

Preclinical pharmacokinetic study of a novel lipid-lowering agent, IMM-H007

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Abstract. – OBJECTIVE: To investigate the pharmacokinetic characteristics of absorption, distribution, metabolism, and excretion *in vivo* after oral administration and sublingual venous injection of the small molecule IMM-H007 in hamsters.

MATERIALS AND METHODS: Pharmacokinetic characteristics, including absorption, distribution, metabolism, and excretion, were studied *in vivo* by LC-MS/MS after oral administration and sublingual venous injection of IMM-H007 in hamsters. Furthermore, IMM-H007 stability in artificial gastric juices, artificial intestinal juices, and Tris-HCl buffer was also analyzed.

RESULTS: There was no significant matrix or impurity interference in golden hamster whole blood as shown using MS/MS to detect the existence of these substances. IMM-H007, M1, and MP exhibited good linearity in the range of 1-500 ng/mL, 2-1000 ng/mL, and 2-1000 ng/mL, respectively. The matrix effect was 105.49%, and IMM-H007, M1, and MP were stable during the process of sample digestion. These results illustrate that the HPLC-MS/MS analytical method is simple, rapid, and sensitive and exhibits high specificity and accuracy, which meets the clinical pharmacokinetic requirements of IMM-H007. IMM-H007 was rapidly absorbed through the oral route in hamsters. The C_{max} and $AUC_{(0-t)}$ of the M1 and MP metabolites in male and female hamsters were increased with increasing dosage and were proportional to the dose. In addition, $T_{1/2}$ and $T_{1/2\beta}$ were significantly prolonged with increasing dosage, exhibiting linear dynamic characteristics and no significant gender differences. Bioavailability in male and female golden hamsters after oral administration of IMM-H007 was calculated using the sum of M1 and MP, resulting in 6.97% and 8.95%, respectively. IMM-H007 and its metabolites were stable in Tris-HCl buffer, artificial gastric juices, and artificial intestinal juices.

CONCLUSIONS: This study provides an experimental basis for elucidating the material pharmacodynamic functions of IMM-H007 and predicting its potential drug interactions.

Keywords: IMM-H007, Preclinical pharmacokinetic, Linear dynamic characteristics

Introduction

The social economy has conveyed great changes in people's life style. Particularly with the ageing population and the acceleration of urbanization, the epidemic trend of risk factors for vascular diseases in China is significant¹. Currently, cardiovascular mortality accounts for the leading cause of overall death among both urban and rural residents. Furthermore, the burden of cardiovascular disease has become an increasingly major public health problem²⁻⁴. IMM-H007 is a novel AMP-activated protein kinase (AMPK) small molecule agonist that specifically binds to AMPK α subunits to activate AMPK. IMM-H007 is a novel type of lipid-lowering compound, and much research⁵⁻⁸ has focused on the experimental study of IMM-H007. Pharmacological studies have shown that IMM-H007 inhibits fatty lesions induced by octadecenoic acid and accumulation of lipids in HepG2 cells⁹. Researchers¹⁰⁻¹² on the *in vivo* pharmacodynamics found that IMM-H007 reduces the concentration of serum triglycerides, total cholesterol, and low-density lipoproteins in golden hamsters with hyperlipidaemia. Mechanistic studies¹³ suggest that IMM-H007 is a new type of AMP-activated protein kinase activator.

In addition, preoperative pharmacokinetic studies have shown that IMM-H007 is a novel lipid-lowering compound with a distinct chemical structure, targets, and metabolic pathways from statins and is expected to be a new drug for the prevention and treatment of cardiovascular disease¹⁴. The pharmacokinetic characteristics of absorption, distribution, metabolism, and excretion *in vivo* after oral administration and sublingual venous injection of IMM-H007 in hamsters were investigated herein using LC-MS/MS. Our findings provide an experimental basis for elucidating the pharmacodynamics activity and predicting potential drug interactions.

Materials and Methods

Drugs and Reagents

IMM-H007 (Batch No. 20120401, 99.7% purity), M1, MP, and the internal standard WS070119 were supplied by the Institute of Medicine of the Chinese Academy of Medical Sciences (Beijing, China). Formic acid and heparin sodium were purchased from Sigma-Aldrich (St. Louis, MO, USA). Sodium carboxy methyl cellulose (CMC), and sodium fluoride were purchased from Sinopharm Chemical Reagent Co., Ltd. (Shanghai, China). Methanol, acetonitrile (chromatographic pure, Merck, Darmstadt, Germany), and other reagents were of analytical grade. Ultra-pure water was sourced from a Milli-Q water instrument (Millipore, Billerica, MA, USA).

Experimental Apparatus

Sartorius-precision weighing were from Sartorius (Goettingen, Germany). Saiguo ultrasound equipment was from Saiguo (Osaka, Japan). The GL-88B combi mixer was from Nanjing Jiancheng Tech (Nanjing, China). The 16B centrifuge was from Shanghai Anting Scientific Instrument Factory (Shanghai, China). The Thermo-Biofuge-PrimoR temperature centrifuge and LC-MS/MS liquid chromatography-mass spectrometry analyzer were from Thermo-Fisher (Waltham, MA, USA) and included the following: surveyor autosampler, syringe for liquid, SQ Quantum Access™ Triple quadrupole Mass Spectrometer, Electrospray Ionization (ESI), and X calibur 1.4 software (used for data collection and analysis).

Experimental Animals

Experimental hamsters were 6-8 weeks old and weighed 100-120 g. Animals were randomly

grouped for experiments. Hamsters were fed in a plastic box and with standard specific pathogen-free (SPF) feed and were given free access to water. Room temperature was maintained at 20-25°C and 50-60% humidity. Animals were kept on a 12 h light/dark cycle, and ventilation frequency was 10-20 times/h. The air velocity was 0.1-0.2 m/s, and the air cleanliness was >10,000. The ammonia concentration was ≤ 1 mg/m³, and the noise level was ≤ 60 decibels. Illumination was 150-300 lux and natural illumination was 15-20 LX. The rat was washed twice per week, and cages were sterilized with high pressure after cleaning. This study was approved by the Animal Ethics Committee of China Pharmaceutical University Animal Center.

