Yang H, Yoon PJ, Yang SH, Lee KW. Yak-Kong Soybean (*Glycine max*) Fermented by a Novel *Pediococcus pentosaceus* Inhibits the Oxidative Stress-Induced Monocyte-Endothelial Cell Adhesion. *Nutrients* 2019; 11: 1380.
- 132) Vogel ME, Idelman G, Konanah ES, Zucker SD. Bilirubin Prevents Atherosclerotic Lesion Formation in Low-Density Lipoprotein Receptor-Deficient Mice by Inhibiting Endothelial VCAM-1 and ICAM-1 Signaling. *J Am Heart Assoc* 2017; 6: e004820.
- 133) Libby P. Inflammation in atherosclerosis. *Nature* 2002; 420: 868-874.
- 134) Lipinski MJ, Campbell KA, Duong SQ, Welch TJ, Garmey JC, Doran AC, Skafien MD, Oldham SN, Kelly KA, Mcnamara CA. Loss of Id3 increases VCAM-1 expression, macrophage accumulation, and atherogenesis in *Ldlr*^{-/-} mice. *Arterioscler Thromb Vasc Biol* 2012; 32: 2855-2861.
- 135) Sun C, Alkhoury K, Wang YI, Foster GA, Radecke CE, Tam K, Edwards CM, Facciotti MT, Armstrong EJ, Knowlton AA, Newman JW, Passerini AG, Simon SI. IRF-1 and miRNA126 modulate VCAM-1 expression in response to a high-fat meal. *Circ Res* 2012; 111: 1054-1064.
- 136) Pan X, Hou R, Ma A, Wang T, Wu M, Zhu X, Yang S, Xiao X. Atorvastatin Upregulates the Expression of miR-126 in Apolipoprotein E-knockout Mice with Carotid Atherosclerotic Plaque. *Cell Mol Neurobiol* 2017; 37: 29-36.
- 137) Yokoyama Y, Mise N, Suzuki Y, Tada-Oikawa S, Izuoka K, Zhang L, Zong C, Takai A, Yamada Y, Ichihara S. MicroRNAs as Potential Mediators for Cigarette Smoking Induced Atherosclerosis. *Int J Mol Sci* 2018; 19: 1097.
- 138) Yuan X, Chen J, Dai M. Paeonol promotes microRNA-126 expression to inhibit monocyte adhesion to ox-LDL-injured vascular endothelial cells and block the activation of the PI3K/Akt/NF-kappaB pathway. *Int J Mol Med* 2016; 38: 1871-1878.
- 139) Parma L, Peters H, Sluiter TJ, Simons KH, Lazari P, de Vries MR, Quax P. bFGF blockade reduces intraplaque angiogenesis and macrophage infiltration in atherosclerotic vein graft lesions in ApoE3*Leiden mice. *Sci Rep* 2020; 10: 15968.
- 140) Yuan R, Shi W, Xin Q, Yang B, Hoi MP, Lee SM, Cong W, Chen K. Tetramethylpyrazine and Paeoniflorin Inhibit Oxidized LDL-Induced Angiogenesis in Human Umbilical Vein Endothelial Cells via VEGF and Notch Pathways. *Evid Based Complement Alternat Med* 2018; 2018: 3082507.
- 141) Luo X, Li W, Bai Y, Du L, Wu R, Li Z. Relation between carotid vulnerable plaques and peripheral leukocyte: a case-control study of comparison utilizing multi-parametric contrast-enhanced ultrasound. *Bmc Med Imaging* 2019; 19: 74.
- 142) Chen H, Chen L, Liang R, Wei J. Ultrasound and magnetic resonance molecular imaging of atherosclerotic neovasculature with perfluorocarbon magnetic nanocapsules targeted against vascular endothelial growth factor receptor 2 in rats. *Mol Med Rep* 2017; 16: 5986-5996.
- 143) Ahmad S, Hewett PW, Wang P, Al-Ani B, Cudmore M, Fujisawa T, Haigh JJ, le Noble F, Wang L, Mukhopadhyay D, Ahmed A. Direct evidence for endothelial vascular endothelial growth factor receptor-1 function in nitric oxide-mediated angiogenesis. *Circ Res* 2006; 99: 715-722.
