

Preclinical study of vitamin D deficiency in the pathogenesis of metabolic syndrome in rats

S.K. MAHJOUB¹, M.A.A. SATTAR AHMAD², F.O. KAMEL², M. ALSEINI², L.M. KHAN²

¹King Fahad General Hospital, Ministry of Health, Jeddah, Saudi Arabia

²Department of Pharmacology, Faculty of Medicine, King Abdulaziz University, Jeddah, Saudi Arabia

Abstract. - OBJECTIVE: To explore the impact of vitamin D deficiency (VD-) in the pathogenesis of metabolic syndrome (MetS).

MATERIALS AND METHODS: Models of (VD-) and (MetS) were induced in male Wister rats by dividing into four groups, group-I for the development of (VD-) by intraperitoneal injection of paricalcitol for 3 weeks, group II for (MetS) model by adding 10% fructose to their drinking water for 8 weeks, the group III for induction of combined (VD- + MetS) and group-IV as a control. Ultimately, the parameters of (VD-) and (MetS) were assessed at zero time and after 8 weeks.

RESULTS: Both (VD-) and (MetS) groups alone displayed a remarkable enhancement of blood pressure, glucose and insulin levels, glycerides, cholesterol, and low-density lipoproteins with a reduction of high-density lipoproteins. Additionally, all distinguishing features of obesity were substantially increased. Nevertheless, the combined group (VD-+MetS) demonstrated an expeditious and established restoration in all the aforesaid parameters compared to the (VD-) and (MetS) groups alone.

CONCLUSION: The hallmark of this study, reinforces a new frontier of awareness of the deleterious effect of (VD-) on each component of (MetS). Eventually, the implementation of vitamin D may circumvent the elements of (MetS) and merits further validation. The determination of (VD-) molecular pathway on the parameters of (MetS) is under investigation.

Key Words: Vitamin D deficiency, Metabolic syndrome, Interleukin-6, Homeostatic model assessment for insulin resistance, Nuclear factor kappa B.

Introduction

An exclusive characteristic of the hormonal active configuration of vitamin D 1,25-dihydroxycholecalciferol (calcitriol) is the coordina-

tion complex or a ligand for vitamin D receptor (VDR) which causes conformational changes that culminate in transcription factor (RXR) heterodimerization. Moreover, this heterodimer subsequently acts as a transcription factor, that activates target gene expression at the transcriptional cell level and thus, reduces protein synthesis¹. Remarkably, over two hundred genes are regulated by calcitriol directly or indirectly, contributing to a wide variety of physiological functions. Moreover, vitamin D is currently of great community health interest because vitamin D deficiency (VD-) is a common worldwide deficiency with high prevalence². Although vitamin D was the main function on bone and skeletal health, it also has many important functions including modulation of the body's immune system, anti-inflammatory, anti-hypertensive activity, and antiproliferative effect in myocardial hypertrophy²⁻⁴. Besides, vitamin D has a role in glycemic control, insulin release, and lipid metabolism. So, all of these elucidate that vitamin D significantly impacts the body's metabolic functions⁵. Remarkably, several studies demonstrated an inadvertent and robust correlation between vitamin D deficiency and cardiometabolic risk, which invariably develops in the presence of other chronic disorders like obesity, diabetes mellitus, atherosclerosis, and hypertension²⁻⁷. Seemingly, vitamin D deficiency is a major contributory factor to the highly prevalent metabolic syndrome⁸. Thus, the basic essence directed towards our objective to conduct this research is to establish a concrete relationship between vitamin D deficiency and etiopathogenesis of cardiovascular metabolic syndrome and further to precisely clarify the underlying mechanisms and cardiovascular complications in rats. Ultimately, the outcome of combined vitamin D deficiency and metabolic syndrome were likewise investigated.

Materials and Methods

This experimental work was done at King Fahd Center for Medical Research (KFCMR), King Abdulaziz University (KAU), Jeddah, Saudi Arabia, the KAU Research Ethical Committee granted the protocol of this animal study.

Chemical and Reagents

Paricalcitol (19-nor-1,25-dihydroxyvitamin D₂) purchased from AbbVie company, Diethyl ether, Krebs-Henseleit buffer solution, Buffered formalin, from Sigma-Aldrich® Co. (Bayouni Trading Co. Ltd., Al-Khobar, KSA).

Formulations

The Krebs-Henseleit buffer solution was used for maintaining the physiological media for the aorta and Fructose solution for induction of metabolic syndrome⁹.

Animals

We obtained 48 adult male Wister rats approximately weighing 140-220 g from King Fahad Medical Research Center, King Abdulaziz University, Jeddah, Saudi Arabia. Furthermore, the rats were kept singly in cages with all possible care, to keep the environmental conditions constant all the time. They were kept in animal care for one week before the commencement of the experimental study with due care to provide free access to food and water at all time.

Diet of the Animals

The Harlan Teklad custom diet [TD87095] (vitamin D deficient) procured from Envigo® Company, Indianapolis, USA. This is brown chow pellets deficient in vitamin D and comprises of 20% fructose, 15% corn starch, 17.6% protein, 5% fat, 2% calcium (Ca²⁺), and 1.25% phosphate (P²⁺).

In addition, a regular rat diet was obtained from the Granulos & Sons mills organization, Jeddah, KSA, and comprises: 20% protein, 4% fat, 1% calcium, 0.6% phosphate, and vitamin D 2.20 IU/g.

Experimental Design

The experimental animals were randomly categorized into four groups, each group comprising 12 rats (n= 12).

1. The control group (C) – recipient of regular diet and water *ad libitum*.
2. The vitamin D deficient group (VD) – from the first day this group of rats received the

Harlan Teklad custom diet, deficient in vitamin D, intended to develop vitamin D deficiency. These rats were injected intraperitoneally with paricalcitol [19-nor-1,25-dihydroxyvitamin D₂, an active form of vitamin D₂ (VDR agonist, Zemplar®) in a dose of 32 ng on days 1, 3, 5, 8, 10, and 12 to enhance the metabolism of endogenous vitamin D metabolites by cytochrome enzyme induction for 3 weeks¹⁰. Subsequently, the serum levels of vitamin D were measured in the third week to confirm the induction of vitamin D deficiency. The rats then remained on a vitamin D deficient diet and regular water *ad libitum* for 8 more weeks.

3. Metabolic syndrome group (MetS): Animals were fed regular diet and regular water was added to drinking water (equivalent to the diet containing 48-57% fructose)^{11,12}. A regular diet with 10% fructose water were supplemented *ad libitum* for 8 weeks to induce metabolic syndrome⁹.

Combined group (vitamin D deficiency and metabolic syndrome) (VD + MetS): after rendering the rats vitamin D deficient, the animals were fed 10% fructose added to their drinking water and on a vitamin D deficient diet for 8 weeks to induce metabolic syndrome.

Body Weight Measurements

The body weights for rats were measured at the baseline and on the 8th week by the end of the experiment using a calibrated, sensitive weighing instrument (acquired from Al-Falak Electronic Industries Company, Jeddah, KSA) and calculated in gram (g).

Blood Samples Collection

The rats were anesthetized by utilization of diluted ether and then blood samples were aspirated from the choroid plexus by using capillary tubes and collected in a 5 ml blood tube then centrifuged at 4,500 rpm for 15 min. Blood samples were taken at Baseline, on the 3rd week to confirm vitamin D deficiency and on the 8th week of the experiment for the combined vitamin D deficient and metabolic syndrome groups.