Experimental Methods

(1) Chromatographic conditions included use of the following: reproSil 120 C18-AQ; mobile phase: A (0.1% formic acid water), B (0.1% formic acid methanol); flow rate: 0.2 mL/min; column temperature: 30°C; injection volume: 5 μ L; and injection temperature: 10°C.

(2) Mass spectrometry conditions included the following: electrospray ion source, 3700 V spray voltage, the sheath gas and auxiliary gas were nitrogen, the pressure and air flow rate were 30 psi and 10 L/min; the capillary temperature was 350°C; and the collision gas pressure was 1.5 mTorr.

(3) Standard product preparation included accurate weighing of standard products. Acetonitrile was added (10 mL) to a volumetric flask that was the corresponding concentration of the reserving solution. Next, the reserving solution was precisely drawn. Acetonitrile was then diluted into a standard solution of suitable concentration by means of gradient dilution.

(4) The matrix effect value was calculated by comparing the peak area ratios of whole blood samples and working fluids of IMM-H007, M1, and MP QC samples with low, medium, and high concentrations.

(5) To determine stability, five samples were taken from each IMM-H007, M1, and MP QC sample with low, medium, and high concentrations. The stability of IMM-H007, M1, and MP was determined by placing the sample concentrations in an auto sampler for 24 h. Before investigation, samples were placed in an ice bath for 1 h, subjected to repeated freezing and thawing (three times), as well as short-term placed and long-term placed.

Table I. Regression equations of IMM-H007, M1 and MP in hamster blood and liver.

Sample type	Substance	C(ng/mL)	Equation	R ²	LLOQ
Blood	IMM-H007	1-500	Y=0.0123654X+0.0154658	0.9991	1
	M1	2-1000	Y=0.0098745X+0.0162354	0.9980	2
	MP	10-5000	Y=0.0046542X+0.00123549	0.9991	10
Liver	IMM-H007	1-500	Y=0.0135648X+0.0178946	0.9996	1
	M1	2-1000	Y=0.0124685X+0.0156987	0.9985	2
	MP	10-5000	Y=0.00329307X-0.00055849	0.9993	10

Results

The Linear Relationship between Prototype Drug IMM-H007 and Metabolites M1 and MP in Whole Blood and Tissue Samples

The prototype drug IMM-H007 and its metabolites M1 and MP exhibited a good linear relationship in whole blood and tissue samples from hamsters at concentration ranges of 1-500, 2-1000, and 10-5000 ng/mL, respectively. The correlation coefficient of standard curves (r^2) was greater than 0.99. The minimum limit of quantification (LLQQ) for these three groups was 1, 2, and 10 ng/mL, respectively, which met the requirements of sensitivity administration of whole blood samples after hamsters were administered the prototype drug. Regression equations for the prototype drug IMM-H007 and its metabolites M1 and MP in biological sample are shown in Table I.

The Linear Relationship between the Prototype Drug IMM-H007 and Metabolites M1 and MP in Faeces and Urine Samples

M228, M244, M344, M404, M440, and M536 in faeces and urine samples from hamsters exhibited good linear relationship at concentration ranges of

0.5-50, 1-200, 0.01-0.5, 0.01-0.5, 0.01-0.5, and 0.01-20 µg/mL, respectively. The correlation coefficient of the standard curves (r^2) was greater than 0.99. The minimum limit of quantification (LLQQ) was 0.5, 1, 0.01, 0.2, 0.5, and 10 µg/mL, respectively, meeting the requirements of sensitivity administration of metabolites in faeces and urine samples after administration of the prototype drug in hamsters. Regression equations for M228, M244, M344, M404, M440, and M536 in biological sample are shown in Table II.

Matrix Effects and Stability

The absolute matrix effect values of whole blood samples, low, medium, and high quality control concentrations (2, 50, and 200 ng/mL) of IMM-H007, M1, and MP are shown in Table III. The results show that the method has no effect on determination of the metabolites in matrix effect, suggesting that the method is accurate and reliable. Samples with low, medium, and high quality control concentrations (IMM-H007: 2, 50, and 200 ng/mL; M1: 5, 100, and 500 ng/mL; MP: 20, 500, and 2000 ng/mL) were placed for 1 h in an ice bath, subjected to repeated freeze-thaw cycles (three times), placed at -80°C for 30 days, and were tested for stability by placement in an auto sampler for 24 h. The results are shown in Table IV and indicate that IMM-H007, M1, and MP remained stable during the above-mentioned treatments.

Table II. Regression equations of M228, M244, M344, M404, M440 and M536 in urine and feces samples.

Metabolites	C (µg/mL)	Equation	R ²	LLOQ (µg/mL)
M228	0.5-50	Urine: Y=0.01834592X+0.00385974	0.9991	0.5
		Feces: Y=0.0160358X+0.00446871	0.9980	
M244	1-200	Urine: Y=0.00465823X+0.0492689	0.9980	1
		Feces: Y=0.0579846X+0.0196438	0.9983	
M344	0.01-0.5	Urine: Y=0.0062938X-0.00062851	0.9999	0.01
		Feces: Y=0.00171527X-0.000302987	0.9996	
M404	0.2-20	Urine: Y=0.0729184X+0.0283431	0.9996	0.2
		Feces: Y=0.475842X+0.0117105	0.9989	
M440	0.5-50	Urine: Y=0.029874X+0.00390267	0.9998	0.5
		Feces: Y=0.29687X+0.0024684	0.9998	
M536	0.2-20	Urine: Y=0.009534581+0.0204985X	0.9990	0.2
		Feces: Y=0.01264587X+0.0150378	0.9978	

Table III. Matrix effects of IMM-H007, M1 and MP in hamster blood (n=5).