- 144) Cai T, Cui X, Zhang K, Zhang A, Liu B, Mu JJ. LncRNA TNK2-AS1 regulated ox-LDL-stimulated HASMC proliferation and migration via modulating VEGFA and FGF1 expression by sponging miR-150-5p. *J Cell Mol Med* 2019; 23: 7289-7298.
- 145) Li J, Cao GY, Zhang XF, Meng ZQ, Gan L, Li JX, Lan XY, Yang CL, Zhang CF. Chinese Medicine She-Xiang-Xin-Tong-Ning, Containing Moschus, Corydalis and Ginseng, Protects from Myocardial Ischemia Injury via Angiogenesis. *Am J Chin Med* 2020; 48: 107-126.
- 146) Zhou Y, Zhu X, Cui H, Shi J, Yuan G, Shi S, Hu Y. The Role of the VEGF Family in Coronary Heart Disease. *Front Cardiovasc Med* 2021; 8: 738325.
- 147) Zhao D, Shao H. Effect of blood purification on serum miR-126 and VEGF levels in the process of atherosclerosis in uremic patients under maintenance hemodialysis. *Kaohsiung J Med Sci* 2018; 34: 447-455.
- 148) Sezer ZC, Timirci-Kahraman O, Amadou FZ, Fazlogullari O, Basaran C, Catal T, Zeybek U, Bermek H. Expression of Eglf7 and miRNA-126-5p in Symptomatic Carotid Artery Disease. *Genet Test Mol Biomarkers* 2016; 20: 125-129.
- 149) Campagnolo L, Leahy A, Chitnis S, Koschnick S, Fitch MJ, Fallon JT, Loskutoff D, Taubman MB, Stuhlmann H. EGFL7 is a chemoattractant for endothelial cells and is up-regulated in angiogenesis and arterial injury. *Am J Pathol* 2005; 167: 275-284.
- 150) Sun YQ, Zhang F, Bai YF, Guo LL. [miR-126 modulates the expression of epidermal growth factor-like domain 7 in human umbilical vein endothelial cells in vitro]. *Nan Fang Yi Ke Da Xue Xue Bao* 2010; 30: 767-770.
- 151) Li Q, Cheng K, Wang AY, Xu QG, Fu ZF, He SY, Xu PX. microRNA-126 inhibits tube formation of HUVECs by interacting with EGFL7 and down-regulating PI3K/AKT signaling pathway. *Biomed Pharmacother* 2019; 116: 109007.
- 152) Yu S, Wong SL, Lau CW, Huang Y, Yu CM. Oxidized LDL at low concentration promotes in-vitro angiogenesis and activates nitric oxide synthase through PI3K/Akt/eNOS pathway in human coronary artery endothelial cells. *Biochem Biophys Res Commun* 2011; 407: 44-48.
- 153) Bir SC, Xiong Y, Kevil CG, Luo J. Emerging role of PKA/eNOS pathway in therapeutic angiogenesis for ischaemic tissue diseases. *Cardiovasc Res* 2012; 95: 7-18.
- 154) Qiu J, Peng Q, Zheng Y, Hu J, Luo X, Teng Y, Jiang T, Yin T, Tang C, Wang G. OxLDL stimulates Id1 nucleocytoplasmic shuttling in endothelial cell angiogenesis via PI3K pathway. *Biochim Biophys Acta* 2012; 1821: 1361-1369.
- 155) Zhang L, Ouyang P, He G, Wang X, Song D, Yang Y, He X. Exosomes from microRNA-126 overexpressing mesenchymal stem cells promote angiogenesis by targeting the PIK3R2-mediated PI3K/Akt signalling pathway. *J Cell Mol Med* 2021; 25: 2148-2162.

- 156) Gao H, Peng C, Wu L, Gao S, Wang Z, Dai L, Wu H. Yiqi-Huoxue granule promotes angiogenesis of ischemic myocardium through miR-126/PI3K/Akt axis in endothelial cells. *Phytomedicine* 2021; 92: 153713.