The underlying biochemical evaluation was executed to study the effect of vitamin D deficiency in the pathogenesis of cardiovascular metabolic syndrome and the possible underlying mechanism:

1. Serum vitamin D level (assessment of vitamin D deficiency).

2. Serum calcium, phosphate & magnesium levels (assessment of mineral homeostasis).

3. Assessment of the parameters of metabolic syndrome (viz: diabetes mellitus, obesity, hyperlipidemia, and hypertension). The development of metabolic syndrome was evaluated according to the guidelines of NCEP ATP III (2001)¹³, which includes insulin resistance (increased insulin values, impairment of fasting blood glucose level (>5.5 mmol/L), hypertension (>130/85), glucose tolerance impairment and type II diabetes mellitus, enhanced abdominal obesity (increased waist circumference and BMI), hyperlipidemia (increased TG levels >1.995 mmol/L or decreased levels of HDL). Notably, the presence of a minimum of three of these parameters is labeled as having metabolic syndrome¹³.

Assessment of Vitamin D Level

Serum vitamin D level was measured at the baseline for all tested groups and on the 3rd week for the VD group and VD + MetS group. The determination of vitamin D level was done by Cobas[®] automated analyzer vitamin D assay (F. Hoffmann-La Roche Ltd., Basel, Switzerland). The vitamin D level was calculated and expressed in terms of nmol/L¹⁰.

Assessment of Calcium (Ca^{+2}), Phosphate (PO_4^{-3}) and Magnesium (Mg^{+2}) Levels

Serum Ca^{+2} , PO_4^{-3} and Mg^{+2} were measured for all test groups, at the baseline and the 8th week by the end of the experiment by using Flex[®] reagent cartridge on the Dimension Vista[®] system, (obtained from Siemens Healthineers, Riyadh, KSA). Subsequently, their concentrations were measured using bichromatic endpoint technique. Ca^{+2} at absorbance (577-450 nm), PO_4^{-3} absorbance (340/700 nm) & Mg^{+2} at absorbance (600 and 510 nm). Their levels were calculated as mmol/L.

Hypertension

Measurement of blood pressure

Measurement of systolic and diastolic blood pressure was done using a Tail Cuff reader (non-invasive technique) by using UGO BASILE[®] 58500 blood pressure recorder (obtained from Gemonio, Varese, Italy), at the baseline, as well as after 8 weeks for all tested groups by the technique of Widdop and Li, 1997^{14,15}.

Diabetes Mellitus

Measurement of glucose level

The serum glucose level was measured at the beginning of the experiment and the 8th week for all experimental groups with the use of Flex[®] reagent cartridge over the Dimension Vista[®] Siemens system (acquired from Healthineers, Riyadh, KSA). However, the level of blood glucose was measured by utilization of a bichromatic (at absorbance 340 and 583 nm) endpoint technique, finally computed as mmol/L.

Measurement of insulin concentration

The serum insulin level was measured at the baseline and the 8th week for all tested groups. This was accomplished by using RayBio[®] rat insulin ELISA [Acquired From-(CGenomix) Amman, Jordan]. Insulin level was calculated in μ IU/L.

Insulin resistance index

The insulin resistance index was measured for all tested groups by the utilization of the standard method of Matthews et al¹⁶, with the mathematical equation:

$$\text{IR} = \frac{\text{FPGMA} - \text{IR} = \text{glucose concentration (mmol/L)}}{\text{fasting insulin } (\mu\text{IU/L})/22.5}$$

Dyslipidemia

Measurement of a lipid profile

The serum concentration of cholesterol, TG, HDL, and LDL was determined at the baseline and the 8th week for all tested groups. The lipid profile was determined using Flex[®] reagent cartridge on the Dimension Vista[®] SIEMENS (acquired from Healthineers, Riyadh, KSA). Serum cholesterol by polychromatic (540, 452 & 700 nm) endpoint technique, TG by bichromatic (510,700 nm) endpoint technique, HDL was measured using bichromatic (600, 700 nm) technique, and LDL concentration was measured using a bichromatic (540, 700 nm) endpoint technique. These parameters were calculated in mmol/L.

Obesity

The percentage of body weight gain, BMI, abdominal fat weight, adiposity index, and AC of the rats were measured as indicators of obesity^{17,18}, essentially, this was done at the start of the experiment and then at the 8th week for all tested groups. This was done using an elec-

Table I. Serum vitamin D level (nmol/l) at the baseline and on the 3rd week after induction of vitamin D deficiency.

Animal Groups Duration	Control (n = 12)	VD- (n = 12)	MetS (n = 12)	(VD- +MetS) (n = 12)
Baseline values	84.3 ± 4.19	88 ± 0.78	88.24 ± 3.11	88.15 ± 3.87
3 rd week of vitamin D deficiency induction (< 12.5 nmol/l)	84.3 ± 4.19	7.33 ± 0.09***	88.24 ± 3.11	7.96 ± 0.2

*** $p < 0.0001$ compared to the respective baseline values.

tronic, calibrated weighing scale (acquired from Al-Falak Electronic Industries Company, Jeddah, KSA). Adiposity index was measured according to the formula:

$$\text{Adiposity index} = \frac{\text{Perennial WAT} + \text{Retroperitoneal WAT} + \text{Epididymal WAT} \times 100}{\text{Bodyweight}}$$

Statistical Analysis

All biochemical outcomes were expressed as mean ± standard error. A paired *t*-test was used to evaluate the mean change in biochemical measurements within the experimental group from time zero to 3 and 8 weeks. One-way analysis of variance (ANOVA) followed by Tukey's HSD test was performed and statistical comparisons among the various groups were conducted using a Graph Pad prism program and excel for data analysis (La Jolla, CA, USA). Statistical significance is set at $p < 0.05$.

Results

Serum Vitamin D Level

By the 3rd week, the vitamin D deficient groups [VD- & VD- + MetS] showed a significant decrease in vitamin D level (< 12.5 nmol/l) with a p -value < 0.0001 compared to the baseline value (Table I).

Table II. Serum calcium, phosphate and magnesium levels (mmol/l) at the baseline and after 8 weeks.

Animal groups	Duration	Calcium (mmol/l)		Phosphate (mmol/l)		Magnesium (mmol/l)	
		Baseline	After 8 weeks	Baseline	After 8 weeks	Baseline	After 8 weeks
	Control n = 12	2.63±0.014	2.63±0.014	2.42±0.13	2.42±0.13	0.9±0.03	0.9±0.03
	VD- n = 12	2.36±0.02	2.34±0.02	2.03±0.07	2.01±0.046	0.87±0.01	0.87±0.02
	MetS n = 12	2.74±0.03	2.49±0.02**	2.7±0.06	2.1±0.09**	0.89±0.018	0.87±0.026
	(VD- + MetS) n = 12	2.60±0.03	2.66±0.03i	2.63±0.07	2±0.07**	0.89±0.018	0.852±0.02

**Significant ($p < 0.001$) compared to baseline of each group.

Serum Level of Calcium (Ca^{+2}), Phosphate (PO_4^{-3}), and Magnesium (Mg^{+2})

The metabolic syndrome (MetS) alone exhibited a significant decrease ($p < 0.0001$) at the end of 8 weeks (Table II) for phosphate levels, only the vitamin D deficiency group (VD) showed a non-significant decrease (p -value > 0.05) vs. each baseline. Moreover, the magnesium levels for all test groups showed a non-significant decrease (p -value > 0.05) vs. each baseline.