Sample	C (ng/mL)	Peak area of matrix sample	Peak area of standard sample	Matrix effect	Precision (RSD%)
IMM-H007	2	60896±5623	84654±1265	85.89	8
	50	1365894±89561	1645612±7543	85.89	7
	200	5365489±198542	6356412±12345	84.41	4
M1	2	81235±4652	94584±8456	85.89	4
	50	1592548±400254	1874562±598742	84.95	2
	200	7845652±295641	8756412±184568	89.21	4
MP	2	61203±2365	72365±8546	84.58	4
	50	1985641±395461	89754±568942	110.94	21
	200	7056945±298543	89745±798456	105.49	5

Table IV. Stability of IMM-H007, M1 and MP in hamster blood under different conditions (n=5).

Sample	C (ng/mL)	Ice bath for 1 h		Free freeze-thaw cycles		Store at -80°C (30 days)		Autosampler at 10°C (24 h)	
		Measured (Mean±SD)	Accuracy (R.E.%)	Measured (Mean±SD)	Accuracy (R.E.%)	Measured (Mean±SD)	Accuracy (R.E.%)	Measured (Mean±SD)	Accuracy (R.E.%)
IMM-H007	2	1.898±0.301	0.6	2.156±0.015	7.1	2.179±0.165	9.2	2.23±0.21	12.5
	50	52.98±1.26	6.54	52.12±1.04	2.1	47.8±3.94	-4.2	44.2±1.3	-11.7
	200	221.6±3.7	10.2	217.6±3.6	9.4	182.5±2.6	-8.5	184.0±6.8	-8.2
M1	5	4.962±0.5	9.9	5.03±0.384	0.4	4.65±0.4	-6.2	5.0±0.41	-0.2
	100	108.89±2.45	9.1	108.56±3.89	9.1	118.26±3.59	11.56	106.14±1.49	6.0
	500	522.6±7.2	4.5	528.6±7.7	5.6	514.1±8.1	2.8	558.2±10.9	12.4
MP	20	21.87±1.23	7.1	21.87±1.09	9.2	18.45±1.02	-8.4	18.5±1.02	-9.4
	500	554.89±3.20	13.26	554.26±16.48	10.84	456.59±35.26	-8.2	567.9±8.26	13.7
	2000	2165.1±25.4	8.6	2165.1±26.4	8.2	2094.2±19.6	4.2	2189.2±20.8	9.8

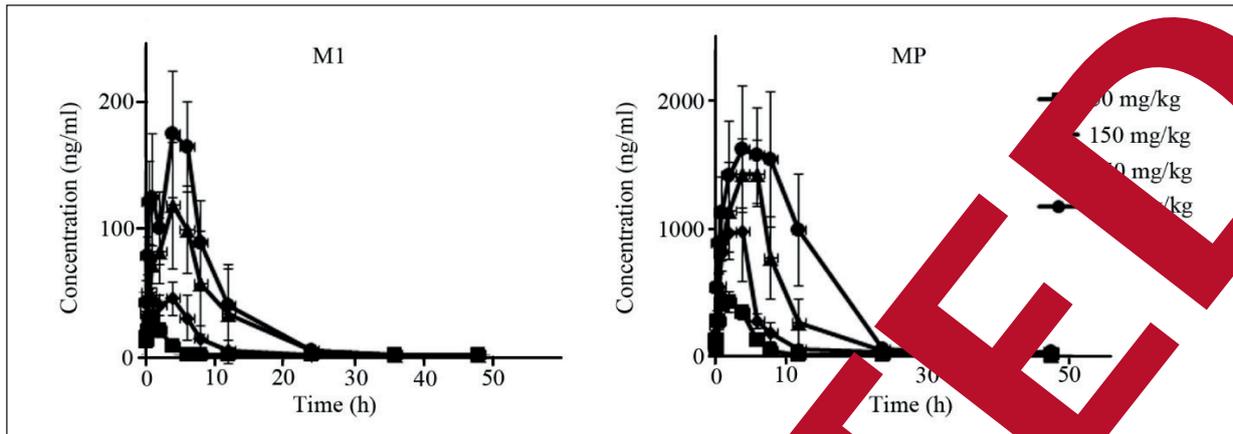


Figure 1. Mean blood concentration-time profiles of M1 and MP in male hamsters after oral IMM-H007 administration at doses of 50, 150, 450, and 900 mg/kg (n=5).

Pharmacokinetics Study of Oral and Intravenous Administration of IMM-H007 in Hamsters

Pharmacokinetics of Oral Administration of IMM-H007 in Hamsters

After hamsters were treated with different doses of IMM-H007 (50, 150, 450, and 900 mg/kg), the concentration-time curve of whole blood concentrations was plotted (Figure 1 and Figure 2). According to the concentration-time curve for M1 and MP after oral administration of different doses of IMM-H007 (50, 150, 450, and 900 mg/kg) in male and female hamsters, the M1 and MP pharmacokinetic parameters were calculated using the non-compartmental model of Phoenix WinNonLin software. Relevant pharmacokinetic parameters are shown in Tables 1 and 2.

The relationship between the average peak concentration (C_{max} , AUC₀₋₂₄) of M1 and MP, and the dosage is shown in Figure 3. Hamsters rapidly absorbed IMM-H007 after oral administration, and the metabolites M1 and MP could be detected in blood 3 minutes after administration in each dose group. Twelve to thirty-six hours after administration, the plasma concentration of the metabolic products M1 and MP was below the detection limit. The concentration of the drug prototype at each time point was close to or below the detection limit.