- 157) Mineo C. Lipoprotein receptor signalling in atherosclerosis. *Cardiovasc Res* 2020; 116: 1254-1274.
- 158) Keramati AR, Singh R, Lin A, Faramarzi S, Ye ZJ, Mane S, Tellides G, Lifton RP, Mani A. Wild-type LRP6 inhibits, whereas atherosclerosis-linked LRP6R611C increases PDGF-dependent vascular smooth muscle cell proliferation. *Proc Natl Acad Sci U S A* 2011; 108: 1914-1918.
- 159) Go GW, Mani A. Low-density lipoprotein receptor (LDLR) family orchestrates cholesterol homeostasis. *Yale J Biol Med* 2012; 85: 19-28.
- 160) Jansen F, Stumpf T, Proebsting S, Franklin BS, Wenzel D, Pfeifer P, Flender A, Schmitz T, Yang X, Fleischmann BK, Nickenig G, Werner N. Intercellular transfer of miR-126-3p by endothelial microparticles reduces vascular smooth muscle cell proliferation and limits neointima formation by inhibiting LRP6. *J Mol Cell Cardiol* 2017; 104: 43-52.
- 161) Abi-Younes S, Sauty A, Mach F, Sukhova GK, Libby P, Luster AD. The stromal cell-derived factor-1 chemokine is a potent platelet agonist highly expressed in atherosclerotic plaques. *Circ Res* 2000; 86: 131-138.
- 162) Murad H, Rafeeq MM, Alqurashi T. Role and implications of the CXCL12/CXCR4/CXCR7 axis in atherosclerosis: still a debate. *Ann Med* 2021; 53: 1598-1612.
- 163) Merckelbach S, van der Vorst E, Kallmayer M, Rischpler C, Burgkart R, Doring Y, de Borst GJ, Schwaiger M, Eckstein HH, Weber C, Pelisek J. Expression and Cellular Localization of CXCR4 and CXCL12 in Human Carotid Atherosclerotic Plaques. *Thromb Haemost* 2018; 118: 195-206.
- 164) Schuster S, Rubil S, Endres M, Princen H, Boeckel JN, Winter K, Werner C, Laufs U. Anti-PCSK9 antibodies inhibit pro-atherogenic mechanisms in APOE*3Leiden.CETP mice. *Sci Rep* 2019; 9: 11079.
- 165) Li L, Du Z, Rong B, Zhao D, Wang A, Xu Y, Zhang H, Bai X, Zhong J. Foam cells promote atherosclerosis progression by releasing CXCL12. *Biosci Rep* 2020; 40.
- 166) Schober A, Knarren S, Lietz M, Lin EA, Weber C. Crucial role of stromal cell-derived factor-1alpha in neointima formation after vascular injury in apolipoprotein E-deficient mice. *Circulation* 2003; 108: 2491-2497.
- 167) Yamaguchi J, Kusano KF, Masuo O, Kawamoto A, Silver M, Murasawa S, Bosch-Marce M, Masuda H, Losordo DW, Isner JM, Asahara T. Stromal cell-derived factor-1 effects on ex vivo expanded endothelial progenitor cell recruitment for ischemic neovascularization. *Circulation* 2003; 107: 1322-1328.
- 168) Gao JH, Yu XH, Tang CK. CXC chemokine ligand 12 (CXCL12) in atherosclerosis: An underlying therapeutic target. *Clin Chim Acta* 2019; 495: 538-544.
- 169) Yang K, Du C, Wang X, Li F, Xu Y, Wang S, Chen S, Chen F, Shen M, Chen M, Hu M, He T, Su Y, Wang J, Zhao J. Indoxyl sulfate induces platelet hyperactivity and contributes to chronic kidney disease-associated thrombosis in mice. *Blood* 2017; 129: 2667-2679.
- 170) Walsh TG, Harper MT, Poole AW. SDF-1alpha is a novel autocrine activator of platelets operating through its receptor CXCR4. *Cell Signal* 2015; 27: 37-46.
- 171) Liehn EA, Radu E, Schuh A. Chemokine contribution in stem cell engraftment into the infarcted myocardium. *Curr Stem Cell Res Ther* 2013; 8: 278-283.