Hypertension

Measurement of blood pressure

A significant increase in both systolic & diastolic blood pressure ($p < 0.0001$). VD-: vitamin D deficiency group, MetS: a group of metabolic syndromes (VD- + MetS): collective group (both vitamin D deficiency and metabolic syndrome), in comparison to the control group (Table III).

Diabetes Mellitus

Glucose level measurement

The level of glucose was found to be significantly augmented ($p < 0.05$) in all the groups compared to the control group (Table IV). Moreover, the combined group (VD- + MetS) demonstrated a significant increase in glucose level ($p < 0.05$) compared to the vitamin D deficient (VD-) and metabolic syndrome (MetS) groups alone.

Table III. Systolic and diastolic blood pressure (mmHg) measurements at baseline and after 8 weeks.

Parameter measured	Groups Duration	Control (C) n = 12	Vitamin D deficient (VD-) n = 12	Metabolic syndrome (MetS) n = 12	Combined (VD- + MetS) n = 12
Systolic blood pressure (mmHg)	Baseline	120±0.57	121.5±0.51	122±0.41	122±0.53
	After 8 weeks	120±0.57	135±0.7 ***	150.5±1.4 ***	166.33±3.8 ***
Diastolic blood pressure (mmHg)	Baseline	81±1.3	81±1.3	81±1.45	81±1.45
	After 8 weeks	81±1.3	94.17±1.7***	98.5±1.22***	111.5±2.1***

***significant ($p < 0.0001$) vs. the control group.

Insulin level measurement

Table V shows a significant increase in serum insulin level ($p < 0.0001$) for the vitamin D deficient (VD-), metabolic syndrome (MetS), and combined (VD- + MetS) groups produced compared to control groups. Interestingly, the combined group exhibited a significant increase in insulin level ($p < 0.0001$) compared to other groups.

Insulin resistance index

As illustrated in Table VI the equilibrium evaluation model for insulin resistance index (HOMA-IR) was found to be significantly augmented in all test groups ($p < 0.0001$) compared to the control group. Moreover, the vitamin D deficient (VD-) group and the combined (VD- + MetS) group had significant increase in insulin resistance index ($p < 0.001$) for (VD-) and ($p < 0.0001$) for (VD- + MetS) compared to the metabolic syndrome (MetS) group alone.

Dyslipidemia

Triglycerides (TG) level

Table VII and Figure 1 showed that the (VD-) group produced a significant increase in TG serum level ($p < 0.05$) compared to the control group whereas both (MetS) and (VD- + MetS) groups produced a significant increase in TG serum level ($p < 0.001$) compared to the control group.

Total cholesterol level

Table VIII showed that the total cholesterol was significantly increased ($p < 0.05$) for the vitamin D

deficient (VD-) group and the combined group (VD- + MetS) in comparison to the control group. In contrast, the metabolic syndrome (MetS) group showed a non-significant ($p > 0.05$) difference compared to the control group.

HDL level

Conventionally, the level of HDL was demonstrated to be quite significantly decreased in the metabolic syndrome (MetS) group ($p < 0.05$) when compared with the control group (Table IX). However, Figure 1 showed that HDL level was increased from baseline to the 3rd week of the experiment followed by a significant decrease in HDL level after 8 weeks for the vitamin D deficient (VD-) group ($p < 0.05$) and the combined (VD- + MetS) group ($p < 0.0001$).

LDL level

Table VII showed that LDL level was significantly increased in vitamin D deficient (VD-) group ($p < 0.001$) and the combined (VD- + MetS) group ($p < 0.0001$) while the metabolic syndrome (MetS) group showed a non-significant ($p > 0.05$) difference vs. control group.

Obesity

Percentage of weight gain

Table VIII and Figure 2A showed a significant increase in the percentage of weight gain of the vitamin D deficient (VD-) group ($p < 0.001$), metabolic syndrome (MetS) group ($p < 0.0001$), and the combined (VD- + MetS) group ($p < 0.0001$) compared to the control group.

Table IV. Serum glucose level (mmol/L) at baseline and after 8 weeks.

Parameter measured	Groups Duration	Control n = 12	Vitamin D deficient (VD-) n = 12	Metabolic syndrome (MetS) n = 12	Combined (VD- + MetS) n = 12
Serum Glucose level (mmol/L)	baseline	7.2±0.02	7.23±0.01	7.47±0.02	7.24±0.02
	After 8 weeks	7.2±0.02	8.3±0.5*	8.2±0.22*	10.2±0.8*

*Significant ($p < 0.05$) vs. control group.

Table V. Serum insulin level (Pmol/L) after 8 weeks.

Groups parameter measured	Control n = 12	Vitamin D deficient (VD-) n = 12	Metabolic syndrome (MetS) n = 12	Combined (VD- + MetS) n = 12
Insulin Level (Pmol/L)	99±6.05	379±15***	272±19***	475.5±5.9***

***Significant ($p < 0.0001$) vs. control.

Body mass index (BMI)

As shown in table VIII and Figure 2B, a significant increase in body mass index ($p < 0.001$) of all tested groups [VD-, MetS & (VD- + MetS)] compared to the control. Moreover, the combined (VD- + MetS) group showed a significant ($p < 0.001$) increase in BMI compared to each metabolic syndrome (MetS) group and vitamin D deficient (VD-) group.

Abdominal circumference (AC)

Table VIII and Figure 3 showed that the abdominal circumference was significantly ($p < 0.0001$) increased in all tested groups [(VD-, MetS & (VD- + MetS)] when compared with the control group.

Total abdominal fat

Table VIII and Figure 4 illustrated that the total abdominal fat was significantly increased for the vitamin D deficient (VD-) group ($p < 0.05$), metabolic syndrome (MetS) group ($p < 0.001$) and vitamin D deficiency with metabolic syndrome (VD- + MetS) group ($p < 0.001$) compared to the control group. Furthermore, both the metabolic syndrome (MetS) group and the combined (VD- + MetS) group produced a significant increase in total abdominal fat ($p < 0.01$) vs. the vitamin D deficient (VD-) group.

Adiposity index: Table VI and Figure 5 showed that the adiposity index was significantly increased for the vitamin D deficient (VD-) group ($p < 0.05$), metabolic syndrome (MetS) group ($p < 0.001$) and vitamin D deficiency with metabolic syndrome (VD- + MetS) group ($p < 0.001$) compared to the control group.

Table VI. Mean serum insulin and glucose levels and insulin resistance index after 8 weeks.

Groups Parameter measured	Control n = 12	VD- n = 12	MetS n = 12	(VD- + MetS) n = 12
Insulin (μ IU/L)	14.256±0.83	54.576±2.1	39.168±1.4	68.472±0.83
Glucose (mmol/L)	7.38±0.02	8.3±0.5	8.2±0.22	10.2±0.8
HOMA-IR	4.678±0.21	20.173±2.4***	14.434±0.7***	31.041±1.43***

***Significant ($p < 0.0001$) vs. control.

Evaluation of the emerging metabolic syndrome by NCEP ATP III (Table VII)

Human metabolic syndrome criteria could be extrapolated to animal models. Except for the central obesity measure (Table VII). All test groups (VD-, MetS) and (VD- + MetS) resulted in a significant increase in insulin resistance, glucose, blood pressure, obesity parameters, and TG level compared to the control group as mentioned above in detail. Table VIII shows that the metabolic syndrome (MetS) and the combined (VD- + MetS) groups accomplished the criteria of metabolic syndrome (marked (✓) in table) according to NCEP-ATP III.