After oral administration of different doses of IMM-H007 (50, 150, 450, and 900 mg/kg) to male hamsters, C_{max} of the M1 metabolite was 35.78 ± 10.82 , 82.65 ± 28.15 , 123.46 ± 30.87 , and 209.46 ± 59.87 ng/mL, while $T_{1/2}$ was 1.38 ± 0.06 , 4.36 ± 1.18 , 6.19 ± 0.67 , and 3.66 ± 0.87 h,

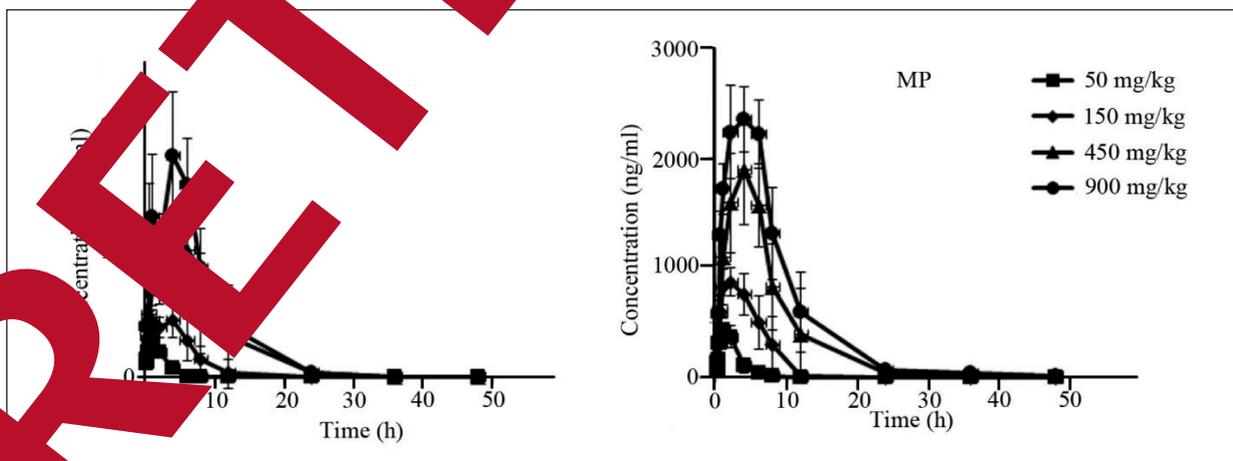


Figure 2. Mean blood concentration-time profiles of M1 and MP in female hamsters after oral IMM-H007 administration at doses of 50, 150, 450, and 900 mg/kg (n=5).

Table V. Pharmacokinetic parameters of M1 in male hamsters after oral administration of IMM-H007 at 50, 150, 450, 900 mg/kg (n=5).

Parameters	50 mg/kg	150 mg/kg	450 mg/kg	900 mg/kg
$T_{1/2}$ (h)	1.38±0.06	4.36±1.18	6.19±0.67	3.66±0.81
T_{max} (h)	1.02±0.84	2.39±1.68	4.79±0.99	5.91±3.22
C_{max} (ng/mL)	35.78±10.82	82.65±28.15	123.46±30.87	209.46±59.87
$AUC_{(0-t)}$ (h*ng/mL)	143.58±12.09	402.69±120.39	982.81±236.42	2340.66±856.14
$AUC_{(0-\infty)}$ (h*ng/mL)	149.74±13.05	420.36±120.69	996.57±230.14	2376.36±798.38
V_z (L/kg)	0.87±0.06	2.04±0.82	4.21±10.6	2.15±0.48
CL _z (L/h/kg)	0.36±0.04	0.38±0.10	0.45±0.10	0.43±0.07
$MRT_{(0-t)}$ (h)	2.86±0.22	4.19±0.68	6.76±0.97	7.59±0.99
$MRT_{(0-\infty)}$ (h)	3.07±0.19	4.47±0.79	7.04±0.69	7.11±0.81

Table VI. Pharmacokinetic parameters of MP in male hamsters after oral administration of IMM-H007 at 50, 150, 450, 900 mg/kg (n=5).

Parameters	50 mg/kg	150 mg/kg	450 mg/kg	900 mg/kg
$T_{1/2}$ (h)	1.42±0.64	5.16±1.08	4.06±0.65	4.47±0.84
T_{max} (h)	1.87±0.56	2.71±1.28	4.79±1.32	4.78±2.98
C_{max} (ng/mL)	479.23±58.24	1123.48±279.36	1579.64±209.67	1865.37±459.87
$AUC_{(0-t)}$ (h*ng/mL)	2130.46±436.52	5897.61±1024.16	13249.87±2360.9	23048.97±6983.27
$AUC_{(0-\infty)}$ (h*ng/mL)	2197.64±504.93	5798.46±1023.97	12997.64±2309.2	23129.83±6974.38
V_z (L/kg)	0.06±0.01	0.20±0.08	0.28±0.07	0.30±0.08
CL _z (L/h/kg)	0.03±0.01	0.03±0.00	0.03±0.01	0.05±0.01
$MRT_{(0-t)}$ (h)	2.86±0.46	5.17±0.81	7.19±0.91	8.16±0.89
$MRT_{(0-\infty)}$ (h)	3.27±1.19	5.77±0.81	7.56±0.78	8.32±0.94

Table VII . Pharmacokinetic parameters of M1 in female hamsters after oral administration of IMM-H007 at 50, 150, 450, 900 mg/kg (n=5).

Parameters	50 mg/kg	150 mg/kg	450 mg/kg	900 mg/kg
$T_{1/2}$ (h)	1.36±0.24	1.94±0.37	2.56±0.69	3.76±0.43
T_{max} (h)	0.99±0.19	2.56±1.61	4.86±1.99	4.65±0.92
C_{max} (ng/mL)	30.67±10.77	79.17±28.77	130.46±39.12	187.34±51.68
$AUC_{(0-t)}$ (h*ng/mL)	153.46±17.85	300.04±109.37	1033.32±498.67	1587.64±423.15
$AUC_{(0-\infty)}$ (h*ng/mL)	83.67±17.85	320.97±100.34	1067.85±485.37	1597.64±428.37
V_z (L/kg)	1.17±0.36	1.39±0.46	1.89±0.97	3.27±0.79
CL _z (L/h/kg)	0.63±0.13	0.53±0.10	0.51±0.22	0.58±0.21
$MRT_{(0-t)}$ (h)	2.86±0.28	3.68±0.78	5.78±1.83	6.57±0.99
$MRT_{(0-\infty)}$ (h)	2.57±0.28	4.47±0.72	6.07±1.67	6.67±1.03

Table VIII. Pharmacokinetic parameters of MP in female hamsters after oral administration of IMM-H007 at 50, 150, 450, 900 mg/kg (n=5).