- 172) Rezaee M, Behnam B, Banach M, Sahebkar A. The Yin and Yang of carbon nanomaterials in atherosclerosis. *Biotechnol Adv* 2018; 36: 2232-2247.
- 173) Van-Solingen C, De-Boer HC, Bijkerk R, Monge M, Van-Oeveren-Rietdijk AM, Seghers L, DeVries MR, Van-Der-Veer EP, Quax PH, Rabelink TJ, Van-Zonneveld AJ. MicroRNA-126 modulates endothelial SDF-1 expression and mobilization of Sca-1(+)/Lin(-) progenitor cells in ischaemia. *Cardiovasc Res* 2011; 92: 449-455.
- 174) Evans WS, Sapp RM, Kim KI, Heilman JM, Hagberg J, Prior SJ. Effects of Exercise Training on the Paracrine Function of Circulating Angiogenic Cells. *Int J Sports Med* 2021; 42: 1047-1057.
- 175) Bassand K, Metzinger L, Naim M, Mouhoubi N, Haddad O, Assoun V, Zaidi N, Sainte-Catherine O, Butt A, Guyot E, Oudar O, Laguillier-Morizot C, Sutton A, Charnaux N, Metzinger-Le MV, Hlawaty H. miR-126-3p is essential for CXCL12-induced angiogenesis. *J Cell Mol Med* 2021; 25: 6032-6045.
- 176) Li J, Chen J, Zhang F, Li J, An S, Cheng M, Li J. LncRNA CDKN2B-AS1 hinders the proliferation and facilitates apoptosis of ox-LDL-induced vascular smooth muscle cells via the ceRNA network of CDKN2B-AS1/miR-126-5p/PTPN7. *Int J Cardiol* 2021; 340: 79-87.
- 177) Zhu J, Liu B, Wang Z, Wang D, Ni H, Zhang L, Wang Y. Exosomes from nicotine-stimulated macrophages accelerate atherosclerosis through miR-21-3p/PTEN-mediated VSMC migration and proliferation. *Theranostics* 2019; 9: 6901-6919.
- 178) Ouyang S, Yao YH, Zhang ZM, Liu JS, Xiang H. Curcumin inhibits hypoxia inducible factor-1alpha-induced inflammation and apoptosis in macrophages through an ERK dependent pathway. *Eur Rev Med Pharmacol Sci* 2019; 23: 1816-1825.
- 179) Celletti FL, Waugh JM, Amabile PG, Brendolan A, Hilfiker PR, Dake MD. Vascular endothelial growth factor enhances atherosclerotic plaque progression. *Nat Med* 2001; 7: 425-429.
- 180) Sun L, Zhou H, Yang Y, Chen J, Wang Y, She M, Li C. Meta-analysis of diagnostic and prognostic value of miR-126 in non-small cell lung cancer. *Biosci Rep* 2020; 40: 89.
- 181) Tafsiri E, Darbouy M, Shadmehr MB, Zagryazhskaya A, Alizadeh J, Karimipour M. Expression of miRNAs in non-small-cell lung carcinomas and their association with clinicopathological features. *Tumour Biol* 2015; 36: 1603-1612.
- 182) Zhou W, Nie J, Zhang D. [Differential expression of miR-126-5p in lung adenocarcinoma and the possible mechanism]. *Nan Fang Yi Ke Da Xue Xue Bao* 2019; 39: 1186-1190.

- 183) Lv J, Zhu S, Chen H, Xu Y, Su Q, Yu G, Ma W. Paeonol inhibits human lung cancer cell viability and metastasis in vitro via miR-126-5p/ZEB2 axis. *Drug Dev Res* 2022; 83: 432-446.
- 184) Liu W, Zhang Y, Huang F, Ma Q, Li C, Liu S, Liang Y, Shi L, Yao Y. The Polymorphism and Expression of EGFL7 and miR-126 Are Associated With NSCLC Susceptibility. *Front Oncol* 2022; 12: 772405.
- 185) Xiang G, Cheng Y. MiR-126-3p inhibits ovarian cancer proliferation and invasion via targeting PLXNB2. *Reprod Biol* 2018; 18: 218-224.