Discussion

Is it Worthwhile Skepticism Is that Vitamin D Deficiency Is Related to Metabolic Syndrome

Essentially, vitamin D is a prime nutrient prerequisite to maintain the standard health and it is an active component of the steroid nuclear hormone family^{1,5}. Low vitamin D level is linked with several diseases like osteoporosis, rickets, cancer, and cardio-metabolic complications^{2-4,19,20}. In addition, there is a great suspicion that vitamin D deficiency is related to metabolic syndrome (MetS), a complex clinical condition that comprises abdominal obesity, disturbed glucose metabolism, atherogenic dyslipidemia, and essential hypertension^{6-8,21}.

The Implication of Vitamin D in the Release and Action of Insulin

In this study, vitamin D level was significantly reduced in all three groups (VD-, MetS alone and

Table VII. Serum cholesterol, triglycerides, HDL, and LDL levels (mmol/L) after 8 weeks.

Groups Parameter measured	Control (C) n = 12	Vitamin D deficient (VD-) n = 12	Metabolic syndrome (MetS) n = 12	Combined (VD- + MetS) n = 12
Triglyceride (mmol/L)	0.83±0.03	1.143±0.03*	1.74±0.09**	2.3±0.18**
Total cholesterol (mmol/L)	1.35±0.04	1.65±0.08*	1.4±0.04	1.76±0.07**
HDL level (mmol/L)	1.26±0.03	1.5±0.04	1.01±0.06*	1.33±0.04
LDL level (mmol/L)	0.14±0.003	0.22±0.02**	0.153±0.002	0.2±0.009

*Significant ($p < 0.05$) vs. control group. **significant ($p < 0.001$) vs. control group. ***significant ($p < 0.0001$) vs. control group.

combined (VD- + MetS) at the end of 3 weeks (Table I). This was well illustrated by the significant increase in insulin resistance, blood glucose level, blood pressure, serum lipids, and obesity (Table III-VIII). The present results were explained based on many different mechanisms like facilitation of the synthesis and secretion of insulin directly by the metabolites of vitamin D^{22,23}. Further substantiation of the role of vitamin D is explained, based on its control over cytosolic calcium-binding protein found in the β -cells. Seemingly, vitamin D performs as a regulator of depolarization-induced insulin release by eventual calcium regulation remarkably authenticated²⁴⁻²⁸.

Accumulating evidence implicates that vitamin D preserves the release of insulin and facilitates insulin actions in the tissues. It is responsive to the insulin, by controlling the intracellular and extracellular calcium levels²⁶. Nevertheless, numerous studies revealed a strong correlation between hypovitaminosis D and metabolic syndrome^{29,30}. Likewise, many epidemiological

studies have shown that there is an inverse correlation between the level of vitamin D, blood glucose, insulin resistance, and the prevalence of type 2 diabetes³¹. Thus, we would lay the emphasis, that our present study has unequivocally demonstrated a significant increase in insulin level and insulin resistance index (HOMA-IR) in all test groups compared to the control group. The insulin level and the insulin resistance index (HOMA-IR) were significantly higher in the combined (VD- + MetS) group (Table VI and VIII). This noteworthy finding is very well corroborated by the fact that TNF and IL-6 are immune mediators which interfere with insulin signaling that leads to insulin resistance^{24,33}.

The Contemplation of Electrolyte Imbalance in Diabetes and Metabolic Syndrome due to an Imbalance of Vitamin D

The reflection of underlying details of serum levels of Calcium (Ca^{+2}), Phosphate (PO_4^{-}

Table VIII. Metabolic syndrome parameter according to NCEP-ATP III.

NCEP-ATP III	Insulin resistance	Blood glucose level	Dyslipidemia	Blood pressure	Central obesity	
	HOMA-IR	> 5.5 mmol/L	TG > 1.695 mmol/L	> 130/85 mmHg	BMI	AC
Control	4.256±0.8	7.2±0.02	0.83±0.03	120±0.57	0.45±0.01	15.3±0.26
VD-	20.17±0.4	8.3±0.5 √	1.143±0.03	135±0.7 /	0.52±0.01	17.33±0.30
MetS	14.434±0.7	8.2±0.22 √	1.74±0.09 √	150.5±1.4 /	0.55±0.008	17±0.23
VD- + MetS	31.041±1.43	10.2±0.8 √	2.3±0.18 √	166.33±3.8 /	0.61±0.01	19.34±0.14
				111.5±2.21 √		

VD-: Vitamin D deficiency group, MetS: a group of metabolic syndrome, (VD- + MetS): collective group (both vitamin D deficiency and metabolic syndrome).

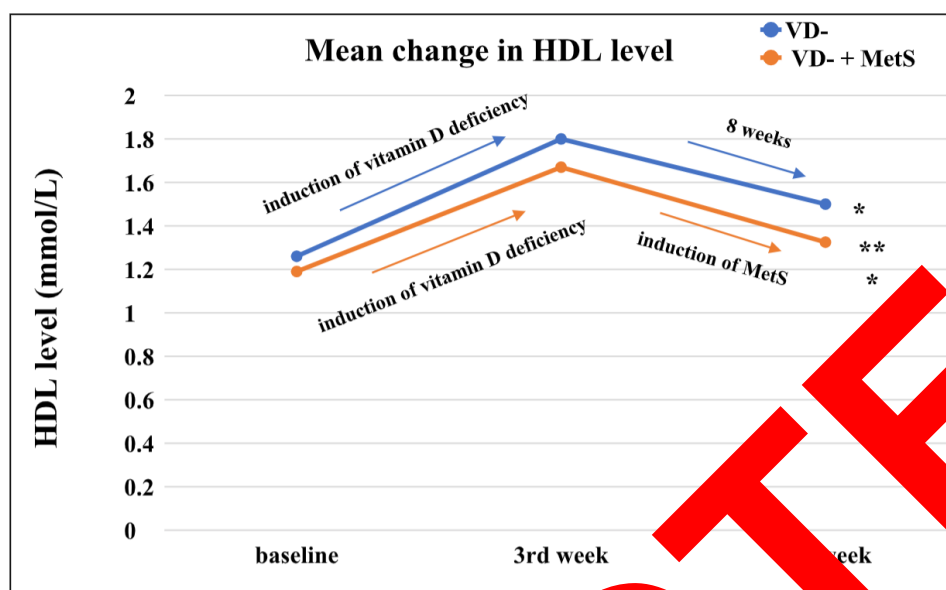


Figure 1. Mean change OF HDL level at baseline, at the 3rd week after induction of vitamin D deficiency, and after 8 weeks (n=12 rat/group). ***significant ($p < 0.0001$) vs. the level on the 3rd week, *significant ($p < 0.05$) vs. the level on the 3rd week.

³), and Magnesium (Mg²⁺) in our results (Table II), could be emphatically attributed to electrolyte imbalance as a consequence of diabetes and metabolic syndrome due to derangement of PTH-vitamin D endocrine system^{34,35}. Interestingly, the enriched phosphate and magnesium diet given to the rats in this study managed to restore the phosphate level in the combined (VD- + MetS) group due to the correction of metabolic syndrome. This observation is significantly endorsed by a streptozotocin-induced diabetes study in which rats developed hypocalcemia³⁶. In addition, a recent study has revealed changes in the magnesium level in all the animal groups, nevertheless several studies demonstrate a correlation between hypomagnesemia with hyperglycemia and diabetes^{37,38}. However, magnesium deficiency leads to hypocalcemia mostly due to impaired PTH release or renal tubule resistance to the action of PTH³⁹.