Parameters	50 mg/kg	150 mg/kg	450 mg/kg	900 mg/kg
$T_{1/2}$ (h)	1.20±0.09	3.56±0.67	6.73±2.34	9.67±2.43
T_{max} (h)	0.99±0.09	2.34±1.11	4.45±1.57	4.32±1.94
C_{max} (ng/mL)	426.39±73.98	912.31±150.12	1978.46±379.52	2546.31±364.35
$AUC_{(0-t)}$ (h*ng/mL)	1304.65±286.36	5963.46±1876.45	16435.23±3765.48	24653.46±4987.56
$AUC_{(0-\infty)}$ (h*ng/mL)	1334.89±267.48	6045.31±1879.46	16423.87±3970.46	24361.21±4974.2
V_z (L/kg)	0.06±0.02	0.16±0.04	0.26±0.07	0.56±0.29
CL _z (L/h/kg)	0.04±0.03	0.03±0.01	0.03±0.01	0.04±0.02
$MRT_{(0-t)}$ (h)	2.16±0.22	4.67±0.56	6.75±1.34	7.27±0.68
$MRT_{(0-\infty)}$ (h)	2.30±0.25	4.98±0.32	7.21±1.27	7.68±0.64

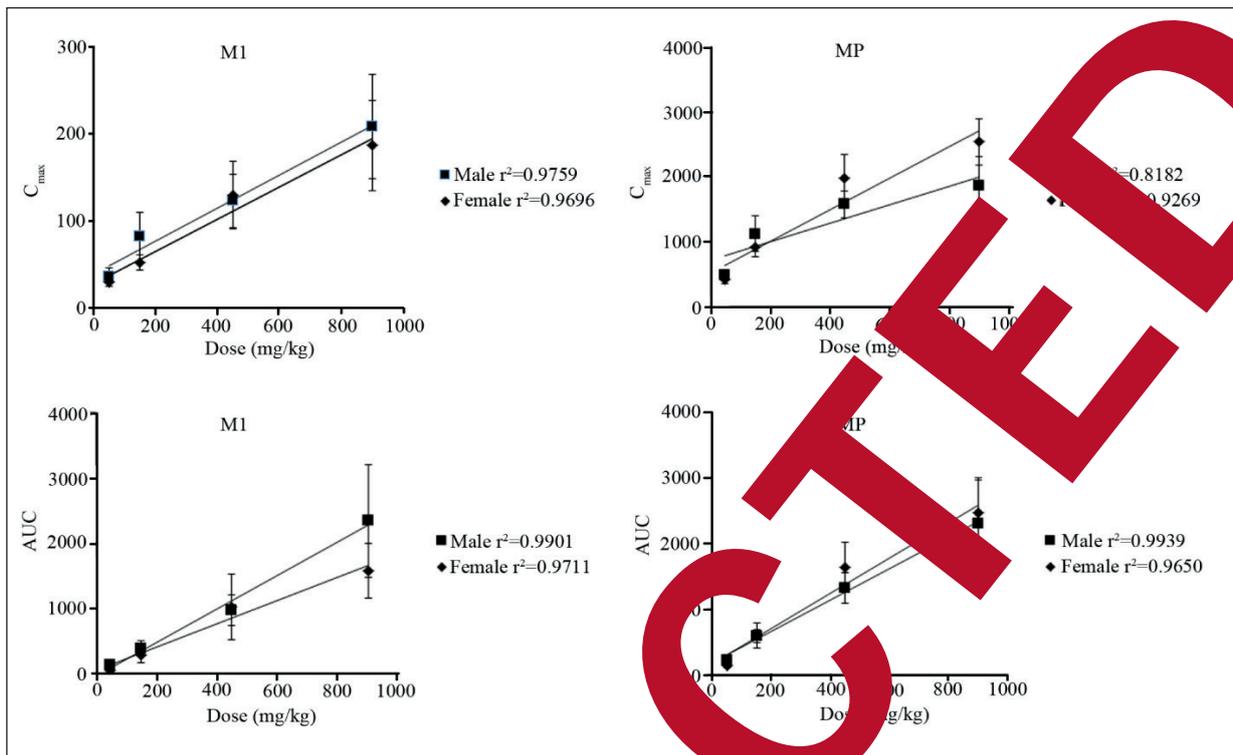


Figure 3. Relationship between Dose (mg/kg) and C_{max} and AUC(0-t) of M1 and MP.

respectively. AUC_(0-t) was 143.58 ± 12.09, 479.19 ± 120.39, 982.81 ± 236.42, and 2345.64 ± 851.12 h*ng/mL, while MRT_(0-t) was 2.86 ± 0.22, 4.11 ± 0.68, 6.76 ± 0.97, and 7.64 ± 0.97 h, respectively. C_{max} of the MP metabolite was 479.19 ± 58.24, 1123.48 ± 279.36, 1579.64 ± 209.67 and 1865.37 ± 459.87 ng/mL; T_{1/2} was 0.64 ± 0.12, 0.72 ± 0.65, and 4.96 ± 0.84 h, respectively. AUC_(0-t) was 213.60 ± 436.52, 5897.61 ± 124.16, 1579.64 ± 2360.98, and 23048.97 ± 691.27 h*Ng/mL. MRT_(0-t) was 2.86 ± 0.46, 4.07, 7.19 ± 0.97 and 8.16 ± 0.89 h, respectively. C_{max} and AUC_(0-t) of the M1 and MP metabolites in male hamsters after IMM-H007 oral administration. IMM-H007 increased with increasing dose, MRT_(0-t) increased slightly with increasing dose, and in the 50-900 mg/kg dose range, the M1 and MP metabolites both exhibited linear kinetic process *in vivo*. After oral administration of different IMM-H007 doses (150, 450, and 900 mg/kg) to male hamsters, the C_{max} of the M1 metabolite was 143.58 ± 5.82, 52.46 ± 8.37, 130.46 ± 39.12, and 187.19 ± 37.12 ng/mL, while T_{1/2} was 1.36 ± 0.24, 1.36 ± 0.37, 1.36 ± 0.69, and 3.76 ± 0.43 h, respectively. AUC_(0-t) was 73.46 ± 16.37, 300.04 ± 109.37, 982.81 ± 498.67, and 1587.64 ± 423.15 h*ng/mL, while MRT_(0-t) was 1.67 ± 0.28, 3.68 ± 0.78, 5.78 ± 1.83,