- 186) Yin J, Bai Z, Song J, Yang Y, Wang J, Han W, Zhang J, Meng H, Ma X, Yang Y, Wang T, Li W, Zhang Z. Differential expression of serum miR-126, miR-141 and miR-21 as novel biomarkers for early detection of liver metastasis in colorectal cancer. *Chin J Cancer Res* 2014; 26: 95-103.
- 187) Li Q, Wang G, Wang H. miR-126 Functions as a Tumor Suppressor by Targeting SRPK1 in Human Gastric Cancer. *Oncol Res* 2018; 26: 1345-1353.
- 188) Bao J, Yu Y, Chen J, He Y, Chen X, Ren Z, Xue C, Liu L, Hu Q, Li J, Cui G, Sun R. MiR-126 negatively regulates PLK-4 to impact the development of hepatocellular carcinoma via ATR/CHEK1 pathway. *Cell Death Dis* 2018; 9: 1045.
- 189) Liu R, Gu J, Jiang P, Zheng Y, Liu X, Jiang X, Huang E, Xiong S, Xu F, Liu G, Ge D, Chu Y. DNMT1-microRNA126 epigenetic circuit contributes to esophageal squamous cell carcinoma growth via ADAM9-EGFR-AKT signaling. *Clin Cancer Res* 2015; 21: 854-863.
- 190) Al-Kafaji G, Said HM, Alam MA, Al NZ. Blood-based microRNAs as diagnostic biomarkers to discriminate localized prostate cancer from benign prostatic hyperplasia and allow cancer-risk stratification. *Oncol Lett* 2018; 16: 1357-1365.
- 191) Chen SR, Cai WP, Dai XJ, Guo AS, Chen HP, Lin GS, Lin RS. Research on miR-126 in glioma targeted regulation of PTEN/PI3K/Akt and MDM2-p53 pathways. *Eur Rev Med Pharmacol Sci* 2019; 23: 3461-3470.
- 192) Soofiyani SR, Hosseini K, Ebrahimi T, Frohander H, Sadeghi M, Beirami SM, Ghasemnejad T, Tarhiz V, Montazersaheb S. Prognostic Value and Biological Role of miR-126 in Breast Cancer. *Microna* 2022 Apr 28. doi: 10.2174/1876402914666220428123203. Epub ahead of print.
- 193) Sibilano M, Tullio V, Adorno G, Savini I, Gasperi V, Catani MV. Platelet-Derived miR-126-3p Directly Targets AKT2 and Exerts Anti-Tumor Effects in Breast Cancer Cells: Further Insights in Platelet-Cancer Interplay. *Int J Mol Sci* 2022; 23: 5484.
- 194) Sharma AR, Sharma G, Bhattacharya M, Lee SS, Chakraborty C. Circulating miRNA in Atherosclerosis: A Clinical Biomarker and Early Diagnostic Tool. *Curr Mol Med* 2022; 22: 250-262.
- 195) Gao J, Yang S, Wang K, Zhong Q, Ma A, Pan X. Plasma miR-126 and miR-143 as Potential Novel Biomarkers for Cerebral Atherosclerosis. *J Stroke Cerebrovasc Dis* 2019; 28: 38-43.
- 196) Li HY, Zhao X, Liu YZ, Meng Z, Wang D, Yang F, Shi QW. Plasma MicroRNA-126-5p is Associated with the Complexity and Severity of Coronary Artery Disease in Patients with Stable Angina Pectoris. *Cell Physiol Biochem* 2016; 39: 837-846.
- 197) Xue S, Liu D, Zhu W, Su Z, Zhang L, Zhou C, Li P. Circulating MiR-17-5p, MiR-126-5p and MiR-145-3p Are Novel Biomarkers for Diagnosis of Acute Myocardial Infarction. *Front Physiol* 2019; 10: 123.
- 198) Weale CJ, Matshazi DM, Davids S, Raghubeer S, Erasmus RT, Kengne AP, Davison GM, Matsha TE. MicroRNAs-1299, -126-3p and -30e-3p as Potential Diagnostic Biomarkers for Prediabetes. *Diagnostics (Basel)* 2021; 11: 949.