Cardiovascular Effects of Vitamin D Deficiency as Risk Factors for the Development of the Metabolic Syndrome

Induction of vitamin D deficiency resulted in noteworthy enhancement of both systolic and diastolic blood pressure in all test groups with the greatest increase in the combined group (Table III and VIII), which is a remarkable observation emerged from this study that needs to be

highlighted. This evidence is splendidly sustained by different studies indicating the expression of genetic factors complicating the myocardial and metabolic functions^{40,41}. Furthermore, recent past studies reported that vitamin D-depleted male rats, developed a significant increase in blood pressure by a mechanism involving renin, and this is independent of Ca²⁺ or PTH^{29,42,43}. The induction of hypertension in the VD- group, MetS, and combined group (Table III) is further reinforced and validated indeed by the finding that vitamin D and its related metabolites can reduce contraction of the vascular tone in rats with elevated blood pressure. This is accomplished, by affecting the calcium influx and reducing the production of endothelium-derived contracting factors (EDCF)^{44,45}. Furthermore, VDR stimulation induces the production of nitric oxide (NO) in the endothelium and improves the angiogenic properties of the endothelium progenitor cells (EPC)^{43,46}. These studies strongly support that vitamin D has a key regulatory part and represents a new paradigm in blood pressure homeostasis.

Impact of Vitamin D Deficiency on Lipids and Triglycerides

A momentous highlight of our present work has revealed that vitamin D deficient group (VD-) and (VD- + MetS) induced a significant increase in TG, LDL and cholesterol levels compared to the control group (Table

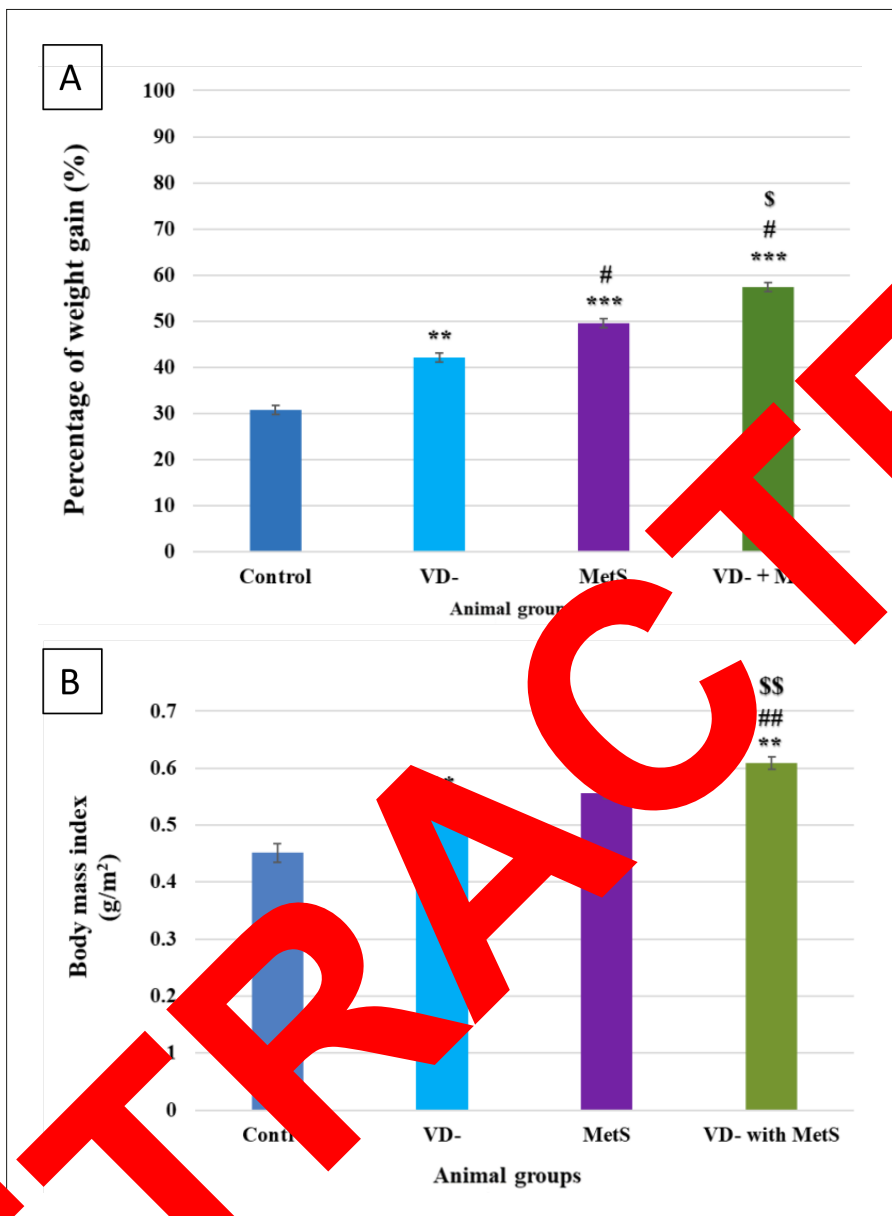


Figure 1. Mean \pm S.E of: (A) Percentage of weight gain, (B) body mass index of control VD-, MetS and (VD- + MetS) groups after 12 weeks (n=10 rat/group). ***Significant ($p < 0.0001$) vs. control group, **significant ($p < 0.001$) vs. control group, #significant ($p < 0.05$) vs. (VD-) group, \$ significant ($p < 0.01$) vs. (MetS) group.

&VIII). The present observation is concordant with a study involving vitamin D deficient mice which reported elevated TG and total cholesterol levels. In addition, vitamin D has an impact on insulin secretion and sensitivity which, indirectly stimulates the metabolism of lipid⁴⁷. Moreover, there is strong evidence of an association between the state of vitamin D deficiency and insulin resistance⁴⁸. Conversely, in the event of insulin resistance and conse-

quent hyperglycemia, the intracellular reactive oxygen species (ROS) has been observed to be substantially elevated and this accumulation of ROS, in turn activates nuclear factor-kappa B (NF- κ B) leading to liver inflammation⁴⁹. Several recent studies emphatically exhibited that vitamin D diminishes the expression of NF- κ B and conversely augments the expression of peroxisome proliferator-activated receptor alfa (PPAR- α)^{50,51}.

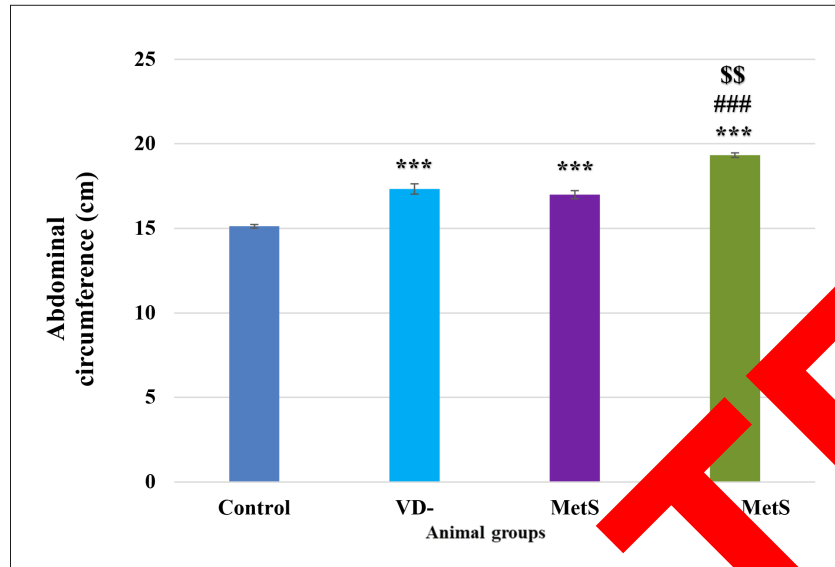


Figure 3. Mean± S.E of abdominal circumference for control, VD-, MetS and (VD- + MetS) groups after 8 weeks (n=12 rat/group). ***significant ($p < 0.0001$) vs. control group, \$\$\$ significant ($p < 0.0001$) vs. MetS group, \$\$ significant ($p < 0.001$) vs. VD- group.