and 6.76 ± 0.99 h. C_{max} of the MP metabolite was 426.39 ± 73.98, 912.31 ± 150.12, 1978.46 ± 379.52, and 2546.31 ± 364.35 ng/mL, while T_{1/2} was 1.20 ± 0.09, 3.56 ± 0.67, 6.73 ± 2.34, and 9.67 ± 2.43 h, respectively. AUC_(0-t) was 1304.65 ± 286.36, 5963.46 ± 1876.45, 16435.23 ± 3765.48, and 24653.46 ± 4987.56 h*ng/mL, while MRT_(0-t) was 2.16 ± 0.22, 4.67 ± 0.56, 6.75 ± 1.34, and 7.27 ± 0.68 h. The C_{max} and AUC_(0-t) of the M1 and MP metabolites in female hamsters after oral IMM-H007 administration increased with increasing dose, and MRT_(0-t) increased slightly with increasing dose. In the 50-900 mg/kg dose range, the M1 and MP metabolites exhibited a linear kinetic process *in vivo*. There was no significant gender difference.

Changes in Blood Concentration and the Drug-Time Curve after Intravenous IMM-H007 Injection at Different Time Points in Hamsters

The drug-time curve of whole blood at each time point in golden hamsters after intravenous IMM-H007 injection (5 mg/kg) is shown in Figure 4. According to the drug-time curve of M1 and MP metabolites after IMM-H007 sublingual venous injection (5 mg/kg) in male and female golden hamsters, M1 and MP pharmacokinetic param-

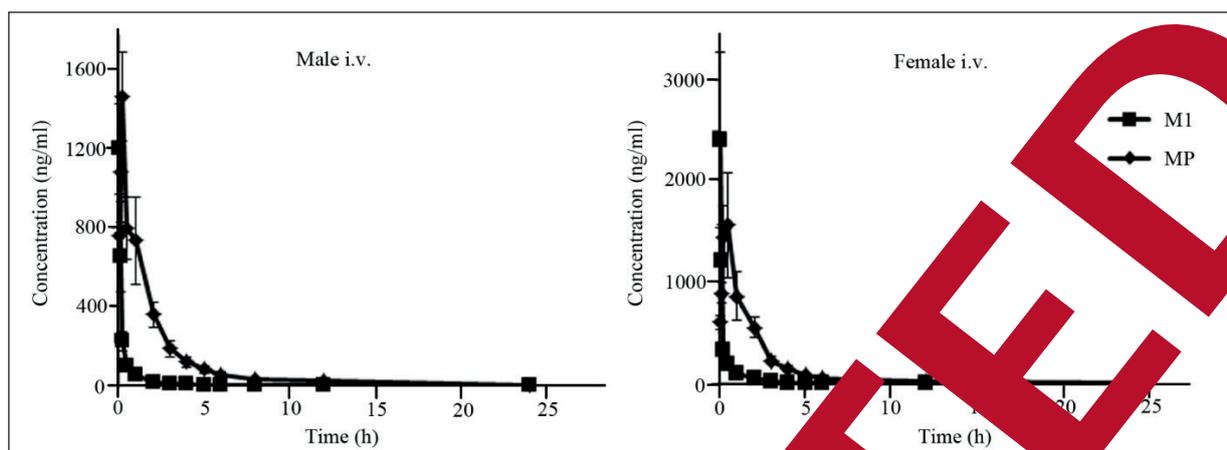


Figure 4. Mean blood concentration-time profiles of M1 and MP in male and female golden hamsters after a single i.v. administration of IMM-H007 at 5 mg/kg (n=5).

Parameters were calculated using the non-compartmental model of Phoenix WinNonLin software. Relevant pharmacokinetic parameters are shown in Table IX. After IMM-H007 injection into the sublingual vein of male and female golden hamsters, the drug prototype and M1 and MP metabolites were detected in the blood at two minutes. Five minutes after administration, the drug prototype concentration was close to the detection limit. Twelve to twenty-five hours after administration, M1 and MP metabolites were close to the detection limit. Five minutes after administration in male hamsters, the average blood concentration of the drug prototype was 7.58 ng/mL, the C_{max} of M1 and MP metabolites was 1198.69 and 1465.23 ng/mL, $T_{1/2}$ was 2.11 and 2.33 h, $AUC_{(0-24)}$ was 332.26 and 2274.35 ng·h/mL, and $MRT_{(0-24)}$ was 1.09 and 2.08 h, respectively. Five minutes after administration in female hamsters, the average blood concentration of the drug prototype was 7.58 ng/mL, the C_{max} of M1 and MP metabolites

was 2406.58 and 1569.37 ng/mL, $T_{1/2}$ was 2.11 and 2.34 h, $AUC_{(0-24)}$ was 569.29 and 2874.66 ng·h/mL, and $MRT_{(0-24)}$ was 0.79 and 1.97 h, respectively. The bioavailability of IMM-H007 after oral administration of IMM-H007 (50 mg/kg) in male golden hamsters for M1 and MP metabolites was 6.97%, and the bioavailability in female golden hamsters was 8.95%. Calculation of the area under the drug curve of the M1 and MP metabolites in whole blood to determine the oral bioavailability of IMM-H007 is shown below.

$$F_{male,M1}(\%) = \frac{5mg/kg \times 73.06ng/ml \cdot h}{50mg/kg \times 332.47ng/ml \cdot h} = 2.19\%$$

$$F_{female,M1}(\%) = \frac{5mg/kg \times 132.08ng/ml \cdot h}{50mg/kg \times 600.02ng/ml \cdot h} = 2.20\%$$

$$F_{male,MP}(\%) = \frac{5mg/kg \times 1079.58ng/ml \cdot h}{50mg/kg \times 2258.71ng/ml \cdot h} = 4.78\%$$

$$F_{female,MP}(\%) = \frac{5mg/kg \times 1901.83ng/ml \cdot h}{50mg/kg \times 2819.45ng/ml \cdot h} = 6.75\%$$

Table IX. Pharmacokinetic parameters of M1 and MP in male and female hamster after a single i.v. administration of IMM-H007 at 5 mg/kg (n=5).

Parameters	Male		Female	
	M1	MP	M1	MP
$T_{1/2}$ (h)	2.11±0.56	2.33±0.26	2.11±0.12	2.34±0.18
$T_{1/2\beta}$ (h)	0.03±0.00	0.25±0.00	0.03±0.00	0.35±0.03
C_{max} (ng/mL)	1198.69±223.65	1465.23±256.58	2406.58±896.25	1569.37±489.56
$AUC_{(0-24)}$ (h*ng/mL)	332.26±55.25	2274.35±336.21	569.29±198.55	2874.66±546.21
$AUC_{(0-5)}$ (h*ng/mL)	339.65±55.26	2345.61±336.78	609.58±199.25	2985±523.86
Cl_z (L/h/kg)	44.05±6.89	0.01±0.00	22.89±8.12	5.79±1.52
Cl_z (L/h/kg)	15.03±2.21	0.00±0.00	8.49±2.56	1.79±0.36
$MRT_{(0-24)}$ (h)	1.09±0.22	2.08±0.06	0.79±0.12	1.97±0.15
$MRT_{(0-5)}$ (h)	1.48±0.36	2.65±0.15	0.97±0.23	2.35±0.2

Stability of IMM-H007 in Artificial Gastric Juices, Artificial Intestinal Juices, and Tris-HCl Buffer

The stability of IMM-H007, M1, and MP in artificial gastric juices, artificial intestinal juices, and Tris-HCl buffer is shown in Table X and Figure 5. IMM-H007, M1, and MP were stable in artificial gastric juices, artificial intestinal juices, and Tris-HCl buffer. After incubating for 4 h in artificial gastric juices, artificial intestinal juices, and buffer, they remained 118.20%/102.38%/101.60%, 92.67%/94.10%/107.42%, and 89.82%/98.40%/103.64%, respectively. After incubating with the artificial gastric juices, artificial intestinal juices, and buffer, neither the primary drug nor the metabolites were significantly reduced, and no other metabolite was produced, indicating that the prototype drugs and metabolites were less affected by gastrointestinal pH changes.

Discussion

Early pharmacological researches have shown that the M1 and MP metabolites are primarily the *in vivo* active metabolites of the prototype drug IMM-H007. In this study, the rapid, sensitive, and reliable LC-MS/MS method was used to detect the prototype drug IMM-H007 and its metabolites, M1 and MP, at the same time. As the plasma content of MP in mice is low and it primarily exists in whole blood, this experiment utilized whole blood for pharmacokinetic studies. The Guidelines of Non-clinical Pharmacokinetic Study in Chemical Drugs, issued by the State Food and Drug Administration, indicate that

a variety of factors could affect the determination of biological samples due to low concentrations of drugs, endogenous substances (such as inorganic salts, lipids, proteins, metabolites), individual differences, and so on. Therefore, it is necessary to establish a suitable biological sample analysis method and to confirm the method according to the structure, biological properties, and expected concentration range of the compound under examination. Among them, the pretreatment of the sample is a prerequisite for establishing a biological sample analysis method. Commonly used pretreatment methods include protein precipitation, organic solvent, liquid-liquid extraction, and solid-phase extraction. There is a large difference between the chemical properties of IMM-H007 and its metabolites M1 and MP. IMM-H007 and M1 being non-polar (MP having high polarity), the recovery rate of the liquid-liquid extraction method is low. In addition, due to IMM-H007 being unstable at room temperature in whole blood, this step needs to be performed in an ice bath. Therefore, for this experiment, we chose the method of protein precipitation and organic solvent. The experimental results showed that the absolute matrix effect value of IMM-H007, M1, and MP in low, medium, and high concentrations of whole blood samples was 95.49%, 95.49%, and 95.49%, and the test value was basically unchanged, indicating that the LC-MS/MS method established in this research is accurate and reliable for detection of the target compounds and can be applied to the study of pharmacokinetics *in vitro* and *in vivo*. Pharmacokinetics are examined to understand the pharmacological characteristics of a drug and to explain the pharmacological effects of a drug in the body by understanding the absorp-

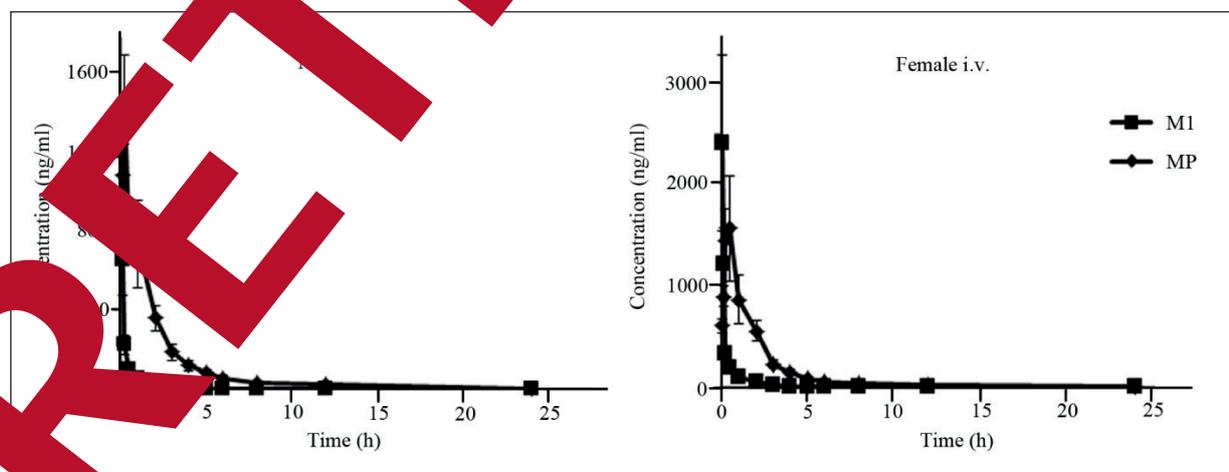


Figure 5. Stability of IMM-H007, M1, and MP in artificial gastric juices, artificial intestinal juices, and Tris-HCl buffer solution.