Attribution of Vitamin D Deficiency to the Distinctive Characteristic of Metabolic Syndrome

A distinct, noteworthy characteristic of metabolic syndrome is the preponderance of abdominal obesity. In the present study, an unequivocal quality not usually correlated

with Vitamin D deficiency⁵². This association has been empirically recognized in our current study (Figure 2-5 and Table VIII) and has generated a significant increase in AC, BMI, percentage of weight gain, adiposity index, and total abdominal fat weight in all the experimental groups. A finding of particular interest is re-

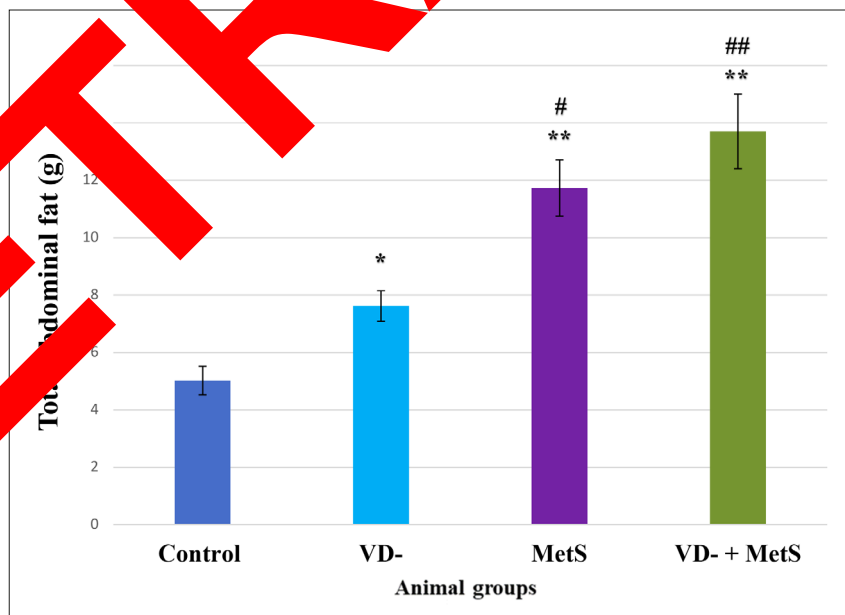


Figure 4. Mean ±S. E of total abdominal fat for control, VD-, MetS and (VD- + MetS) groups after 8 weeks (n=12 rat/group). **significant ($p < 0.001$) vs. control group, *significant ($p < 0.05$) vs. control group, #significant ($p < 0.05$) vs. VD- group, ##significant ($p < 0.001$) vs. VD- group.

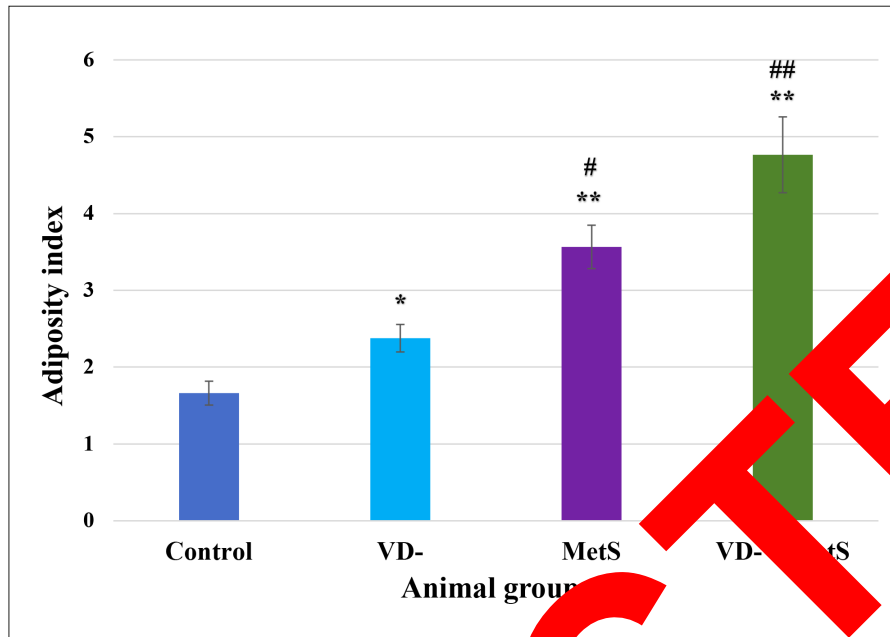


Figure 5. Mean \pm S.E. of adiposity index for control, VD-, MetS (Metabolic Syndrome) (VD- + MetS) groups after 8 weeks (n=12 rat/group). **significant ($p < 0.001$) vs. control group, *significant ($p < 0.05$) vs. control group, #significant ($p < 0.01$) vs. VD- group, ##significant ($p < 0.001$) vs. VD- group.

lated to the mean increase in BMI and AC for the VD- group and (MetS) group which were similar, while a stronger effect was indicated in the combined group than in either group alone (Figure 2-4 and Table VIII). These findings undoubtedly reinforces the deleterious effect of vitamin D deficiency on metabolic syndrome. Interestingly, a reassuring cross-sectional study acquired from the insulin resistance atherosclerotic study of Hispanic adults exhibited a higher level of calcidiol and, in turn, is inversely related to the body mass index (BMI) and parameters of visceral and subcutaneous adiposity^{53,54}. Uncontroversially, higher risk of metabolic syndrome, obesity, increased waist circumference, and metabolic dyslipidemia were found to be strongly associated with low calcidiol levels and high PTH concentrations^{52,55}. The agglomeration of abdominal adipose tissue leads to the progression of dyslipidemia, hyperglycemia, and hyper-

Correlation of Hypovitaminosis D with Inflammatory Markers and Obesity

Another focus of interest of this study is that hypovitaminosis D is related to increased inflammatory markers such as: TNF and C-reactive protein in obese subjects⁵⁷. The basic essence of

mechanisms recommended to explain the correlation between vitamin D deficiency and obesity incorporates the requisitioning of vitamin D in adipose tissue, which is attributable to its inherent lipophilic property and diminished synthesis of calcidiol in the liver as a result of hepatic steatosis or to the proinflammatory cytokines inhibitory actions⁵⁸.

Thus, the present study has illustrated a new frontier of awareness that vitamin D deficiency can induce different components of metabolic syndrome; hypertension, diabetes mellitus, insulin resistance, dyslipidemia, and obesity.

Conclusions

The basic hallmark of this study has emphatically validated that hypovitaminosis D could worsen the progression of each component of the metabolic syndrome. Further studies are required to elucidate the underlying mechanism and molecular pathways of vitamin D deficiency in the progression of metabolic syndrome. Eventually, healthcare providers should supplement vitamin D to minimize the variety of elements of metabolic syndrome.

Conflicts of Interest

We declare no conflict of interest.