Table X. Stability of IMM-H007, M1 and MP in artificial gastric juice, artificial intestinal juice and Tris-HCl buffer solution.

Matrix	Time	IMM-H007 (Mean±SD)			M1 (Mean±SD)			MP (Mean±SD)		
		IMM-H007	M1	MP	IMM-H007	M1	MP	IMM-H007	M1	MP
Artificial gastric juice	0 h	4.23±0.02	ND	ND	ND	5.27±0.24	ND	ND	ND	4.91±0.02
	2 h	4.24±0.20	ND	ND	ND	5.17±0.17	ND	ND	ND	4.66±0.16
	4 h	5.00±0.12	ND	ND	ND	5.51±0.11	ND	ND	ND	4.41±0.13
Artificial intestinal juice	0 h	4.42±0.16	ND	ND	ND	5.95±0.08	ND	ND	ND	5.64±0.06
	2 h	4.46±0.16	ND	ND	ND	5.62±0.07	ND	ND	ND	5.83±0.08
	4 h	4.30±0.11	ND	ND	ND	5.67±0.19	ND	ND	ND	5.55±0.26
Tris-HCl buffer solution	0 h	4.37±0.13	ND	ND	ND	5.08±0.05	ND	ND	ND	4.94±0.23
	2 h	4.38±0.08	ND	ND	ND	5.65±0.11	ND	ND	ND	5.15±0.26
	4 h	4.44±0.06	ND	ND	ND	5.81±0.01	ND	ND	ND	5.12±0.26

tion, distribution, metabolism, and excretion of the drug *in vivo*¹⁵. Early drug evaluation, drug design, dosage form modification, optimization of dosing regimen, and guiding clinical rational drug use are of great significance. It is generally believed that the linear kinetic process *in vivo* should have the following characteristics: concentration of drug in the blood decreases exponentially with time; amount of drug transferred in unit time decreases with time; transfer percentage remains unchanged in unit time; half-life is independent of the dose; and $AUC_{(0-t)}$ is proportional to the dose¹⁶. In the efficacy test using golden hamsters, 50 mg/kg were used as an oral dose and administered twice daily. Therefore, this study selected four doses of 50, 150, 450, and 900 mg/kg for pharmacokinetic studies. After IMM-H007 administration in male and female golden hamsters, the $AUC_{(0-t)}$ of the major metabolites MP and M1 in the whole blood increased with increasing dose with good correlation, indicating that the metabolism of M1 and MP is essentially a linear kinetic process *in vivo* in the selected dose range. Since the blood prototype drug concentration was low, close to, or below, the detection limit after oral or intravenous IMM-H007 administration, we used M1 and MP to calculate the bioavailability, which was 6.97% and 4.78%, respectively, in the whole blood of male golden hamsters and 2.20% and 6.75%, respectively, in female golden hamsters. The bioavailability of male and female golden hamsters was 6.97% and 8.95% using the sum of M1 and MP, which was relatively low. There are three main possible factors that dictate bioavailability: (1) physico-chemical properties of the compound, such as stereo-chemical structure, solubility, permeability, etc. (2) gastrointestinal environment. There are many transporters on small intestinal epithelial cells, such as P-gp, which affect the transport of drugs in the gastrointestinal tract. In addition, abundant intestinal bacteria are also involved in the metabolism of drugs *in vivo*. Other factors may also include pathological conditions, drug interaction, and individual differences. The specific reasons for the low bioavailability of IMM-H007 need to be investigated for the study of bile excretion and intestinal metabolism. The drug-time curves of the primary metabolites in whole blood after oral administration and sublingual venous injection of IMM-H007 in golden hamsters was irregular, likely due to the heterogeneity of gastrointestinal absorption and reabsorption of the drug in tissue. After drugs are discharged from the gastric juices into the intestine, a large number

of drugs are re-transformed into non-ionic forms due to the neutral pH of the intestinal fluid. At this point, transmembrane transport and absorption are facilitated, so that the plasma concentration has multiple peaks. In addition, different parts of the gastrointestinal tract, the organelles exhibit differential permeability for certain drugs that can also cause irregular curves because high concentrations of drugs in the tissue may be re-released into the blood and then redistributed throughout the body. This phenomenon also causes irregular changes for the drug-time curves. The ribbing of IMM-H007 contains H acceptor groups, which, like other ester structures, are easily hydrolyzed in animal plasma both *in vitro* and *in vivo*. Therefore, the study of IMM-H007 biological transformation of *in vivo* biological samples helps to understand its metabolic pathway *in vivo*. In this work, the relationship between stability and actual transformation of IMM-H007 and its major metabolites M1 and MP was studied in *in vitro* biological samples using *in vitro* thermolysis. The results showed that IMM-H007, M1, and MP were stable in artificial gastric juices, artificial intestinal juices, and Tris-HCl buffer, suggesting that the prototype drug and metabolites were less affected by the acidic environment in gastrointestinal pH and digestive enzymes.

Conclusions

In this research, an efficient, sensitive, and reliable LC-MS/MS method was established and applied to pharmacokinetics and related studies after oral or sublingual intravenous IMM-H007 administration in hamsters. The experimental results demonstrated that IMM-H007 is rapidly absorbed through the oral route in hamsters. The C_{max} and $AUC_{(0-t)}$ of the metabolites M1 and MP in male and female hamsters were increased with increasing dose and were proportional to the dose, and $T_{1/2}$ and $MRT_{(0-t)}$ were significantly prolonged with increasing dose, which basically show linear dynamic characteristics and no significant gender differences. The bioavailability in male and female golden hamsters after oral IMM-H007 administration was calculated using the sum of M1 and MP, resulting in 6.97% and 8.95%, respectively. The IMM-H007 and its metabolites were stable in Tris-HCl buffer, artificial gastric juices, and artificial intestinal juices. These findings provide reference for clinical application research.

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Conflict of Interests

The authors declare that they have no conflicts of interest.

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