Funding

Thanks to King Abdulaziz City for Science and Technology for its financial support for the project of the research number (IT-1979-38).

Ethics Approval

King Abdulaziz University (KAU), Jeddah, Saudi Arabia, the KAU Research Ethical Committee granted the protocol of this animal study.

Authors' Contribution

S.K. Mahjoub: formal analysis, experimental work, investigation, M.A.A. Sattar Ahmad: conceptualization, data curation, supervision, and methodology F.O. Kamel: visualization, original draft preparation, M. Alseini: methodology and statistical analysis, L.M. Khan: interpretation of data for the work, writing review and editing, validation, and software. All authors have read and agreed to the published version of the manuscript.

References

- 1) Kato S. The function of vitamin D receptor in vitamin D action. *J Biochem* 2004; 127: 717-722.
- 2) Mandarino NR, Júnior P, Magalhães CS, Filho NS. Is vitamin D deficiency a new risk factor for cardiovascular disease? *Arterioscler Thromb Vasc Biol* 2015; 35: 198-209.
- 3) Levin A, Li X, Shumway-Cook A. Vitamin D and its analogs: do they protect against cardiovascular disease in patients with kidney disease? *Am J Hypertens* 2005; 18: 1973-1983.
- 4) Fan J, Wong SL, Lau CW, Lee HK, Ng CF, Zhang Y, Yao X, Chen ZY, Vanhoutte PM, Huang Y. Calcitriol protects renovascular function in hypertensive mice by down-regulating angiotensin II type 1 receptor and reducing oxidative stress. *Eur Heart J* 2012; 33: 2987-2990.
- 5) Wang H, Li W, Li D, Yin X, Zhang X, Olsen N, Zheng SC. Vitamin D and chronic diseases. *Aging* 2017; 8: 346-353.
- 6) López-Carretero JL, Alvarez-Blasco F, Villafrauela JJ, Balsa JA, Vázquez C, Escobar-Morreale HF. Vitamin D deficiency is associated with the metabolic syndrome in morbid obesity. *Clin Nutr* 2007; 26: 573-580.
- 7) Pereira-Santos M, Costa PR, Assis AM, Santos CA, Santos DB. Obesity and vitamin D deficiency: a systematic review and meta-analysis. *Obes Rev* 2015; 16: 341-349.
- 8) Park JE, Pichiah PBT, Cha Y-S. Vitamin D and metabolic diseases: growing roles of vitamin D. *J Obes Metab Syndr* 2018; 27: 223-232.
- 9) Dai S, McNeill JH. Fructose-induced hypertension in rats is concentration- and duration-dependent. *J Pharmacol Toxicol Methods* 1995; 33: 101-107.
- 10) Stavenuiter AW, Arcidiacono MV, Ferruzzi E, Keuning ED, Vila Cuenca M, Ter Weert Beelen RH, Vervloet MG, Dusso AS. A novel model of vitamin D deficiency: safe and rapid induction of vitamin D and calcitriol deficiency without hyperparathyroidism. *Biomed Res Int* 2015; 2015: 191234.
- 11) Patel J, Iyer A, Brown L. Evaluation of the chronic complications of diabetes on a high-fructose diet in rats. *Indian J Biochem Biophys* 2009; 46: 66-73.
- 12) Chang KC, Lian J, Tseng YC, Wu ET, Chen KL, Wu MS, Lin Y, Tseng YZ. Andrographolide prevents fructose-induced deterioration of ventricular-arterial coupling in Wistar rats. *J Pharmacol* 2007; 151: 341-347.
- 13) Thakur R, Arora G. Metabolic Syndrome: Definition, Pathophysiology, and discussion goes on. *Phys Pharm Adv* 2013; 3: 48-56.
- 14) Daugherty A, Frieri D, Hong L, Balakrishnan A. Measuring blood pressure in mice using volume pressure recording, a tail-cuff method. *J Vis Exp* 2007; 27: 1291.
- 15) Wu Y, Li XC. A simple versatile method for measuring tail cuff systolic blood pressure in conscious rats. *Clin Sci* 1997; 93: 191-194.
- 16) News DR, Hosker J, Rudenski A, Naylor B, Treacher D, Turner R. Homeostasis model assessment: insulin resistance and β -cell function from fasting plasma glucose and insulin concentrations in man. *Diabetologia* 1985; 28: 412-419.
- 17) Singh R. Antiobesity activity of aqueous and ethanol extracts of *Enicostemma littorale* in high-fat diet-induced obese rats. *Int J Phytomedicine* 2014; 6: 433-443.
- 18) Garg A, Singh R. Anti-obesity activity of ethanolic extract of *cassia auriculata* in high-fat diet-induced obese rats. *Int J Pharm Sci* 2015; 7: 237-243.
- 19) Makariou S, Liberopoulos EN, Elisaf M, Challa A. Novel roles of vitamin D in disease: what is new in 2011? *Eur J Intern Med* 2011; 22: 355-362.
- 20) Lavie CJ, Lee JH, Milani RV. Vitamin D and cardiovascular disease: will it live up to its hype? *J Am Coll Cardiol* 2011; 58: 1547-1556.
- 21) Alberti K, Eckel RH, Grundy SM, Zimmet PZ, Cleeman JI, Donato KA, Fruchart J-C, James WPT, Loria CM, Smith Jr SC. Harmonizing the metabolic syndrome: a joint interim statement of the international diabetes federation task force on epidemiology and prevention; national heart, lung, and blood institute; American heart association; world heart federation; international atherosclerosis society; and international association for the study of obesity. *Circulation* 2009; 120: 1640-1645.
- 22) Karpe F, Dickmann JR, Frayn KN. Fatty acids, obesity, and insulin resistance: time for a re-evaluation. *Diabetes* 2011; 60: 2441-2449.

- 23) Wimalawansa SJ. Associations of vitamin D with insulin resistance, obesity, type 2 diabetes, and metabolic syndrome. *J Steroid Biochem Mol Biol* 2018; 175: 177-189.
- 24) Palomer X, González-Clemente J, Blanco-Vaca F, Mauricio D. Role of vitamin D in the pathogenesis of type 2 diabetes mellitus. *Diabetes Obes Metab* 2008; 10: 185-197.
- 25) Mitri J, Pittas AG. Vitamin D and diabetes. *Endocrinol Metab Clin* 2014; 43: 205-232.
- 26) Sergeev IN. 1, 25-Dihydroxyvitamin D3 and type 2 diabetes: Ca²⁺-dependent molecular mechanisms and the role of vitamin D status. *Horm Mol Biol Clin Inv* 2016; 26: 61-65.
- 27) Kadowaki S, Norman AW. Pancreatic vitamin D-dependent calcium-binding protein: biochemical properties and response to vitamin D. *Arch Biochem Biophys* 1984; 233: 228-236.
- 28) Sooy K, Schermerhorn T, Noda M, Surana M, Rhoten WB, Meyer M, Fleischer N, Sharp GW, Christakos S. Calbindin-D28k Controls [Ca²⁺] I and Insulin Release: Evidence Obtained From Calbindin-D28k Knockout Mice And B Cell Lines. *J Biol Chem* 1999; 274: 34343-34349.
- 29) Weishaar RE, Simpson RU. Vitamin D3 and cardiovascular function in rats. *J Clin Investig* 1987; 79: 1706-1712.
- 30) Pittas AG, Nelson J, Mitri J, Hillmann W, Garber AL, C, Nathan DM, Hu FB, Dawson-Hughes B, Borch-Johnsen K, et al. 25-hydroxyvitamin D, and progression to diabetes in patients at risk for diabetes: an ancillary analysis in the Diabetes Prevention Program. *Diabetes Care* 2012; 35: 565-573.
- 31) Forouhi NG, Luan Ja, Cook DG, Sattar N, Wareham NJ. Baseline serum 25-hydroxyvitamin D is predictive of future metabolic status and insulin resistance: the Medical Research Council Prospective Study of 1990-2000. *Diabetes* 2008; 57: 2619-2625.
- 32) Gradinaru D, Ionescu C, Cioba D, Prada GI, Japaneau R. Vitamin D status and oxidative stress markers in the elderly with impaired fasting glucose and type 2 diabetes mellitus. *Aging Clin Exp Res* 2012; 24: 595-600.
- 33) Fumelle G, Guardado Mendoza R, Winnier D, Brentino M, Pengou Z, Cornell J, Andreozzi F, Jenkins JC, Cersosimo E, Federici M. The inflammatory status score including IL-6, TNF- α , osteocalcin, frataktine, MCP-1, and adiponectin underlies whole body insulin resistance and hyperglycemia in type 2 diabetes mellitus. *Acta Diabetol* 2011; 48: 123-131.
- 34) Rodenburg VM, Vervloet MG, Marx N. The role of vitamin D in cardiovascular disease: from present evidence to future perspectives. *Atherosclerosis* 2012; 225: 253-263.
- 35) Osadnik K, Osadnik T, Delijewski M, Lejawa M, Fronczek M, Reguła R, Gaşior M, Pawlas N. Calcium and phosphate levels are among other factors associated with metabolic syndrome in patients with normal weight. *Diabetes Metab Syndr* 2020; 13: 1281-1288.
- 36) Schedl HP, Heath H, Hwenger A. Serum calcitonin and parathyroid hormone in experimental diabetes: effects of insulin treatment. *Endocrinology* 1978; 103: 1368-1373.
- 37) White JR Jr, Campbell RK. Magnesium and diabetes: a review. *Ann Pharmacother* 1993; 27: 777-783.
- 38) Liamis G, Liberopoulos E, Barkas F, Elisaf M. Diabetes mellitus and electrolyte disorders. *World J Clin Cases* 2014; 2: 488-496.
- 39) Liamis G, Rodenburg EM, Hofman A, Zitterman R, Stricker BH, Hoorn EJ. Electrolyte disorders in community subjects: prevalence and risk factors. *Am J Med* 2013; 126: 255-263.
- 40) Argacha J-F, Egrise D, Fardet S, Fougère D, Lefort A, Libert F, Goldmann S, Vignery A, Burnier B, Lombard G, Morencoyes R. Vitamin D deficiency-induced hypertension is associated with vascular oxidative stress and altered heart geometry. *J Cardiovasc Pharmacol* 2011; 58: 65-71.
- 41) Muscogiuri G, Annunzio G, Duval G, Karras S, Tirabassi C, Salvio G, Biondi A, Kimball S, Kotsa M, et al. Vitamin D and cardiovascular disease: From atherosclerosis to myocardial infarction and stroke. *Int J Cardiol* 2017; 230: 577-584.
- 42) Bare M, Emmert-SJ, Coleman HA, Skordilis C, Jones DW, Mody R, Parkington HC. Vitamin D deficiency associated with impaired vascular endothelial and smooth muscle function and hypertension in young rats. *J Physiol* 2011; 589: 4777-4786.
- 43) Grossini E, Uberti F, Grossini E, Vacca G, Carda S, Invernizzi M, Cisari C. 1 α , 25-dihydroxycholecalciferol induces nitric oxide production in cultured endothelial cells. *Cell Physiol Biochem* 2011; 27: 661-668.
- 44) Wong MS, Delansorne R, Man RY, Vanhoutte PM. Vitamin D derivatives acutely reduce endothelium-dependent contractions in the aorta of the spontaneously hypertensive rat. *Am J Physiol Heart Circ Physiol* 2008; 295: 289-296.
- 45) Kassi E, Adamopoulos C, Basdra EK, Papavassiliou AG. Role of vitamin D in atherosclerosis. *Circulation* 2013; 128: 2517-2531.
- 46) Grundmann M, Haidar M, Placzko S, Niendorf R, Darashchonak N, Hubel C, Avon Versen-Höyneck F. Vitamin D improves the angiogenic properties of endothelial progenitor cells. *Am J Physiol Cell Physiol* 2012; 303: 954-962.
- 47) Kamycheva E, Jorde R, Figenschau Y, Haug E. Insulin sensitivity in subjects with secondary hyperparathyroidism and the effect of a low serum 25-hydroxyvitamin D level on insulin sensitivity. *J Endocrinol. Investig* 2007; 30: 126-132.
- 48) Judd SE, Tangpricha V. Vitamin D deficiency and risk for cardiovascular disease. *Am J Med Sci* 2009; 338: 40-44.
- 49) Cohen-Lahav M, Shany S, Tobvin D, Chaimovitz C, Douvdevani A. Vitamin D decreases NF κ B activity by increasing I κ B α levels. *Nephrol Dial Transplant* 2006; 21: 889-897.
- 50) Labudzynski D, Zaitseva O, Latyshko N, Gudkova O, Veliky M. Vitamin D3 contribution to the

- regulation of oxidative metabolism in the liver of diabetic mice *Ukr Biochem J T* 2015; 87: 75-90.
- 51) Ning C, Liu L, Lv G, Yang Y, Zhang Y, Yu R, Wang Y, Zhu J. Lipid metabolism and inflammation modulated by Vitamin D in the liver of diabetic rats. *Lipids Health Dis* 2015; 14: 1-9.
- 52) Cheng S, Massaro JM, Fox CS, Larson MG, Keyes MJ, McCabe EL, Robins SJ, O'Donnell CJ, Hoffmann U, Jacques PF. Adiposity, cardiometabolic risk, and vitamin D status: the Framingham Heart Study. *Diabetes* 2010; 59: 242-248.
- 53) Young KA, Engelman CD, Langefeld CD, Hairston KG, Haffner SM, Bryer-Ash M, Norris JM. Association of plasma vitamin D levels with adiposity in Hispanic and African Americans. *J Clin Endocrinol Metab* 2009; 94: 3306-3313.
- 54) Aggarwal N, Reis JP, Michos ED. Vitamin D deficiency and its implications on cardiovascular disease. *Curr Cardiovasc Risk Rep* 2010; 4: 68-75.
- 55) Guasch A, Bulló M, Rabassa A, Bonada A, Del Castillo D, Sabench F, Salas-Salvadó J. Plasma vitamin D, and parathormone are associated with obesity and atherogenic dyslipidemia: a cross-sectional study. *Cardiovasc Diabetol* 2012; 11: 1-11.
- 56) Kershaw EE, Flier JS. Adipose Tissue as an Endocrine Organ. *J Clin Endocrinol Metab* 2004; 89: 2548-2556.
- 57) Bellia A, Garcovich C, D'Adamo M, Lombardi M, Tesaro M, Donadel G, Gentile P, Lauro R, Federici M, Lauro R. Serum 25-hydroxyvitamin D levels are inversely associated with systemic inflammation in severely obese subjects. *Intern Emerg Med* 2013; 8: 33-37.
- 58) Miñambres I, Sánchez-Quevedo L, Pérez. The association between hypovitaminosis D and metabolic syndrome: current understanding. *Clin Lipidol* 2015; 10: 51-57.